

From: [Ayres, Joshua](#)
To: [Reaves, Shawn D](#)
Cc: [Homann, Lisa](#); [Watts, John](#); [Sprague, Kara](#)
Subject: RE: times off in LIMS
Date: Friday, November 13, 2020 8:29:02 AM
Attachments: [Re RO-2020-11-02-44 NW-DIST-2020-11-05-01.msg](#)
[FW RO-2020-11-02-30 NE-DIST-2020-11-04-02.msg](#)
[Re RO-2020-10-19-70 NE-DIST-2020-11-04-01.msg](#)
[image001.png](#)
Importance: High

Shawn –

We got confirmation from the samplers about the time difference.

Please subtract one hour from the collection times in LIMS for all samples in these three events & add the email threads to the event files.

Thanks.



JOSHUA AYRES
Environmental Specialist III
DEP Laboratories
Voice: 850.245.8206
Joshua.Ayres@dep.state.fl.us

From: Ayres, Joshua
Sent: Thursday, November 12, 2020 13:59
To: Sprague, Kara <Kara.Sprague@floridadep.gov>
Cc: Homann, Lisa <Lisa.Homann@FloridaDEP.gov>; Watts, John <John.Watts@FloridaDEP.gov>; Reaves, Shawn D <Shawn.Reaves@FloridaDEP.gov>
Subject: RE: times off in LIMS
Importance: High

Kara –

So.. It looks like the tablets weren't adjusted for daylight savings but the handwrite overflow COC were.

I just pulled additional containers for NE-DIST-2020-11-04-01 & NE-DIST-2020-11-04-02, they confirm the collection times listed on the overflow COCs.

The containers for NW-DIST-2020-11-05-01 don't have collection information on them for me to compare but looking at the submittal forms & pictures of overflow COC it looks like everything is 1 hr. off.
2 hrs. off in LIMS when adjusting for central time zone.

I'll have to contact the customers to get confirmation before making any corrections



RQ-2020-10-19-70

Customer: WQAP

Project ID: SMP

Collected By: Kelly McDonald, Brian Davis

Field Report Prepared By: Kelly McDonald

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to laboratory			
WIN ID/Field ID:									
		G5NE0591							
Location:		Unnamed Drain South Entrance							
Matrix: Marine						(Check one) ☒ GRAB			
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp C (µmho)
11/3/2020	13:30 EST	7.93	112.6	23.1	8.04	0.3	0.9 ☐ VOB (S)	1.4	509

Project Name Unnamed CM		Project Number RQ-2020-10-19-70		ANALYSIS REQUESTED (Include)					
Report To J. Perry		Report CC DEP-Overflows@dep.state.fl.us		Preservative					
Company/Address 8800 Baymeadows Way W Suite 100 Jax, FL, 32256		Phone # 904-256-1700		FAX #		NUMBER OF CONTAINERS			
Sampler's Signature B. Davis		Sampler's Printed Name Brian Davis				Entero Fecal			
CLIENT SAMPLE ID	LAB ID	SAMPLING DATE TIME		Matrix					
G5NE0591		11/3/20 12:30		SW	2	1	1		



JOSHUA AYRES
Environmental Specialist III
DEP Laboratories
Voice: 850.245.8206
Joshua.Ayres@dep.state.fl.us

From: Sprague, Kara <Kara.Sprague@FloridaDEP.gov>

Sent: Thursday, November 12, 2020 12:57

To: Ayres, Joshua <Joshua.Ayres@dep.state.fl.us>Cc: Homann, Lisa <Lisa.Homann@FloridaDEP.gov>

Subject: times off in LIMS

Hey Josh,

It looks like the sampling times are an hour off for events starting sometime around 11/3.

We've found NE-DIST-2020-11-04-01, NE-DIST-2020-11-04-02, and NW-DIST-2020-11-05-01 so far.

Can you take a look?

Thanks,

Kara

Kara Sprague

Environmental Specialist II

FDEP Laboratory - Biology Program

Florida Department of Environmental Protection

Kara.Sprague@Floridadep.gov

Office: 850.245.8826

Login Checklist

RQ-2020-11-02-08

Login Station ID: #4

Shipping Method: FedEx UPS | HD | Greyhound

Date/Time of Receipt: 11/05/2020 @ 9:25

*Cooler Temperature	Samples Frozen?	* All Samples in Cooler Preserved with		Number of Sample Containers in Cooler	Evidence Tape				Waybill Tracking #
					Present?		*Intact?		
	YES	HNO ₃	Formalin		Yes	No	Yes	No	
1.7°C				11		x			9117 6381 6496
1.6°C				10		X			9293 8006 8738

COOLER CHECK

*All samples received on ice unless otherwise noted. HNO₃ and Formalin preserved samples do NOT have a temperature requirement.

