

Biological Analysis Report

SW-DIST-2018-04-12-01

Florida Department of Environmental Protection
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Overflow Analyses Performed By:
Advanced Environmental Laboratories - Tampa
9610 Princess Palm Avenue
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DOH Accreditation E84589

Event Description: **Spartman, Flint, Bellows, Baker**
Request ID: **RQ-2018-04-09-07**
Customer: **SW-DIST**
Project ID: **SMP**

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Date Certified: 24-MAY-2018 16:33



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: BAKER CREEK @ THONOTOSASSA RD

Collection Date/Time: 04/11/2018 09:45

Field ID: G2SW0012

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983994	EPA 1603	Escherichia coli-Membrane Filter**	2500		cfu/100 mL		

Sample Location: BAKER CREEK @ THONOTOSASSA RD

Collection Date/Time: 04/11/2018 09:45

Field ID: G2SW0012

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983989	SOP-PCR-1.0	HF183-qPCR**	100	T	GEU/100 mL	P345261	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Sample Location: BELLOWS LAKE OUTLET DUP

Collection Date/Time: 04/11/2018 11:45

Field ID: G1SW0060_DUP

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983967	SM 10200 H (mod.)	Chlorophyll-a, Corrected	25		ug/L	P344004	
		Phaeophytin-a	7.7		ug/L	P344004	
		Chlorophyll-a, Uncorrected	30		ug/L	P344004	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: BELLOWS LAKE OUTLET DUP

Collection Date/Time: 04/11/2018 11:45

Field ID: G1SW0060_DUP

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983995	EPA 1603	Escherichia coli-Membrane Filter**	8000	Z	cfu/100 mL		

Sample Location: Field Blank

Collection Date/Time: 04/11/2018 12:20

Field ID: BLANK

Matrix: W-FIELD-BK

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983968	SM 10200 H (mod.)	Chlorophyll-a, Corrected	0.55	U	ug/L	P344004	
		Phaeophytin-a	0.40	U	ug/L	P344004	
		Chlorophyll-a, Uncorrected	0.40	U	ug/L	P344004	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Field Blank

Collection Date/Time: 04/11/2018 12:20

Field ID: BLANK

Matrix: W-FIELD-BK

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983996	EPA 1603	Escherichia coli-Membrane Filter**	1	U	cfu/100 mL		

Sample Location: SPARTMAN BRANCH AT HARVEY TEW RD

Collection Date/Time: 04/11/2018 10:25

Field ID: G2SW0067

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983997	EPA 1603	Escherichia coli-Membrane Filter**	1600	B	cfu/100 mL		

Sample Location: SPARTMAN BRANCH AT HARVEY TEW RD

Collection Date/Time: 04/11/2018 10:25

Field ID: G2SW0067

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983990	SOP-PCR-1.0	HF183-qPCR**	58	T	GEU/100 mL	P345261	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Sample Location: BELLOWS LAKE OUTLET

Collection Date/Time: 04/11/2018 11:40

Field ID: G1SW0060

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983969	SM 10200 H (mod.)	Chlorophyll-a, Corrected	33		ug/L	P344004	
		Phaeophytin-a	8.2		ug/L	P344004	
		Chlorophyll-a, Uncorrected	39		ug/L	P344004	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: BELLOWS LAKE OUTLET

Collection Date/Time: 04/11/2018 11:40

Field ID: G1SW0060

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983998	EPA 1603	Escherichia coli-Membrane Filter**	8000	Z	cfu/100 mL		

Sample Location: FLINT CREEK @ KNIGHTS GRIFFIN RD

Collection Date/Time: 04/11/2018 10:45

Field ID: G2SW0066

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983999	EPA 1603	Escherichia coli-Membrane Filter**	4000		cfu/100 mL		

Sample Location: FLINT CREEK @ KNIGHTS GRIFFIN RD

Collection Date/Time: 04/11/2018 10:45

Field ID: G2SW0066

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983991	SOP-PCR-1.0	HF183-qPCR**	11	T	GEU/100 mL	P345261	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Sample Location: FLINT CREEK @ SR301, TP39

Collection Date/Time: 04/11/2018 11:05

Field ID: G2SW0004

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1984000	EPA 1603	Escherichia coli-Membrane Filter**	1500	B	cfu/100 mL		

