

Biological Analysis Report

NE-DIST-2017-12-20-01

Florida Department of Environmental Protection
Central Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Overflow Analyses Performed By:
Advanced Environmental Laboratories - Gainesville
6815 SW Archer Road
Gainesville, Florida 32608
DOH Accreditation E82001

Event Description: **Suwannee SCIs**
Request ID: **RQ-2017-12-18-11**
Customer: **NE-DIST**
Project ID: **SMP**

Send Reports to:
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8800 Baymeadows Way West
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Certified by: Cheryl A. Swanson, Program Administrator

Date Certified: 18-APR-2018 09:11



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, InvertZool, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: Mill Creek @ Bellamy/Alligator Rd

Collection Date/Time: 12/19/2017 09:20

Field ID: G1NE0605 12192017

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1955263	SM 10200 H (mod.)	Chlorophyll-a, Corrected	74		ug/L	P337877	
		Phaeophytin-a	0.80	U	ug/L	P337877	
		Chlorophyll-a, Uncorrected	77		ug/L	P337877	
1955265	EPA 1603	Escherichia coli-Membrane Filter	420	J	cfu/100 mL		
1955266	SOP-PCR-1.0	HF183-qPCR**	54	T	GEU/100 mL	P336939	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Ref. Method and Comment:
EPA 1603: Incubation period exceeded method limit by less than one hour

Sample Location: Mill Creek @ Bellamy/Alligator Rd

Collection Date/Time: 12/19/2017 09:20

Field ID: G1NE0605 12192017

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1955264	SOP-IZ06	Stream Condition Index (SCI)**	86	A	Points		
		Macroinvert-FW-Qual-DipnetX20-#Taxa-Rep2**	37		# Taxa		

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)

Batch ID: P337877

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SOP-PCR-1.0

Batch ID: P336939

Component	Result	Code	Units
HF183-qPCR	280	U	GEU/100 mL
HF183-qPCR	280	U	GEU/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)

Batch ID: P337877

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	95.0		P	84 - 116

Reference Method: SOP-PCR-1.0

Batch ID: P336939

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	141	104	P/P	50 - 175
HF183-qPCR	164	127	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
 Batch ID: P336939

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1954866	HF183-qPCR	97.7		P	50 - 175
1954892	HF183-qPCR	96.3		P	50 - 175
1954999	HF183-qPCR	95.2		P	50 - 175
1955000	HF183-qPCR	110	106	P/P	50 - 175
1955001	HF183-qPCR	108		P	50 - 175
1955239	HF183-qPCR	93.9		P	50 - 175
1955240	HF183-qPCR	78.6		P	50 - 175
1955241	HF183-qPCR	90.6		P	50 - 175
1955266	HF183-qPCR	86.9		P	50 - 175
1956283	HF183-qPCR	85.9		P	50 - 175
1956284	HF183-qPCR	80.4		P	50 - 175
1956285	HF183-qPCR	80.8		P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 10200 H (mod.)
 Batch ID: P337877

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1955321	Chlorophyll-a, Corrected	1.11	Sample	P	0 - 25
1955321	Chlorophyll-a, Uncorrected	1.25	Sample	P	0 - 25
1955321	Phaeophytin-a	2.57	Sample	P	0 - 25

Reference Method: SOP-PCR-1.0
 Batch ID: P336939

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1955000	HF183-qPCR	3.09	Spike	P	0 - 25
1958501	HF183-qPCR	3.82	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
 Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Lab Sample ID: 1955266
 Field Sample ID: G1NE0605 12192017

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	106	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery	MS % Recovery	Precision	MS
				LCS SMP	

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery			MS % Recovery			LCS	Precision	MS
									SMP	
SM 10200 H (mod.)	Chlorophyll-a, Corrected	95.0							1.11	
	Chlorophyll-a, Uncorrected								1.25	
	Phaeophytin-a								2.57	
SOP-PCR-1.0	HF183-qPCR	141	127	164	78.6	97.7	96.3			3.82 3.09
		104			95.2					

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
EPA 1603 / E82001	Escherichia coli by membrane filter method using modified mTEC by Advanced Environmental Gainesville Overflow Laboratories for NED	1955265
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	1955263
SOP-IZ06 / E31780	No. Taxa of FW macroinverts, qual samp, 20 dipnets.	1955264
SOP-PCR-1.0 / E31780	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	1955266
SOP-PCR-4.0 / E31780	Preparation of Samples by Filtration and held for qPCR Analysis	1955266

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
EPA 1603	12/19/2017	12/19/2017 14:22		12/20/2017 16:25	AEL - Gainesville	1955265
SM 10200 H (mod.)	12/20/2017	12/20/2017 12:13	Cathy Sherrer	01/05/2018 09:45	Jennifer Mihalic	1955263
SOP-IZ06	12/20/2017	02/01/2018 07:18	Cathy Sherrer	03/01/2018 08:16	Michael Heyn	1955264
	12/20/2017	02/01/2018 07:18	Cathy Sherrer	03/13/2018 08:16	Michael Heyn	1955264
SOP-PCR-1.0	12/20/2017	12/20/2017 13:30	Avril Wood	01/09/2018 09:55	Sarah Menz	1955266
SOP-PCR-4.0	12/20/2017	12/20/2017 13:30	Avril Wood	01/16/2018 11:31	Puja Jasrotia	1955266