- If the temperature of a cooler exceeds 6.0 oC (above 10.0 oC for W-1-4-DIOX and microbiology) or received without ice, identify affected samples in an NCR. NCR# _____
- DOH coolers can exceed 2.0 °C above the standard temperature (6.0 or 10.0 °C for W-1-4-DIOX) without login confirmation from customer for samples requiring ice preservation; identify affected samples in an NCR. NCR# _____
- If cooler or samples are received with a damaged Evidence Seal; identify the affected samples in an NCR. NCR# _____
- If coolers or samples are received late; identify the affected samples in an NCR. NCR# _____

Chain of Custody/ Field Sheet(s) Included?

Yes ☒ No ☐

Micro-Biology Overflow Chain of Custody/ Field Sheet(s) Included?

Yes ☒ No ☐

CONTAINER CHECK

Evidence Tape on Bottles?

Yes ☐ No ☐ NA ☒

Evidence Tape intact? Yes ☐ No ☐

If Criminal, photographs taken?

Yes ☐ No ☐

Containers intact? Yes ☒ No ☐

Caps on tight?

Yes ☒ No ☐

Sufficient Sample Volume:

Yes ☒ No ☐

If NO is checked above, generate an NCR listing affected sample IDs in its appropriate category. NCR# _____

CHLORINE CHECK REQUIRED? (Blue dot on container)

Yes ☒ No ☐ Init: Tyler R

Chlorine detected? (One container checked per Field ID)

Yes ☐ No ☒ Init: Tyler R

If chlorine is detected, generate an NCR listing affected sample IDs . NCR# _____

PRESERVATION CHECK (EXCLUDING SAMPLES SENT TO OVERFLOW LABS, "OV-")

**Acid Preserved Samples: pH< 2.0?

Yes ☒ No ☐ NA ☐ Init: Tyler R

**W-1-4-DIOX (Green dot on container) preserved to pH<4.0?

Yes ☐ No ☐ NA ☒ Init: Tyler R

If samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

Coolers Unpacked/Checked by: Tyler Reynolds

Date: 11/05/2020

Event contains NCRs (Y/N)? none

Event ID: Nassau SCI's
NE-DIST-2020-11-05-01 (SMP)

Page 1 of 2, 06/ Date Received: 05-NOV-2020 09:23

Login Checklist

Login Preservation Equipment

Lot # & Exp: Date

Additional Comments:

Infrared Thermometer ID:	#4 Temp Gun EXP:11/05/2020
pH Test Strips 0 – 6	
pH Paper 0 – 3	
pH Paper 0 – 13	231019 EXP:11/01/2022
Chlorine Test Strips	8261 EXP:07/2021

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ENCORE SAMPLES

S-VOC-MS samples in Encores must be frozen within 48 hours of collection.

Were Encore samples received? Yes _____ Init: _____

Date and time placed in freezer _____

Were all samples placed in freezer within 48 hours of collection? Yes _____ No _____

If no, were samples shipped on dry ice? Yes _____ No _____

If S-VOC-MS samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

FOR CLEAN LAB USE ONLY – Total Mercury Preservation Check (W-HG-AF;W-HG-AF-F):

Samples preserved within 48 hours? Yes _____ No _____

Date and time samples preserved: _____

FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION
CHAIN OF CUSTODY FORM

Date: 11/4/2020
Customer: NE-DIST
RQ: RQ-2020-11-02-08

Collected By: Jeremy Parrish, Kim Appleby
Sampling Agency: FDEP
Project ID: SMP

PLEASE RETURN THE ORIGINAL OF THIS FORM WITH SAMPLE SUBMITTAL
FORMS AND THE COOLER PACKING WORKSHEET TO THE LAB

Number of Coolers Shipped: 1

FIELD ID (Labels) submitted under this RQ for this collection date.

Site Location	Field ID/WIN ID
Deep Creek at CR 121	19010048

RELINQUISHED BY(SIGNATURE):

Jeremy Parrish

DATE/TIME: 11/4/2020

THIS SECTION TO BE COMPLETED BY LABORATORY

Rec'd/Inspected By: *Tyler Reynolds*

Rec'd/Inspected Date: *11/05/2020*

CALIBRATION AND VERIFICATION LOG (FDEP SOP FT 1000-FT 1500, FD 1000-FD 4000)

Boldly "X" this box if there are qualified data on this page.

Meter ID: 5126

RQ: 2020-11-02-08

Project: SCE Pauphery

- Notes: (1) Always wait for meter to stabilize before recording any readings.
 (2) Report all digits displayed. Do not round before reporting measurements. (See special instructions for depth).
 (3) For Calibrations, record calibrated meter reading. Do not record initial meter reading before calibration.