Sample Location: FLINT CREEK @ SR301, TP39

Collection Date/Time: 04/11/2018 11:05

Field ID: G2SW0004

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983992	SOP-PCR-1.0	HF183-qPCR**	280	U	GEU/100 mL	P345261	
	SOP-PCR-1.2	GULL2-qPCR**	1.7E+04	U	TSC/100 mL	P345261	LCS
	SOP-PCR-1.3	GFD-purified-qPCR**	1.34E+03		TSC/100 mL	P345032	LCS
	SOP-PCR-1.4	BacR-qPCR**	3.0E+03	U	TSC/100 mL	P345261	

Field ID: G2SW0004

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983992	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Ref. Method and Comment:
 SOP-PCR-1.0: No amplification of HF183 marker.
 SOP-PCR-1.2: No amplification of GULL2 marker. Precision data unavailable for Analysis Batch A83964.
 SOP-PCR-1.4: No amplification of BACR marker. Precision data unavailable for Analysis Batch A83966.

Sample Location: SPARTMAN BRANCH @ US 92

Collection Date/Time: 04/11/2018 10:05

Field ID: G2SW0013

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1984001	EPA 1603	Escherichia coli-Membrane Filter**	2300		cfu/100 mL		

Sample Location: SPARTMAN BRANCH @ US 92

Collection Date/Time: 04/11/2018 10:05

Field ID: G2SW0013

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1983993	SOP-PCR-1.0	HF183-qPCR**	9.2	T	GEU/100 mL	P345261	
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold				

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)

Batch ID: P344004

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.55	U	ug/L
Chlorophyll-a, Uncorrected	0.40	U	ug/L
Phaeophytin-a	0.40	U	ug/L

Reference Method: SOP-PCR-1.0

Batch ID: P345261

Component	Result	Code	Units
HF183-qPCR	280	U	GEU/100 mL

Reference Method: SOP-PCR-1.2

Batch ID: P345261

Component	Result	Code	Units
GULL2-qPCR	1.7E+04	U	TSC/100 mL

Reference Method: SOP-PCR-1.3

Batch ID: P345032

Component	Result	Code	Units
GFD-purified-qPCR	340	U	TSC/100 mL

Reference Method: SOP-PCR-1.4

Batch ID: P345261

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
Batch ID: P344004

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	86.4		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P345261

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	55.0	95.7	P/P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P345261

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GULL2-qPCR	92.0	46.8	P/F	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P345032

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GFD-purified-qPCR	102	30.8	P/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P345261

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	92.3	74.4	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P345261

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1981703	HF183-qPCR	85.7		P	50 - 175
1981704	HF183-qPCR	87.3		P	50 - 175
1983989	HF183-qPCR	81.6		P	50 - 175
1983990	HF183-qPCR	92.1	86.0	P/P	50 - 175
1983991	HF183-qPCR	82.5		P	50 - 175
1983992	HF183-qPCR	80.2		P	50 - 175
1983993	HF183-qPCR	89.6		P	50 - 175
1985974	HF183-qPCR	86.5		P	50 - 175
1985975	HF183-qPCR	105		P	50 - 175
1986052	HF183-qPCR	84.1		P	50 - 175
1986054	HF183-qPCR	82.1		P	50 - 175
1986378	HF183-qPCR	70.9		P	50 - 175
1986379	HF183-qPCR	75.1		P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P345261

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
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Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.2
Batch ID: P345261

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1983992	GULL2-qPCR	70.6		P	50 - 300
1985974	GULL2-qPCR	72.5		P	50 - 300
1986052	GULL2-qPCR	85.2		P	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P345032

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1983992	GFD-purified-qPCR	51.1		P	50 - 175
1985936	GFD-purified-qPCR	95.2		P	50 - 175
1985938	GFD-purified-qPCR	107		P	50 - 175
1985974	GFD-purified-qPCR	142		P	50 - 175
1986052	GFD-purified-qPCR	97.3		P	50 - 175
1986379	GFD-purified-qPCR	125		P	50 - 175
1991711	GFD-purified-qPCR	89.1		P	50 - 175
1991712	GFD-purified-qPCR	147	160	P/P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P345261

