

Biological Analysis Report

NE-DIST-2018-04-18-01

Florida Department of Environmental Protection
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DOH Accreditation E31780

Overflow Analyses Performed By:
ALS Environmental - Jacksonville
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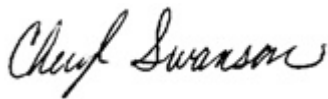
Event Description: **Suwannee North 2**
Request ID: **RQ-2018-04-16-11**
Customer: **NE-DIST**
Project ID: **SMP**

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Certified by: Cheryl A. Swanson, Program Administrator

Date Certified: 01-JUN-2018 15:49



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: Rocky Creek @ Woodpecker Road

Collection Date/Time: 04/17/2018 11:05

Field ID: 21010032

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985403	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.3	I	ug/L	P344149	
		Phaeophytin-a	0.76	I	ug/L	P344149	
		Chlorophyll-a, Uncorrected	1.8		ug/L	P344149	
1985413	SM 9223 B	Escherichia Coli-Quanti-Tray**	341		MPN/100 mL		

Sample Location: Little Creek at US 441

Collection Date/Time: 04/17/2018 11:40

Field ID: 21010033

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985402	SM 10200 H (mod.)	Chlorophyll-a, Corrected	0.55	U	ug/L	P344149	
		Phaeophytin-a	0.40	U	ug/L	P344149	
		Chlorophyll-a, Uncorrected	0.65	I	ug/L	P344149	
1985412	SM 9223 B	Escherichia Coli-Quanti-Tray**	110		MPN/100 mL		
1985417	SOP-PCR-4.0	HF183 - Human Source Marker**	Below Threshold				

Sample Location: Hunters Creek at SR 135

Collection Date/Time: 04/17/2018 09:40

Field ID: 21020063

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985404	SM 10200 H (mod.)	Chlorophyll-a, Corrected	2.5		ug/L	P344149	
		Phaeophytin-a	3.2		ug/L	P344149	
		Chlorophyll-a, Uncorrected	4.6		ug/L	P344149	
1985414	SM 9223 B	Escherichia Coli-Quanti-Tray**	613		MPN/100 mL		

Sample Location: Falling Creek @ US 441

Collection Date/Time: 04/17/2018 12:20

Field ID: 21010064

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985405	SM 10200 H (mod.)	Chlorophyll-a, Corrected	0.98	I	ug/L	P344150	
		Phaeophytin-a	0.40	U	ug/L	P344150	
		Chlorophyll-a, Uncorrected	1.2		ug/L	P344150	
1985415	SM 9223 B	Escherichia Coli-Quanti-Tray**	122		MPN/100 mL		

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Deep Creek @ HYWY 441

Collection Date/Time: 04/17/2018 12:00

Field ID: 21010051

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985418	SOP-PCR-1.4	BacR-qPCR**	1.5E+03	U	TSC/100 mL	P342991	RPD
	SOP-PCR-4.0	HF183 - Human Source Marker**	Below Threshold				

Ref. Method and Comment:

SOP-PCR-1.4: No amplification of BACR marker.

Sample Location: Deep Creek @ HYWY 441

Collection Date/Time: 04/17/2018 12:00

Field ID: 21010051

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1985406	SM 10200 H (mod.)	Chlorophyll-a, Corrected	0.55	U	ug/L	P344150	
		Phaeophytin-a	0.40	U	ug/L	P344150	
		Chlorophyll-a, Uncorrected	0.40	U	ug/L	P344150	
1985416	SM 9223 B	Escherichia Coli-Quanti-Tray**	148		MPN/100 mL		

Ref. Method and Comment:
 SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)
 Batch ID: P344149

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SM 10200 H (mod.)
 Batch ID: P344150

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SOP-PCR-1.4
 Batch ID: P342991

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
 Batch ID: P344149

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	93.6		P	84 - 116

Reference Method: SM 10200 H (mod.)
 Batch ID: P344150

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	88.7		P	84 - 116

Reference Method: SOP-PCR-1.4
 Batch ID: P342991

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	109	61.8	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.4
Batch ID: P342991

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1974132	BacR-qPCR	88.3		P	50 - 175
1983204	BacR-qPCR	101		P	50 - 175
1983223	BacR-qPCR	79.1	61.3	P/P	50 - 175
1985418	BacR-qPCR	80.9		P	50 - 175
1985611	BacR-qPCR	82.2		P	50 - 175
1985612	BacR-qPCR	72.5		P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 10200 H (mod.)
Batch ID: P344149

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1985317	Chlorophyll-a, Corrected	8.19	Sample	P	0 - 25
1985317	Chlorophyll-a, Uncorrected	7.94	Sample	P	0 - 25
1985317	Phaeophytin-a	5.75	Sample	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P342991

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1983223	BacR-qPCR	25.3	Spike	F	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Lab Sample ID: 1985418
Field Sample ID: 21010051

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.4	SKETA-qPCR	99.4	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			Precision
				LCS	SMP	MS	
SM 10200 H (mod.)	Chlorophyll-a, Corrected	93.6			8.19		
	Chlorophyll-a, Corrected	88.7					
	Chlorophyll-a, Uncorrected				7.94		
	Phaeophytin-a				5.75		
SOP-PCR-1.4	BacR-qPCR	61.8	109	88.3	101	79.1	25.3
				61.3			

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	1985402, 1985403, 1985404, 1985405, 1985406
SM 9223 B / E82502	Escherichia coli by Colilert Quanti-Tray method by ALS Environmental Overflow Laboratory for Northeast District	1985412, 1985413, 1985414, 1985415, 1985416
SOP-PCR-1.4 / E31780	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	1985418
SOP-PCR-4.0 / E31780	Preparation of Samples by Filtration and held for qPCR Analysis	1985417, 1985418

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SM 10200 H (mod.)	04/18/2018	04/18/2018 14:00	Mariam Hanna	04/24/2018 11:45	Carina A. Tull	1985402, 1985403, 1985404
	04/18/2018	04/18/2018 14:00	Mariam Hanna	04/25/2018 12:25	Jennifer Mihalic	1985405, 1985406
SM 9223 B	04/17/2018	04/17/2018 17:25		04/18/2018 20:44	ALS Environmental, Jax	1985412, 1985413, 1985414, 1985415, 1985416
SOP-PCR-1.4	04/18/2018	04/18/2018 12:45	Sarah Menz	04/20/2018 14:30	Sarah Menz	1985418
SOP-PCR-4.0	04/18/2018	04/18/2018 12:45	Angela Smith	05/07/2018 15:05	Puja Jasrotia	1985417, 1985418