

Biological Analysis Report

NE-DIST-2018-08-23-01

Florida Department of Environmental Protection
Central Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Overflow Analyses Performed By:
ALS Environmental - Jacksonville
9143 Philips Highway, Suite 200
Jacksonville, FL 32256
DOH Accreditation E82502

Event Description: **LSJR Southside and East**
Request ID: **RQ-2018-08-20-18**
Customer: **NE-DIST**
Project ID: **SMP**

Send Reports to:
FL Dept. of Environmental Protection
8800 Baymeadows Way West
Suite 100
Jacksonville, FL 32256-7577
Attn: Jeremy Parrish

For additional information please contact
Cheryl Swanson - Administrator
J. Marshall Faircloth - Aquatic Toxicology
Jacqueline Savage - Bench Biology
Jacqueline Savage - Microbiology
Puja Jasrotia, Ph.D. - Molecular Biology
Amelia Rankin - Taxonomy (Invertebrate & Algal)
Thekkekalathil Chandrasekhar, Ph.D, QA Officer
Phone (850) 245-8177

Certified by: Cheryl A. Swanson, Program Administrator

Date Certified: 20-SEP-2018 20:39



NON-CONFORMANCE REPORT INCLUDED

Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: NEWCASTLE CR @ FT CAROLINE HILLS RD. Collection Date/Time: 08/22/2018 09:20

Field ID: 20030872

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019419	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	1230		MPN/100 mL		

Sample Location: MILLER CR @ SCHUMACHER Collection Date/Time: 08/22/2018 10:15

Field ID: 20030794

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019420	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	15500		MPN/100 mL		
2019428	SOP-PCR-1.0	HF183-qPCR**	190	U	GEU/100 mL	P350953	
	SOP-PCR-1.2	GULL2-qPCR**	1.1E+04	U	TSC/100 mL	P350953	
	SOP-PCR-1.3	GFD-purified-qPCR**	110	U	TSC/100 mL	P351074	LCS, MS
	SOP-PCR-1.4	BacR-qPCR**	2.0E+03	U	TSC/100 mL	P350953	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Ref. Method and Comment:
 SOP-PCR-1.2: Precision data unavailable for Analysis Batch A86375

Sample Location: MILLER CR @ BEACH BLVD Collection Date/Time: 08/22/2018 09:55

Field ID: 20030828

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019413	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.3	I	ug/L	P351471	
		Phaeophytin-a	0.40	U	ug/L	P351471	
		Chlorophyll-a, Uncorrected	1.2	I	ug/L	P351471	
2019416	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	7700		MPN/100 mL		
2019427	SOP-PCR-1.0	HF183-qPCR**	300	I	GEU/100 mL	P350953	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Sample Location: MIRAMAR CR @ GADSEN RD Collection Date/Time: 08/22/2018 10:35

Field ID: 20030825

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019421	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	10	U	MPN/100 mL		
2019429	SOP-PCR-4.0	HF183 - Human Source Marker**	Below Threshold				

Sample Location: MIRAMAR AT SAN JOSE Collection Date/Time: 08/22/2018 10:45

Field ID: G2NE1164

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019422	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	605		MPN/100 mL		
2019430	SOP-PCR-1.0	HF183-qPCR**	770		GEU/100 mL	P350953	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Sample Location: OLDFIELD CR @ JULINGTON CREEK ROAD Collection Date/Time: 08/22/2018 11:30

Field ID: 20030684

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019414	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.1	I	ug/L	P351471	

Field ID: 20030684

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019414	SM 10200 H (mod.)	Phaeophytin-a	0.40	U	ug/L	P351471	
		Chlorophyll-a, Uncorrected	1.2		ug/L	P351471	
2019417	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	1660		MPN/100 mL		

Sample Location: OLDFIELD CR @ LORETTO RD

Collection Date/Time: 08/22/2018 12:05

Field ID: 20030563

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2019415	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.8	I	ug/L	P351471	
		Phaeophytin-a	0.64	I	ug/L	P351471	
		Chlorophyll-a, Uncorrected	2.2		ug/L	P351471	
2019418	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	341		MPN/100 mL		

Non-Conformance Report

NCR ID: 7188

Event(s)

NE-DIST-2018-08-23-01

Job(s)

Sample(s)

Test(s)

NCR Type: SHIPPING/RECEIVING

NCR Category: Receiving

Observation: PCR-FILTER container for NEWCASTLE CR @ FT CAROLINE HILLS RD container was received empty.