Temperature (Quarterly) FT 1400

Date of Last Temperature Verification 10/14/20

DO DEP SOP FT 1500	Name	Date	Time CT-ET	Temp °C	Baro-meter mmHg	D.O. Chart mg/L	Meter D.O. mg/L	% DO	Probe Charge	Probe Gain	Pass / Fail	Lab / Field
Calibr.	J. Lamsh	11/4/20	738	22.0	769	8.8	8.8	101.2	—	1.000	P/F	L/F
ICV	J. Lamsh	11/4/20	746	22.1	769	8.8	8.84	101.3	—	1.000	P/F	L/F
CCV	Kim Appleby	11/4/20	1517	22.8	769.2	8.7	8.75	101.7	—	1.020	P/F	L/F
CCV											P/F	L/F

DO Acceptance criteria from Table ± 0.3 mg/L

Rapid-Pulse Sensors: DO Gain Range 0.7 to 1.4; DO Charge Range 25-75.

Optical: DO gain range 0.85 to 1.15 (Pro DSS 0.75 to 1.50); DO charge N/A. Steady-state & Galvanic Sensors: DO Gain & Charge N/A.

Spec. Cond. FT 1200	Name	Date	Time CT-ET	Lot #	Expir. Date	Standard μ hos/cm	Meter Reading μ hos/cm	Pass / Fail	Lab / Field
Calibr.	J. Lamsh	11/4/20	747	200902B	9/21	1000	1000	P/F	L/F
ICV	J. Lamsh	11/4/20	748	200902A	9/21	100	102.5	P/F	L/F
CCV	Kim Appleby	11/4/20	1520	191025A	11/20	1418	1417	P/F	L/F
CCV								P/F	L/F

Conductivity Acceptance criteria $\pm 5\%$

pH DEP SOP FT 1100	Name	Date	Time CT-ET	Lot #	Expir. Date	pH Buffer SU	Temp °C	Meter reading SU	mV	Pass / Fail	Lab / Field
Calibr.	J. Lamsh	11/4/20	750	2001C31	1/22	7.00	22.7	7.01	-49.4	P/F	L/F
Calibr.	J. Lamsh	11/4/20	752	2006B06	5/22	4.00	22.7	4.00	130	P/F	L/F
Calibr.	J. Lamsh	11/4/20	755	202911	8/21	10.00	22.7	10.00	-214.5	P/F	L/F
ICV	J. Lamsh	11/4/20	757	2006B06	5/22	4.00	22.3	4.02	127.8	P/F	L/F
CCV	Kim Appleby	11/4/20	1525	20-2911	8/21	10.00	23.5	9.96	-216.3	P/F	L/F
CCV										P/F	L/F

pH Acceptance criteria ± 0.2 SU; mV pH 7 Range 0 ± 50 ; mV pH 4 Range $+180 \pm 50$; mV pH 10 Range -180 ± 50 ;

If mV are recorded: slope from 7 to 10 _____, slope from 4 to 7 _____ (both must be between 165 and 180 mV)

Does meter have a depth sensor that will be used to measure total depth & sample depth? YES / NO / NA (not surf. water project)

If YES, complete daily Calibr. & ICV below and list date of last quarterly depth verification: _____

If NO, what will be used? (circle one) Secchi Disk Line / Sonar Unique ID: _____; Date of last verification: _____

Depth Sensor (Daily Calibration & ICV)	Name	Date	Time CT-ET	Calibrated Value (0.00 or Offset), meters	ICV Value, meters	Pass / Fail	Lab / Field
Pressure mode in air	J. Lamsh	11/4/20	755	0.00	0.02	P/F	L/F

Report two decimal places. Round numbers ≤ 4 down, ≥ 5 up. ICV acceptance criteria $\pm 5\%$ or ± 0.05 m, whichever is greater.

COMMENTS:

FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION
CHAIN OF CUSTODY FORM

Date: 11/4/2020
Customer: NE-DIST
RQ: RQ-2020-11-02-08

Collected By: Kelly McDonald, Brian Davis
Sampling Agency: FDEP
Project ID: SMP

PLEASE RETURN THE ORIGINAL OF THIS FORM WITH SAMPLE SUBMITTAL
FORMS AND THE COOLER PACKING WORKSHEET TO THE LAB

Number of Coolers Shipped: 1

FIELD ID (Labels) submitted under this RQ for this collection date.

Site Location	Field ID/WIN ID
Calkins At Turner Cemetary Rd	 19010076

RELINQUISHED BY(SIGNATURE): *Brian Davis*

DATE/TIME: 11/4/20 1600

THIS SECTION TO BE COMPLETED BY LABORATORY

Rec'd/Inspected By: *Tyler Reynolds*

Rec'd/Inspected Date: *11/05/2020*

CALIBRATION AND VERIFICATION LOG (FDEP SOP FT 1000-FT 1500, FD 1000-FD 4000)

Boldly "X" this box if there are qualified data on this page.

Meter ID: ProDSS Desi

RQ: 2020-11-02-08

Project: Calkins SCI

Notes: (1) Always wait for meter to stabilize before recording any readings.

