

Biological Analysis Report

NE-DIST-2019-03-19-01

Florida Department of Environmental Protection
Central Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Overflow Analyses Performed By:
Advanced Environmental Laboratories - Gainesville
6815 SW Archer Road
Gainesville, Florida 32608
DOH Accreditation E82001

Event Description: **Alachua County March SMP**
Request ID: **RQ-2019-03-11-13**
Customer: **NE-DIST**
Project ID: **SMP**

Send Reports to:
FL Dept. of Environmental Protection
8800 Baymeadows Way West
Suite 100
Jacksonville, FL 32256
Attn: Jeremy Parrish

For additional information please contact
Cheryl Swanson - Administrator
Jacqueline Savage - Microbiology
Diane Power - Bench Biology
Andrew Tintle - Aquatic Toxicology
Puja Jasrotia, Ph.D. - Molecular Biology
Amelia Rankin - Taxonomy (Invertebrate & Algal)
Thekkekalathil Chandrasekhar, Ph.D, QA Officer
Phone (850) 245-8177

Certified by: Jacqueline Savage, Environmental Administrator

Date Certified: 07-MAY-2019 13:03



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: Pareners Branch Creek E of CR1491

Collection Date/Time: 03/18/2019 11:00

Field ID: G1NE0906 - PAR1491

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2070974	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.1	U	ug/L	P361686	
		Phaeophytin-a	1.2	U	ug/L	P361686	
		Chlorophyll-a, Uncorrected	1.4	I	ug/L	P361686	
2070977	EPA 1603	Escherichia coli-Membrane Filter	630		cfu/100 mL		
2070981	SOP-PCR-1.0	HF183-qPCR**	94	U	GEU/100 mL	P361140	
	SOP-PCR-1.4	BacR-qPCR**	4.35E+06		TSC/100 mL	P361140	MS
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold			P360637	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Pareners Branch Creek DS of CR262

Collection Date/Time: 03/18/2019 11:45

Field ID: G1NE0917 - PAR262

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2070975	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.1	U	ug/L	P361686	
		Phaeophytin-a	1.2	U	ug/L	P361686	
		Chlorophyll-a, Uncorrected	1.0	I	ug/L	P361686	
2070978	EPA 1603	Escherichia coli-Membrane Filter	820	B	cfu/100 mL		
2070982	SOP-PCR-1.0	HF183-qPCR**	140	U	GEU/100 mL	P361140	
	SOP-PCR-1.4	BacR-qPCR**	1.55E+06		TSC/100 mL	P361140	MS
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold			P360637	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Mill Creek E of Old Belamy Rd

Collection Date/Time: 03/18/2019 12:15

Field ID: G1NE0907 - MillBel

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2070976	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.2	U	ug/L	P361686	
		Phaeophytin-a	1.4	U	ug/L	P361686	
		Chlorophyll-a, Uncorrected	0.90	U	ug/L	P361686	
2070979	EPA 1603	Escherichia coli-Membrane Filter	570		cfu/100 mL		
2070983	SOP-PCR-1.0	HF183-qPCR**	94	U	GEU/100 mL	P361140	
	SOP-PCR-1.2	GULL2-qPCR**	5.6E+03	U	TSC/100 mL	P361140	
	SOP-PCR-1.3	GFD-purified-qPCR**	140	IJ	TSC/100 mL	P361911	LCS, MS, RPD
	SOP-PCR-1.4	BacR-qPCR**	1.0E+03	U	TSC/100 mL	P361140	MS
	SOP-PCR-4.0	HF183 - Human Source Marker**	Above Threshold			P360637	

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

SOP-PCR-1.2: Precision data unavailable for Analysis Batch # A90582

SOP-PCR-1.3: Quality control failure(s) observed. See QA report for details.