Resolution: Jeremy Parrish was notified; email added to file.

Authorized by/Date: Joshua Ayres 9/4/2018

The Non-Conformance Report details exceptions or problems encountered with the events/jobs/samples/test.
Please address questions to:

Chemistry Colin Wright (850) 245-8085
Biology Cheryl Swanson (850) 245-8177

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)

Batch ID: P351471

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SOP-PCR-1.0

Batch ID: P350953

Component	Result	Code	Units
HF183-qPCR	280	U	GEU/100 mL

Reference Method: SOP-PCR-1.2

Batch ID: P350953

Component	Result	Code	Units
GULL2-qPCR	1.7E+04	U	TSC/100 mL

Quality Assurance Report Method Blank Results

Reference Method: SOP-PCR-1.3
Batch ID: P351074

Component	Result	Code	Units
GFD-purified-qPCR	340	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P350953

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
Batch ID: P351471

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	85.9		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P350953

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	115	129	P/P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P350953

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GULL2-qPCR	103	99.2	P/P	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P351074

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GFD-purified-qPCR	121	49.7	P/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P350953

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	91.4	162	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P350953

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2016142	HF183-qPCR	93.8		P	50 - 175
2016293	HF183-qPCR	102		P	50 - 175
2017670	HF183-qPCR	95.6		P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0

Batch ID: P350953

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2017671	HF183-qPCR	85.5		P	50 - 175
2017672	HF183-qPCR	96.8		P	50 - 175
2019427	HF183-qPCR	94.7		P	50 - 175
2019428	HF183-qPCR	98.1		P	50 - 175
2019430	HF183-qPCR	86.6		P	50 - 175
2020083	HF183-qPCR	87.2		P	50 - 175
2020084	HF183-qPCR	92.8		P	50 - 175
2020736	HF183-qPCR	94.8		P	50 - 175
2022097	HF183-qPCR	76.5	85.6	P/P	50 - 175
2022098	HF183-qPCR	84.9		P	50 - 175

Reference Method: SOP-PCR-1.2

Batch ID: P350953

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2016293	GULL2-qPCR	82.2		P	50 - 300
2017670	GULL2-qPCR	90.7		P	50 - 300
2017671	GULL2-qPCR	79.8		P	50 - 300
2019428	GULL2-qPCR	96.0		P	50 - 300
2020083	GULL2-qPCR	76.4		P	50 - 300
2020084	GULL2-qPCR	80.8		P	50 - 300

Reference Method: SOP-PCR-1.3

Batch ID: P351074

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2013669	GFD-purified-qPCR	157		P	50 - 175
2015786	GFD-purified-qPCR	172		P	50 - 175
2016247	GFD-purified-qPCR	50.0		F	50 - 175
2016293	GFD-purified-qPCR	138		P	50 - 175
2017669	GFD-purified-qPCR	177		F	50 - 175
2017670	GFD-purified-qPCR	130		P	50 - 175
2017671	GFD-purified-qPCR	143		P	50 - 175
2018597	GFD-purified-qPCR	172		P	50 - 175
2018598	GFD-purified-qPCR	133		P	50 - 175
2019428	GFD-purified-qPCR	153		P	50 - 175
2023260	GFD-purified-qPCR	153		P	50 - 175
2023261	GFD-purified-qPCR	172	150	P/P	50 - 175
2023262	GFD-purified-qPCR	95.4		P	50 - 175
2023263	GFD-purified-qPCR	44.8		F	50 - 175
2023264	GFD-purified-qPCR	66.9		P	50 - 175
2023265	GFD-purified-qPCR	106		P	50 - 175
2023266	GFD-purified-qPCR	131		P	50 - 175
2023267	GFD-purified-qPCR	160		P	50 - 175

Reference Method: SOP-PCR-1.4

Batch ID: P350953

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2016293	BacR-qPCR	68.7		P	50 - 175
2017670	BacR-qPCR	68.7		P	50 - 175
2017671	BacR-qPCR	62.0		P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.4
Batch ID: P350953