(2) Report all digits displayed. Do not round before reporting measurements. (See special instructions for depth).

(3) For Calibrations, record calibrated meter reading. Do not record initial meter reading before calibration.

Temperature (Quarterly) FT 1400

Date of Last Temperature Verification 10/14/20

DO FDEP SOP FT 1500	Name	Date	Time CT-ET	Temp °C	Baro- meter mmHg	D.O. Chart mg/L	Meter D.O. mg/L	% DO	Probe Charge	Probe Gain	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/4/20	853	21.8	769.1	8.9	8.9	101.2	✓	1.07	P/F	L/F
ICV			854	21.8	769.2	8.9	8.87	101.1	✓	1.07	P/F	L/F
CCV			1544	22.3	768.3	8.8	8.87	102.1	✓	1.07	P/F	L/F
CCV											P/F	L/F

DO Acceptance criteria from Table ± 0.3 mg/L.

Rapid-Pulse Sensors: DO Gain Range 0.7 to 1.4; DO Charge Range 25-75.

Optical: DO gain range 0.85 to 1.15 (Pro DSS 0.75 to 1.50); DO charge N/A. Steady-state & Galvanic Sensors: DO Gain & Charge N/A.

Spec Cond. FT 1200	Name	Date	Time CT-ET	Lot #	Expir. Date	Standard μ hos/cm	Meter Reading μ hos/cm	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/4/20	856	200902B	09/2021	1000	1000	P/F	L/F
ICV			858	200902A	09/2021	10.0	103.0	P/F	L/F
CCV			1546	191025A	11/20	1413	1405	P/F	L/F
CCV								P/F	L/F

Conductivity Acceptance criteria $\pm 5\%$

pH FDEP SOP FT 1100	Name	Date	Time CT-ET	Lot #	Expir. Date	pH Buffer SU	Temp °C	Meter reading SU	mV	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/4/20	1003	2001C31	01/2022	7.0	22.7	7.01	23.1	P/F	L/F
Calibr.	Kelly McDonald		1011	2006B06	05/2022	4.00	22.7	4.00	155.5	P/F	L/F
Calibr.			1014	2002911	08/2021	10.03	22.6	10.03	193.1	P/F	L/F
ICV			1018	2006B06	05/2022	4.00	22.7	4.07	150.9	P/F	L/F
CCV	Brian Davis	11/4/20	1548	2001C31	01/22	7.01	23.5	7.07	23.6	P/F	L/F
CCV										P/F	L/F

pH Acceptance criteria ± 0.2 SU; mV pH 7 Range 0 $\pm$ 50;

mV pH 4 Range $+180 \pm 50$;

mV pH 10 Range -180 ± 50 ;

If mV are recorded: slope from 7 to 10 _____, slope from 4 to 7 _____ (both must be between 165 and 180 mV)

Does meter have a depth sensor that will be used to measure total depth & sample depth? YES / NO / NA (not surf. water project)

If YES, complete daily Calibr. & ICV below and list date of last quarterly depth verification: 10/14/20

If NO, what will be used? (circle one) Secchi Disk Line / Sonar Unique ID: _____; Date of last verification: _____

Depth Sensor (Daily Calibration & ICV)	Name	Date	Time CT-ET	Calibrated Value (0.00 or Offset), meters	ICV Value, meters	Pass / Fail	Lab / Field
Pressure mode in air	K. McDonald	11/4/20	1019	0.000	0.000	P/F	L/F

Report two decimal places. Round numbers ≤ 4 down, ≥ 5 up. ICV acceptance criteria $\pm 5\%$ or ± 0.05 m, whichever is greater.

COMMENTS:



FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION STRATEGIC MONITORING PROGRAM FIELD SHEET

January 2020

Meter Passed End
of Day CCV
☒ Yes / ☐ No

WIN Station Name: Deep Creek At Cr 121

Date: 11/4/2020

WIN/Field ID: 19010048

WBID: 2196

ENR:

Latitude: 30.4557

County: NASSAU

Org ID: 21FLA

Longitude: -82.0041

Receiving Body of Water: Nassau - St. Marys

RQ- 2020-11-02-08

Sampling Team Members		WQ Measurements	Sample Collection	Documentation	Preservation	Preservation Check
☒	Jeremy Parrish	X		X		
☒	Kimberly Applebee		X		X	X
☐						
☐						

Sampling Equipment	
☒ Sample Container	☐ Carboy
☐ Dipper	☐ Van Dorn
☐ Syringe	☐ Pole
Depth measured with: Meter	
Secchi Equipment ID:	
Collection Method	
☒ Wading	☐ Canoe/Kayak
☐ From Shore	☐ Electric Motor
☐ Other	☐ Gasoline Motor