Quality Assurance Report

Method Blank Results

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)
Batch ID: P361686

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.62	U	ug/L
Chlorophyll-a, Uncorrected	0.45	U	ug/L
Phaeophytin-a	0.68	U	ug/L

Reference Method: SOP-PCR-1.0
Batch ID: P361140

Component	Result	Code	Units
HF183-qPCR	280	U	GEU/100 mL

Reference Method: SOP-PCR-1.2
Batch ID: P361140

Component	Result	Code	Units
GULL2-qPCR	1.7E+04	U	TSC/100 mL

Reference Method: SOP-PCR-1.3
Batch ID: P361911

Component	Result	Code	Units
GFD-purified-qPCR	340	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P361140

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
Batch ID: P361686

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	105		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P361140

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	131	110	P/P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P361140

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GULL2-qPCR	137	210	P/P	50 - 300

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SOP-PCR-1.3
Batch ID: P361911

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GFD-purified-qPCR	151	13.2	P/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P361140

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	109	121	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P361140

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2068511	HF183-qPCR	71.5		P	50 - 175
2068767	HF183-qPCR	98.2		P	50 - 175
2068768	HF183-qPCR	95.3		P	50 - 175
2070871	HF183-qPCR	96.3		P	50 - 175
2070981	HF183-qPCR	93.0		P	50 - 175
2070982	HF183-qPCR	102		P	50 - 175
2070983	HF183-qPCR	114		P	50 - 175
2071059	HF183-qPCR	96.7		P	50 - 175
2073222	HF183-qPCR	92.1		P	50 - 175
2073223	HF183-qPCR	112		P	50 - 175
2073224	HF183-qPCR	83.9		P	50 - 175
2073225	HF183-qPCR	93.8	95.7	P/P	50 - 175
2073226	HF183-qPCR	68.8		P	50 - 175

Reference Method: SOP-PCR-1.2
Batch ID: P361140

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2068767	GULL2-qPCR	106		P	50 - 300
2068768	GULL2-qPCR	81.9		P	50 - 300
2070871	GULL2-qPCR	81.0		P	50 - 300
2070983	GULL2-qPCR	82.4		P	50 - 300
2071059	GULL2-qPCR	102		P	50 - 300

Reference Method: SOP-PCR-1.3
Batch ID: P361911

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2068767	GFD-purified-qPCR	239		F	50 - 175
2068768	GFD-purified-qPCR	253		F	50 - 175
2069251	GFD-purified-qPCR	234		F	50 - 175
2069252	GFD-purified-qPCR	152		P	50 - 175
2069253	GFD-purified-qPCR	260		F	50 - 175
2069298	GFD-purified-qPCR	146		P	50 - 175
2070024	GFD-purified-qPCR	143		P	50 - 175
2070548	GFD-purified-qPCR	153		P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.3
Batch ID: P361911

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2070871	GFD-purified-qPCR	231		F	50 - 175
2070983	GFD-purified-qPCR	182		F	50 - 175
2071059	GFD-purified-qPCR	141		P	50 - 175
2071300	GFD-purified-qPCR	138		P	50 - 175
2071762	GFD-purified-qPCR	140		P	50 - 175
2072205	GFD-purified-qPCR	133		P	50 - 175
2077083	GFD-purified-qPCR	137		P	50 - 175
2077084	GFD-purified-qPCR	210		F	50 - 175
2077085	GFD-purified-qPCR	183		F	50 - 175
2077086	GFD-purified-qPCR	293	218	F/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P361140

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2068767	BacR-qPCR	102		P	50 - 175
2068768	BacR-qPCR	85.2		P	50 - 175
2070871	BacR-qPCR	86.0		P	50 - 175
2070981	BacR-qPCR	85.1		P	50 - 175
2070982	BacR-qPCR	92.6		P	50 - 175
2070983	BacR-qPCR	85.7		P	50 - 175
2071059	BacR-qPCR	88.5		P	50 - 175
2073222	BacR-qPCR	56.9		P	50 - 175
2073223	BacR-qPCR	68.9		P	50 - 175
2073224	BacR-qPCR	56.8		P	50 - 175
2073225	BacR-qPCR	69.1	61.4	P/P	50 - 175
2073226	BacR-qPCR	47.6		F	50 - 175

