

Biological Analysis Report

NE-DIST-2016-07-08-01

Florida Department of Environmental Protection
Central Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Event Description: **Nassau Reference Streams-SCI**
Request ID: **RQ-2016-06-27-22**
Customer: **NE-DIST**
Project ID: **SCIREF**

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Certified by: Jacqueline Savage, Environmental Specialist III

Date Certified: 03-OCT-2016 15:54

A handwritten signature in black ink that reads "Jackie Savage". The signature is written in a cursive, flowing style.

Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: AlgalBio, Chlorophyll/Grain Size/BOD, InvertZool and Microbiology.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: CEDAR RUN ABOVE BLUEHOLE

Collection Date/Time: 07/07/2016 13:15

Field ID: CEDAR RUN ABOVE BLUEHOLE

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1814736	SM 10200 H (mod.)	Chlorophyll-a, Corrected	0.75	I	ug/L	P308320	
		Phaeophytin-a	0.40	U	ug/L	P308320	
		Chlorophyll/Phaeophytin Ratio	1.6			P308320	
		Chlorophyll-a, Uncorrected	0.87	I	ug/L	P308320	
1814737	SM 9223 Quanti-Tray	Escherichia Coli-Quanti-Tray	5.2	Q	MPN/100 mL	P307988	
1814739	SOP-PCR01	HF183-qPCR**	1.0E+03	U	GEU/100 mL	P307127	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

SOP-PCR01: No amplification of HF183 marker.

Sample Location: CEDAR RUN ABOVE BLUEHOLE

Collection Date/Time: 07/07/2016 13:15

Field ID: CEDAR RUN ABOVE BLUEHOLE

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1814732	SOP-AB05	Dominant sample taxon**	Cyanophyceae				
1814738	SOP-IZ06	Stream Condition Index (SCI)**	64	A	Points		
		Macroinvert-FW-Qual-DipnetX20-#Taxa-Rep2**	25		# Taxa		

Ref. Method and Comment:

SOP-AB05: The dominant taxon was Plectonema wollei.

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)

Batch ID: P308320

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SM 9223 Quanti-Tray

Batch ID: P307988

Component	Result	Code	Units
Escherichia Coli-Quanti-Tray	1.0	U	MPN/100 mL

Reference Method: SOP-PCR01

Batch ID: P307127

Component	Result	Code	Units
HF183-qPCR	4.1E+03	U	GEU/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)

Batch ID: P308320

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
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Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
 Batch ID: P308320

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	114		P	84 - 116

Reference Method: SOP-PCR01
 Batch ID: P307127

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	138	75.9	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR01
 Batch ID: P307127

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1812484	HF183-qPCR	126		P	50 - 175
1812952	HF183-qPCR	116		P	50 - 175
1813484	HF183-qPCR	108	116	P/P	50 - 175
1814327	HF183-qPCR	124		P	50 - 175
1814739	HF183-qPCR	147		P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 9223 Quanti-Tray
 Batch ID: P307988

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1814462	Escherichia Coli-Quanti-Tray	5.42	Sample	P	0 - 80

Reference Method: SOP-PCR01
 Batch ID: P307127

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1813484	HF183-qPCR	7.63	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
 Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Lab Sample ID: 1814739
 Field Sample ID: CEDAR RUN ABOVE BLUEHOLE

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR01	SKETA-qPCR	139	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	114							
SM 9223 Quanti-Tray	Escherichia Coli-Quanti-Tray							5.42	
SOP-PCR01	HF183-qPCR	75.9	138	126	116	108			7.63
				116					

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, phaeophytin and ratio by spectrophotometry	1814736
SM 9223 Quanti-Tray / E31780	Escherichia coli by Colilert 18 Quanti-Tray method - non-chlorinated water systems	1814737
SOP-AB05 / E31780	Qualitative assessment of algal taxa present without counts	1814732
SOP-IZ06 / E31780	No. Taxa of FW macroinverts, qual samp, 20 dipnets.	1814738
SOP-PCR01 / E31780	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	1814739

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SM 10200 H (mod.)	07/08/2016	07/08/2016 14:15	Amelia G. Rankin	07/20/2016	Kim M. Spencer	1814736
SM 9223 Quanti-Tray	07/08/2016	07/08/2016 16:40	Brian Cumbie	07/09/2016 14:25	Brian Cumbie	1814737
SOP-AB05	07/08/2016			07/11/2016	Isaura Lorenzo-Perez	1814732
SOP-IJ06	07/08/2016	09/16/2016	Elizabeth Lovern	09/27/2016	Lisa Homann	1814738
	07/08/2016	09/16/2016	Elizabeth Lovern	09/29/2016	Lisa Homann	1814738
SOP-PCR01	07/08/2016	07/08/2016 13:49	Sarah Menz	07/13/2016	Puja Jasrotia	1814739