Water Level: ☒ Normal ☐ Low ☐ High ☐ Dry ☐ Pooled ☐ Flooded		Meter ID# STITCH	
Photos: NO	Flow: FLOW	Tide:	Tide Times: High: Low:
Biological Samples Collected: ☐ None ☒ HA ☒ SCI ☒ RPS ☐ Algae ID ☒ LVS ☐ LVI Other:			
SBIO ID Numbers:			
Notes:			

Duplicate Sample

Time Zone: EST	Time: N/A	Field ID: N/A	(WIN ID_DUP)
WIN DMT Activity ID:			

Blank

Time Zone:	Time: N/A	Field ID: N/A	(Blank Type_DDMMYYYY)
DI Source: N/A	Location: N/A		
DI Type: N/A	Equipment ID: N/A		

LIMS EVENT ID: WIN Import ID:

Enter Field Parameters, Verify Station ID In LIMS DMT: (Initials/Date) QC Station ID, Field Parameters In LIMS DMT: (Initials/Date)

Scan/Save Field Sheets (Initials/Date) Migrate Data to WIN: (Initials/Date) Verify Data In WIN: (Initials/Date)



RQ-2020-11-02-08

Customer: WQAP

Project ID: SMP

Collected By: Jeremy Parrish, Kimberly Applebee

Field Report Prepared By: Jeremy Parrish

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to contract laboratory				
WIN ID/Field ID:  19010048										
Location: Deep Creek At Cr 121										
Matrix: Fresh						(Check one) ☒ GRAB ☐ COMPOSITE				
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp COND (µmhos/cm)	Salinity (PPTH)
11/4/2020	11:30 EST	6.98	67.7	14	4.26	0.15	0.3 ☒ VOB (S)	0.3	56	0.03
Central Lab please use top row for login purposes										
EST										
Parameter Suite		Parameter Methods				Preservation		pH Check	# Bottles sent to Lab	Bottle Group
Preserved Nutrients (P-500ML)		☒ W-NH3 / W-NO2NO3 / W-TKN / W-TOC / W-S-A-TP				☒ Ice ☒ H2SO4*		☒ <2	1	A
		Lot #	Sa0038110		Exp:	2/7/20				
Metals (P-500ML)		☒ W-ICPMS ☒ W-ICP ☒ W-HARD				☒ Ice ☒ HNO3*		☒ <2	1	A
		Lot #	Na9354060		Exp:	12/20/20				
Filtered Nutrients (P125ML)		☒ W-PO4-F ☒ Field Filtered 0.45 µm Filter				☒ Ice			1	A
		Filter Lot #	Rocb48617							
		Syringe Lot #								
Chlorophyll (BP-1L)		☒ CHLSUITE-W				☒ Ice			1	A
Physical/Aggregate (Fresh) (P-1L)		☒ Alkalinity / W-F / W-TSS / TURBIDITY / W-CL-IC / W-COLOR / W-SO4-IC / W-TDS				☒ Ice			1	A
Physical/Aggregate (Marine) (P-1L)		☐ Alkalinity / W-BR-IC / W-F / W-TSS / TURBIDITY				☐ Ice				
Macroinvertebrates (PJ-2L)		☒ MI-FW-QLDC				☒ Formalin			2	B
Periphyton (PT-50ML)		☐ ALGAE-ID				☐ Ice				
Microbiology (Contract Lab Bottle)		☒ E Coli ☐ F Coli ☐ Enterococci ☒ Contract Lab				☒ Ice			1	A
Molecular (QPCR-P-500ML)		☐ PCR-HF183 ☐ PCR-BacR ☐ PCR-DG3 ☐ PCR-GULL2 ☐ PCR-GFD ☐				☐ Ice				
Tracers (BG-500ML)		☒ W-E8321-DI ☒ W-E8321-MS				☒ Ice			1	A
Pesticides (BG-500ML) (BG-1L)		☐ W-E8321-DI ☐ W-E8321-MS ☒ W-PSNP-TQ ☐ W-CT-CL-R ☐ W-PCL-SQ-R ☐ W-CARB-AA (5 ml of MCAA** Buffer Solution)				☒ Ice			3	A
						☐ Ice				
Name: Jeremy Parrish Kimberly Applebee				Signature: 					Date: 11/4/2020	

*2.0 ml of Preservative used unless noted otherwise.