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1981703	BacR-qPCR	64.8		P	50 - 175
1983992	BacR-qPCR	60.0		P	50 - 175
1985974	BacR-qPCR	63.6		P	50 - 175
1986052	BacR-qPCR	73.8		P	50 - 175
1986054	BacR-qPCR	50.3		P	50 - 175

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P345261

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1983990	HF183-qPCR	6.83	Spike	P	0 - 25

Reference Method: SOP-PCR-1.3
Batch ID: P345032

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1991712	GFD-purified-qPCR	8.56	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Quality Assurance Report Surrogates

Lab Sample ID: 1983989
Field Sample ID: G2SW0012

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	102	P	50 - 300

Lab Sample ID: 1983990
Field Sample ID: G2SW0067

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	100	P	50 - 300

Lab Sample ID: 1983991
Field Sample ID: G2SW0066

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	99.5	P	50 - 300

Lab Sample ID: 1983992
Field Sample ID: G2SW0004

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	91.8	P	50 - 300
SOP-PCR-1.2	SKETA-qPCR	78.8	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	82.4	P	50 - 300

Lab Sample ID: 1983993
Field Sample ID: G2SW0013

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	105	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery	MS % Recovery	LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	86.4				
SOP-PCR-1.0	HF183-qPCR	95.7 55.0	85.7 87.3 81.6			6.83
			92.1			
SOP-PCR-1.2	GULL2-qPCR	46.8 92.0	70.6 72.5 85.2			
SOP-PCR-1.3	GFD-purified-qPCR	30.8 102	89.1 147 160			8.56
			95.2			
SOP-PCR-1.4	BacR-qPCR	74.4 92.3	64.8 60.0 63.6			
			73.8			

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
EPA 1603 / E84589	Escherichia coli by membrane filter method using modified mTEC by Advanced Environmental Tampa Lab for SWD	1983994, 1983995, 1983996, 1983997, 1983998, 1983999, 1984000, 1984001
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	1983967, 1983968, 1983969
SOP-PCR-1.0 / E31780	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	1983989, 1983990, 1983991, 1983992, 1983993

Reference Method Descriptions

Method / DoH Cert #	Description	<u>Associated Samples</u>
SOP-PCR-1.2 / E31780	Detection and quantification of coastal bird specific <i>Catelliboccus marimammalium</i> Gull2 genetic marker with quantitative polymerase chain reaction	1983992
SOP-PCR-1.3 / E31780	Detection and quantification of bird specific <i>Helicobacter</i> GFD genetic marker with quantitative polymerase chain reaction	1983992
SOP-PCR-1.4 / E31780	Detection and quantification of Ruminant specific <i>Bacteroidetes</i> BacR genetic marker with quantitative polymerase chain reaction	1983992
SOP-PCR-4.0 / E31780	Preparation of Samples by Filtration and held for qPCR Analysis	1983989, 1983990, 1983991, 1983992, 1983993

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
EPA 1603	04/11/2018	04/11/2018 17:40		04/12/2018 16:10	AEL - Tampa	1983994, 1983995, 1983996, 1983997, 1983998, 1983999, 1984000, 1984001
SM 10200 H (mod.)	04/12/2018	04/12/2018 13:50	Peterson Janvier	04/18/2018 16:20	Carina A. Tull	1983967, 1983968, 1983969
SOP-PCR-1.0	04/12/2018	04/12/2018 14:15	Puja Jasrotia	05/15/2018 16:44	Mailin Lopez	1983989, 1983990, 1983991, 1983992, 1983993
SOP-PCR-1.2	04/12/2018	04/12/2018 14:15	Puja Jasrotia	05/16/2018 15:33	Sarah Menz	1983992
SOP-PCR-1.3	04/12/2018	04/12/2018 14:15	Angela Smith	05/17/2018 14:48	Sarah Menz	1983992
SOP-PCR-1.4	04/12/2018	04/12/2018 14:15	Puja Jasrotia	05/16/2018 12:41	Sarah Menz	1983992
SOP-PCR-4.0	04/12/2018	04/12/2018 14:15	Angela Smith	05/14/2018 15:47	Puja Jasrotia	1983989, 1983990, 1983991, 1983992, 1983993