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P361140

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2073225	HF183-qPCR	2.00	Spike	P	0 - 25

Reference Method: SOP-PCR-1.3
Batch ID: P361911

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2077086	GFD-purified-qPCR	29.4	Spike	F	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P361140

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2073225	BacR-qPCR	11.7	Spike	P	0 - 25

Quality Assurance Report Precision

* Sample, spike and/or laboratory control sample precision (LCS) is reported.
Replicate spike precision may be reported when sample results are below quantifiable levels.

Quality Assurance Report Surrogates

Lab Sample ID: 2070981
Field Sample ID: G1NE0906 - PAR1491

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	96.3	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	93.9	P	50 - 300

Lab Sample ID: 2070982
Field Sample ID: G1NE0917 - PAR262

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	107	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	102	P	50 - 300

Lab Sample ID: 2070983
Field Sample ID: G1NE0907 - MillBel

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	100	P	50 - 300
SOP-PCR-1.2	SKETA-qPCR	259	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	97.6	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	105							
SOP-PCR-1.0	HF183-qPCR	110	131	71.5	92.1	112			2.00
				83.9					
SOP-PCR-1.2	GULL2-qPCR	210	137	106	81.9	81.0			
				82.4					
SOP-PCR-1.3	GFD-purified-qPCR	13.2	151	137	210	183			29.4
				293					
SOP-PCR-1.4	BacR-qPCR	121	109	56.9	68.9	56.8			11.7
				69.1					

Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
EPA 1603 / E82001	Escherichia coli by membrane filter method by Advanced Environmental Laboratory in Gainesville	2070977, 2070978, 2070979
SM 10200 H (mod.) / E31780	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2070974, 2070975, 2070976
SOP-PCR-1.0 / E31780	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2070981, 2070982, 2070983
SOP-PCR-1.2 / E31780	Detection and quantification of coastal bird specific Catellicoccus marimammalium Gull2 genetic marker with quantitative polymerase chain reaction	2070983
SOP-PCR-1.3 / E31780	Detection and quantification of bird specific Helicobacter GFD genetic marker with quantitative polymerase chain reaction	2070983

Reference Method Descriptions

Method / DoH Cert #	Description	<u>Associated Samples</u>
SOP-PCR-1.4 / E31780	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2070981, 2070982, 2070983
SOP-PCR-4.0 / E31780	Preparation of Samples by Filtration and held for qPCR Analysis	2070981, 2070982, 2070983

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
EPA 1603	03/18/2019	03/18/2019 15:35		03/19/2019 13:05	AEL - Gainesville	2070977, 2070978, 2070979
SM 10200 H (mod.)	03/19/2019	03/19/2019 14:20	Edwin Gonzalez-Soto	04/03/2019 08:48	Jennifer Mihalic	2070974, 2070975, 2070976
SOP-PCR-1.0	03/19/2019	03/19/2019 11:25	Sarah Menz	04/04/2019 07:48	Mailin Lopez	2070981, 2070982, 2070983
SOP-PCR-1.2	03/19/2019	03/19/2019 11:25	Sarah Menz	04/10/2019 12:46	Mailin Lopez	2070983
SOP-PCR-1.3	03/19/2019	03/19/2019 11:25	Angela Smith	04/22/2019 10:54	Sarah Menz	2070983
SOP-PCR-1.4	03/19/2019	03/19/2019 11:25	Sarah Menz	04/04/2019 11:55	Mailin Lopez	2070981, 2070982, 2070983
SOP-PCR-4.0	03/19/2019	03/19/2019 11:25	Angela Smith	04/01/2019 11:32	Sarah Menz	2070981, 2070982, 2070983