**MCAA: Monochloroacetic acid buffer solution



RQ-2020-11-02-08

Customer: WQAP

Project ID: SMP

Collected By: Kelly McDonald, Brian Davis

Field Report Prepared By: Kelly McDonald

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to contract laboratory				
WIN ID/Field ID: 										
19010076										
Location: Calkins At Turner Cemetary Rd										
Matrix: Fresh						(Check one) ☒ GRAB ☐ COMPOSITE				
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp COND (µmhos/cm)	Salinity (PPTH)
11/4/2020	12:00 EST	7.27	72.1	15	5.78	0.15	0.4 ☒ VOB (S)	0.4	61.5	0.03
Central Lab please use top row for login purposes										
EST										
Parameter Suite		Parameter Methods				Preservation		pH Check	# Bottles sent to Lab	Bottle Group
Preserved Nutrients (P-500ML)		☒ W-NH3 / W-NO2NO3 / W-TKN / W-TOC / W-S-A-TP				☒ Ice ☒ H2SO4*		☒ <2	1	A
		Lot #		SA0038110		Exp:		2/7/21		
Metals (P-500ML)		☒ W-ICPMS ☒ W-ICP ☒ W-HARD				☒ Ice ☒ HNO3*		☒ <2	1	A
		Lot #		NA9354060		Exp:		12/20/2020		
Filtered Nutrients (P125ML)		☒ W-PO4-F ☒ Field Filtered 0.45 µm Filter				☒ Ice			1	A
		Filter Lot #		R9BA86875						
		Syringe Lot #								
Chlorophyll (BP-1L)		☒ CHLSUITE-W				☒ Ice			1	A
Physical/Aggregate (Fresh) (P-1L)		☒ Alkalinity / W-F / W-TSS / TURBIDITY / W-CH-IC / W-COLOR / W-SO4-IC / W-TDS				☒ Ice			1	A
Physical/Aggregate (Marine) (P-1L)		☐ Alkalinity / W-BR-IC / W-F / W-TSS / TURBIDITY				☐ Ice				
Macroinvertebrates (PJ-2L)		☒ MI-FW-QLDC				☐ Formalin			2	
Periphyton (PT-50ML)		☐ ALGAE-ID				☐ Ice				
Microbiology (Contract Lab Bottle)		☒ E Coli ☐ F Coli ☐ Enterococci ☒ Contract Lab				☒ Ice			1	A
Molecular (QPCR-P-500ML)		☐ PCR-HF183 ☐ PCR-BacR ☐ PCR-DG3 ☐ PCR-GULL2 ☐ PCR-GFD ☐				☐ Ice				
Tracers (BG-500ML)		☐ W-E8321-DI ☐ W-E8321-MS				☐ Ice				
Pesticides (BG-500ML) (BG-1L)		☒ W-E8321-DI ☒ W-E8321-MS ☒ W-PSNP-TQ ☐ W-CT-CL-R ☐ W-PCL-SQ-R ☐ W-CARB-AA (5 ml of MCAA** Buffer Solution)				☒ Ice			3	A
						☐ Ice				
Name: Kelly McDonald Brian Davis				Signature: <i>Brian Davis</i>				Date: 11/4/2020		

*2.0 ml of Preservative used unless noted otherwise.

**MCAA: Monochloroacetic acid buffer solution



FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION STRATEGIC MONITORING PROGRAM FIELD SHEET

January 2020

Meter Passed End
of Day CCV
Yes / No

WIN Station Name: Calkins At Turner Cemetary Rd **Date:** 11/4/2020
WIN/Field ID: 19010076 **WBID:** 2264 **ENR:** **Latitude:** 30.328
County: BAKER **Org ID:** 21FLA **Longitude:** -82.2307
Receiving Body of Water: Nassau - St. Marys **RQ:** 2020-11-02-08

Sampling Team Members		WQ Measurements	Sample Collection	Documentation	Preservation	Preservation Check
☒	Kelly McDonald	x	x		x	x
☒	Brian Davis	x		x	x	x
☐						
☐						

Sampling Equipment	
☒ Sample Container	☐ Carboy
☐ Dipper	☐ Van Dorn
☐ Syringe	☐ Pole
Depth measured with: Meter	
Secchi Equipment ID:	
Collection Method	
☒ Wading	☐ Canoe/Kayak
☐ From Shore	☐ Electric Motor
☐ Other	☐ Gasoline Motor

Water Level: ☒ Normal ☐ Low ☐ High ☐ Dry ☐ Pooled ☐ Flooded						Meter ID# DESI					
Photos: No	Flow: FLOW			Tide:		Tide Times: High: Low:					
Biological Samples Collected: ☐ None ☒ HA ☒ SCI ☒ RPS ☐ Algae ID ☒ LVS ☐ LVI Other:											
SBIO ID Numbers:											
Notes:											

Duplicate Sample

Time Zone: EST	Time: N/A	Field ID: N/A	(WIN ID_DUP)
WIN DMT Activity ID:			

Blank

Time Zone:	Time: N/A	Field ID: N/A	(Blank Type_DDMMYYYY)
DI Source: N/A	Location: N/A		
DI Type: N/A	Equipment ID: N/A		

LIMS EVENT ID: **WIN Import ID:**
Enter Field Parameters, Verify Station ID In LIMS DMT: **QC Station ID, Field Parameters In LIMS DMT:**
(Initials/Date) (Initials/Date)
Scan/Save Field Sheets BD 11/4/2020 **Migrate Data to WIN:** **Verify Data In WIN:**
(Initials/Date) (Initials/Date) (Initials/Date)

