

Biological Analysis Report

WQAP-2022-10-27-03

Florida DEP Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Event Description: **Hurricane Ian Post Storm Monitoring - CEN ROC**
Request ID: **RQ-2022-10-24-44**
Customer: **WQAP**
Project ID: **HURR-IAN**

Send Reports to:
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3319 Maguire Blvd, Suite 232
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Date Certified: 10-NOV-2022 12:54



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Microbiology and Molecular.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: ST JOHNS RIVER AT HWY 40 NEAR ASTOR

Collection Date/Time: 10/26/2022 12:45

Field ID: 3515

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2365718	SM 10200 H (mod.)	Chlorophyll-a, Corrected	4.0	I	ug/L	P421626	
		Phaeophytin-a	1.8	U	ug/L	P421626	
		Chlorophyll-a, Uncorrected	4.1	A	ug/L	P421626	
2365719	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	13.4	Q	MPN/100 mL	P421697	
2365720	SOP-PCR-1.0	HF183-qPCR**	3.3E+03	U	TSC/100 mL	P421541	
	SOP-PCR-1.2	GULL2-qPCR**	3.6E+03	U	TSC/100 mL	P421541	
	SOP-PCR-1.3	GFD-qPCR**	3.6E+03		TSC/100 mL	P421541	
	SOP-PCR-1.4	BacR-qPCR**	3.0E+03	U	TSC/100 mL	P421541	
	SOP-PCR-1.5	DG3-qPCR**	2.6E+03	U	TSC/100 mL	P421541	

Ref. Method and Comment:

SM 10200 H (mod.): Laboratory duplicate result is 3.4 I for Chlorophyll-a Corrected and 1.8 U for Phaeophytin-a. Precision data is unavailable due to the small amount of analyte in the QC sample.

SOP-PCR-1.3: Duplicate was analyzed and result is 3.0E+03 I.

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)

Batch ID: P421626

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.82	U	ug/L
Chlorophyll-a, Uncorrected	0.60	U	ug/L
Phaeophytin-a	0.90	U	ug/L

Reference Method: SM 9223 B Quanti-Tray

Batch ID: P421697

Component	Result	Code	Units
Escherichia Coli-Quanti-Tray	1.0	U	MPN/100 mL

Reference Method: SOP-PCR-1.0

Batch ID: P421541

Component	Result	Code	Units
HF183-qPCR	3.3E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.2

Batch ID: P421541

Component	Result	Code	Units
GULL2-qPCR	3.6E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.3

Batch ID: P421541

Component	Result	Code	Units
GFD-qPCR	720	U	TSC/100 mL

Quality Assurance Report Method Blank Results

Reference Method: SOP-PCR-1.4
Batch ID: P421541

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.5
Batch ID: P421541

Component	Result	Code	Units
DG3-qPCR	2.6E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
Batch ID: P421626

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	87.6		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P421541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	78.9	78.5	P/P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P421541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GULL2-qPCR	90.4	91.9	P/P	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P421541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GFD-qPCR	99.3	84.2	P/P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P421541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	83.0	76.1	P/P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P421541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
DG3-qPCR	96.9	84.6	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P421541

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2365680	HF183-qPCR	79.5		P	50 - 175
2365681	HF183-qPCR	87.9		P	50 - 175
2365682	HF183-qPCR	85.8		P	50 - 175
2365720	HF183-qPCR	90.1	91.8	P/P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P421541

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2365680	GULL2-qPCR	91.3		P	50 - 300
2365681	GULL2-qPCR	64.6		P	50 - 300
2365682	GULL2-qPCR	78.2		P	50 - 300
2365720	GULL2-qPCR	92.6	92.7	P/P	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P421541

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2365680	GFD-qPCR	105		P	50 - 175
2365681	GFD-qPCR	113		P	50 - 175
2365682	GFD-qPCR	108		P	50 - 175
2365720	GFD-qPCR	107	108	P/P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P421541

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2365680	BacR-qPCR	94.9		P	50 - 175
2365681	BacR-qPCR	68.4		P	50 - 175
2365682	BacR-qPCR	75.1		P	50 - 175
2365720	BacR-qPCR	97.7	80.5	P/P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P421541