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2019428	BacR-qPCR	72.0		P	50 - 175
2020083	BacR-qPCR	61.9		P	50 - 175
2020084	BacR-qPCR	54.3		P	50 - 175
2020736	BacR-qPCR	62.5		P	50 - 175
2022097	BacR-qPCR	62.1	64.0	P/P	50 - 175
2022098	BacR-qPCR	61.7		P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 10200 H (mod.)
Batch ID: P351471

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2019601	Chlorophyll-a, Corrected	8.19	Sample	P	0 - 25
2019601	Chlorophyll-a, Uncorrected	6.81	Sample	P	0 - 25
2019601	Phaeophytin-a	4.50	Sample	P	0 - 25

Reference Method: SOP-PCR-1.0
Batch ID: P350953

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2022097	HF183-qPCR	11.3	Spike	P	0 - 25

Reference Method: SOP-PCR-1.3
Batch ID: P351074

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2023261	GFD-purified-qPCR	13.4	Spike	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P350953

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2022097	BacR-qPCR	2.98	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Lab Sample ID: 2019427
Field Sample ID: 20030828

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	107	P	50 - 300

Quality Assurance Report Surrogates

Lab Sample ID: 2019428
Field Sample ID: 20030794

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	112	P	50 - 300
SOP-PCR-1.2	SKETA-qPCR	112	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	104	P	50 - 300

Lab Sample ID: 2019430
Field Sample ID: G2NE1164

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	95.9	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	85.9						8.19	
	Chlorophyll-a, Uncorrected							6.81	
	Phaeophytin-a							4.50	
SOP-PCR-1.0	HF183-qPCR	129	115	98.1	86.6	87.2			11.3
				92.8					
SOP-PCR-1.2	GULL2-qPCR	99.2	103	90.7	79.8	82.2			
				96.0					
SOP-PCR-1.3	GFD-purified-qPCR	49.7	121	172	177	157			13.4
				50.0					
SOP-PCR-1.4	BacR-qPCR	162	91.4	68.7	62.0	68.7			2.98
				62.1					

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2019413, 2019414, 2019415
SM 9223 B Quanti-Tray / E82502	Escherichia coli by Colilert Quanti-Tray method by ALS Environmental Laboratory in Jacksonville	2019416, 2019417, 2019418, 2019419, 2019420, 2019421, 2019422
SOP-PCR-1.0 / E31780	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2019427, 2019428, 2019430
SOP-PCR-1.2 / E31780	Detection and quantification of coastal bird specific Catellicoccus marimammalium Gull2 genetic marker with quantitative polymerase chain reaction	2019428
SOP-PCR-1.3 / E31780	Detection and quantification of bird specific Helicobacter GFD genetic marker with quantitative polymerase chain reaction	2019428
SOP-PCR-1.4 / E31780	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2019428
SOP-PCR-4.0 / E31780	Preparation of Samples by Filtration and held for qPCR Analysis	2019427, 2019428, 2019429, 2019430

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SM 10200 H (mod.)	08/23/2018	08/23/2018 13:25	Edwin Gonzalez-Soto	09/05/2018 08:15	Carina A. Tull	2019413, 2019414, 2019415
SM 9223 B Quanti-Tray	08/22/2018	08/22/2018 16:08		08/23/2018 16:15	ALS Environmental, Jax	2019416, 2019417, 2019418, 2019419, 2019420, 2019421, 2019422
SOP-PCR-1.0	08/23/2018	08/23/2018 15:45	Sarah Menz	09/07/2018 11:00	Mailin Lopez	2019427, 2019428, 2019430
SOP-PCR-1.2	08/23/2018	08/23/2018 15:45	Sarah Menz	09/10/2018 13:07	Mailin Lopez	2019428
SOP-PCR-1.3	08/23/2018	08/23/2018 15:45	Sarah Menz	09/13/2018 15:38	Sarah Menz	2019428
SOP-PCR-1.4	08/23/2018	08/23/2018 15:45	Sarah Menz	09/07/2018 15:36	Mailin Lopez	2019428
SOP-PCR-4.0	08/23/2018	08/23/2018 15:45	Angela Smith	09/06/2018 08:42	Sarah Menz	2019427, 2019428, 2019429, 2019430