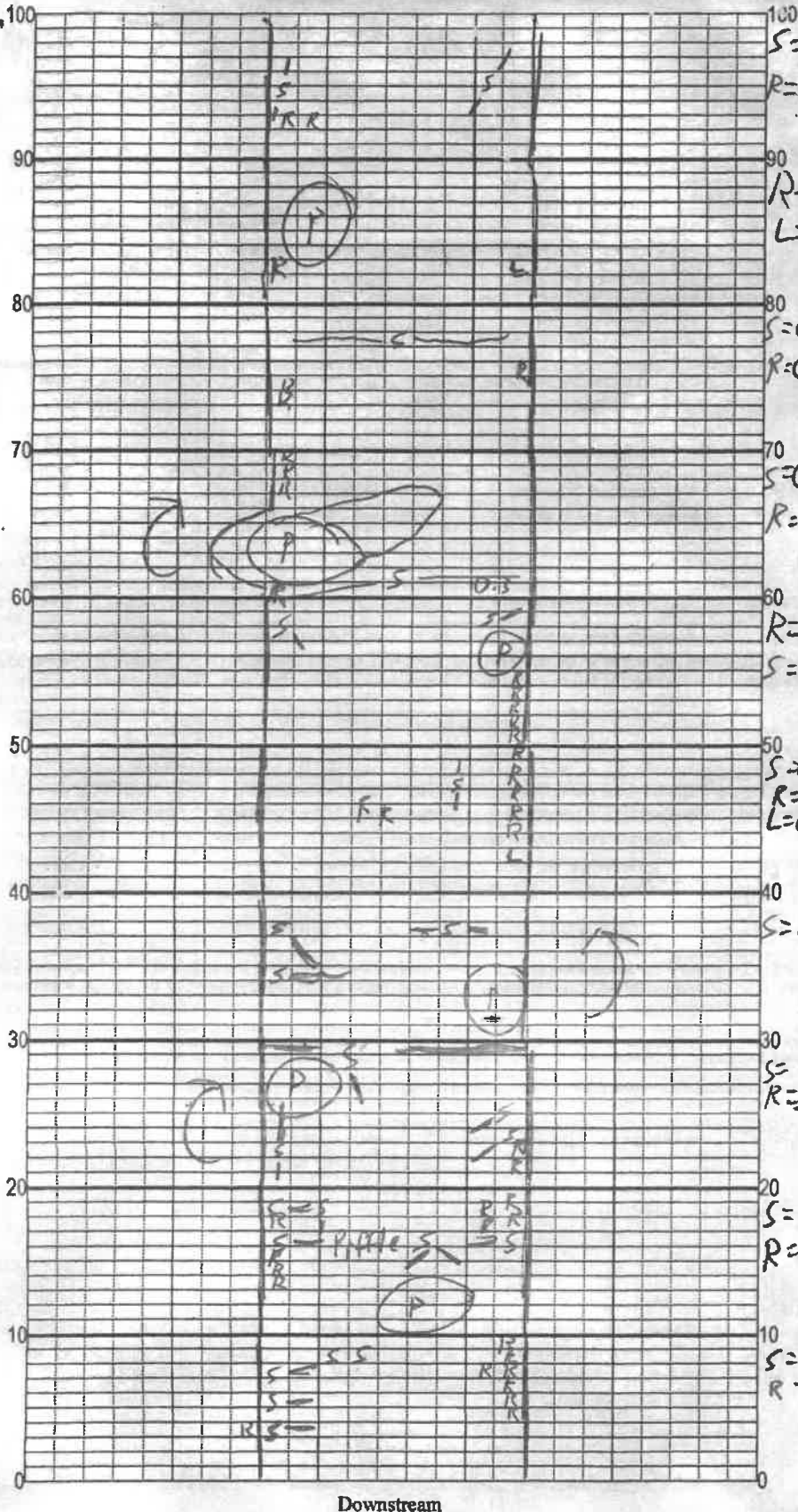
DEP Form FD 9000-4 Stream/River Habitat Assessment Sketch Sheet

0.6 m

Left Bank

0.25 m/s
= 0.5
= 2.5 m

0.25 m
= 0.1 m/s
= 0.4 m



S=0.5
R=0.2
R=0.2
L=0.1

Location: Calkins of Tupper Lake
Date: 11/4/20
Sampler: Brian Davis Kelly McBride

100 m: Latitude _____
Longitude _____
0 m: Latitude _____
Longitude _____

Substrates: Code key, draw proportionate habitat abundance. %

☐ S	Snags	2
☐ R	Roots/undercut banks	2.5
☐ L	Leaf Packs (or mats)	< 1
☐	Aquatic Macrophytes	
☐		
☐		
☐ P	Pools	total # observed = 6 # range expected = 3

Average width 2.5 m
Average depth 0.5 m

Velocity measured at 9.50 meter mark.