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2365680	DG3-qPCR	88.3		P	50 - 175
2365681	DG3-qPCR	76.6		P	50 - 175
2365682	DG3-qPCR	87.9		P	50 - 175
2365720	DG3-qPCR	91.2	94.6	P/P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 10200 H (mod.)
Batch ID: P421626

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365718	Chlorophyll-a, Uncorrected	22.5	Sample	P	0 - 25

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P421541

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365720	HF183-qPCR	1.82	Spike	P	0 - 25

Reference Method: SOP-PCR-1.2
Batch ID: P421541

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365720	GULL2-qPCR	0.0879	Spike	P	0 - 25

Reference Method: SOP-PCR-1.3
Batch ID: P421541

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365720	GFD-qPCR	0.918	Spike	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P421541

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365720	BacR-qPCR	19.4	Spike	P	0 - 25

Reference Method: SOP-PCR-1.5
Batch ID: P421541

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2365720	DG3-qPCR	3.67	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.

Quality Assurance Report Surrogates

Lab Sample ID: 2365720
Field Sample ID: 3515

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	96.8	P	50 - 300
SOP-PCR-1.0	SKETA-qPCR	93.0	P	50 - 300
SOP-PCR-1.2	SKETA-qPCR	108	P	50 - 300
SOP-PCR-1.2	SKETA-qPCR	106	P	50 - 300
SOP-PCR-1.3	SKETA-qPCR	106	P	50 - 300
SOP-PCR-1.3	SKETA-qPCR	102	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	105	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	107	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	97.3	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	98.4	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	87.6							
	Chlorophyll-a, Uncorrected							22.5	
SOP-PCR-1.0	HF183-qPCR	78.5	78.9	79.5	87.9	85.8			1.82
				90.1					
SOP-PCR-1.2	GULL2-qPCR	91.9	90.4	91.3	64.6	78.2			0.0879
				92.6					
SOP-PCR-1.3	GFD-qPCR	84.2	99.3	105	113	108			0.918
				107					
SOP-PCR-1.4	BacR-qPCR	76.1	83.0	94.9	68.4	75.1			19.4
				97.7					
SOP-PCR-1.5	DG3-qPCR	84.6	96.9	91.2	94.6	88.3			3.67
				76.6					

Reference Method Descriptions

Method	Description	Associated Samples
SM 10200 H (mod.)	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2365718
SM 9223 B Quanti-Tray	Escherichia coli by Colilert 18 Quanti-Tray method - non-chlorinated water systems	2365719
SOP-PCR-1.0	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2365720
SOP-PCR-1.2	Detection and quantification of coastal bird specific Catellicoccus marimammalium Gull2 genetic marker with quantitative polymerase chain reaction	2365720
SOP-PCR-1.3	Detection and quantification of bird specific Helicobacter GFD genetic marker with quantitative polymerase chain reaction	2365720
SOP-PCR-1.4	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2365720
SOP-PCR-1.5	Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction	2365720

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SM 10200 H (mod.)	10/27/2022	10/27/2022 14:27	Tynan A Branch	10/28/2022 10:14	Megan Higbee	2365718
SM 9223 B Quanti-Tray	10/27/2022	10/27/2022 14:52	Callie Williams	10/28/2022 12:24	Callie Williams	2365719
SOP-PCR-1.0	10/27/2022	10/27/2022 14:20	Claudia Castro	10/31/2022 09:04	Roberta Tatti	2365720
SOP-PCR-1.2	10/27/2022	10/27/2022 14:20	Claudia Castro	11/01/2022 09:17	Roberta Tatti	2365720
SOP-PCR-1.3	10/27/2022	10/27/2022 14:20	Claudia Castro	11/01/2022 12:18	DeAsia Armster	2365720
SOP-PCR-1.4	10/27/2022	10/27/2022 14:20	Claudia Castro	10/31/2022 10:16	DeAsia Armster	2365720
SOP-PCR-1.5	10/27/2022	10/27/2022 14:20	Claudia Castro	11/01/2022 08:16	Roberta Tatti	2365720