WQ & chemistry taken at 0 meter mark.

Habitat Smothering:
Note areas (on map) where sand or silt is smothering substrates, limiting habitability.

Bank Stability:
Note areas (on map) with unstable, eroding banks.

Riparian Buffer Width:
Note areas (on map) where natural vegetation is altered or eliminated.

Plants observed / other notes:

S=1.0
R=0.6
S=1.0
R=1.5

S=50/250 2%
R=6.2/250 2.5%
L

Length of grid represents 100 m of stream (not linear meters).
(Horizontal scale is double vertical scale, draw proportionately).

DEP Form FD 9003-4 Stream/River Habitat Assessment Field Sheet

SAMPLING AGENCY: <u>DEAR NERO</u>		STORET STATION NUMBER: <u>19010076</u>	DATE (mm/dd/yyyy): <u>11/4/20</u>	RECEIVING BODY OF WATER: <u>Cedar Creek</u>
REMARKS:	COUNTY: <u>Baker</u>	LOCATION: <u>Turner Glicks Cemetery Rd</u>		FIELD ID/NAME:

Habitat Parameter	Optimal	Suboptimal	Marginal	Poor
Primary Habitat Components	Four or more major productive habitats present (snags, tree roots, aquatic vegetation, leaf packs (partially decayed), rock)	Three major productive habitats present. Adequate habitat. Some substrates may be new fall (fresh leaves or snags)	Two major productive habitats present. Less than desirable habitat, frequently disturbed or removed	One or less major productive habitat. Lack of habitat is obvious, substrates unstable or smothered
Substrate Diversity <u>8</u>	20 19 18 17 16	15 14 13 12 11	10 9 8 7 6	5 4 3 2 1
Substrate Availability <u>5</u>	Greater than 30% major productive habitat present at site 20 19 18 17 16	16% to 30% major productive habitat, by aerial extent 15 14 13 12 11	6% to 15% major productive habitat 10 9 8 7 6	Less than 5% major productive habitat (5) 4 3 2 1
Water Velocity <u>14</u>	Max. observed at typical transect: > 0.25 m/sec. But < 1 m/sec ≥ 0.33 0.31 0.29 0.27 > 0.25 20 19 18 17 16	Max. observed at typical transect: > 0.1 to 0.25 m/sec 0.25 0.21 0.17 0.13 > 0.1 15 (14) 13 12 11	Max. observed at typical transect: 0.05 to 0.1 m/sec 0.1 0.09 0.07 0.06 0.05 10 9 8 7 6	Max. observed at typical transect: < 0.05 m/sec; or spate occurring: > 1 m/sec < 0.05 0.04 0.03 0.01 < 0.01 5 4 3 2 1
Habitat Smothering <u>13</u> Primary Score <u>40</u>	Adequate number of stable pools (1-2 per 12 times width) and < 25% of habitats affected by sand, silt, or algae. 20 19 18 17 16	Adequate number of stable pools (1-2 per 12 times width) and > 25% of habitats affected by sand, silt, or algae. 15 14 (13) 12 11	Does not have required number of stable pools (1-2 per 12 x width) and/or has shallow pools (< 2 x prevailing depth). 10 9 8 7 6	Stable pools are absent. Most habitats affected by sand, silt, or algae accumulation. 5 4 3 2 1
Secondary Habitat Components Artificial Channelization <u>16</u>	Expected sinuosity given the stream width. No evidence of dredging or artificial straightening. No spoil banks. 20 19 18 17 (16)	Good sinuosity within old channelized area. Evidence of dredging or straightening in the past (> 25 yrs) but mostly recovered. 15 14 13 12 11	Straightened with trapezoidal cross section, but has a small degree of sinuosity developed within channelized area. 10 9 8 7 6	Straightened or engineered by dredging, has trapezoidal or box cut cross section, lacks required pools. May have spoil banks. 5 4 3 2 1
Bank Stability Right Bank <u>8</u> Left Bank <u>8</u>	Bankfull > 60% of bank height. Slope of bank ≤ 60° from bankfull to top of bank. Bankfull is within or above the woody root zone with few raw, eroded areas. 10 9	Only meets 2 of the 3 requirements for optimal bank stability. 8 7 6	Only meets 1 of the 3 requirements for optimal bank stability. 5 4	Bankfull < 60% of bank height. Slope of bank > 60°. Bankfull is below the woody root zone with raw, eroded areas. 3 2 1
Riparian Buffer Zone Width Right Bank <u>10</u> Left Bank <u>10</u>	Width of vegetation greater than 18 m 10 9	Width of vegetation > 12 to 18 m 8 7 6	Width of vegetation 6 to 12 m. human activities close to system 5 4	Less than 6 m of buffer zone due to intensive human activities 3 2 1
Riparian Zone Vegetation Quality Right Bank <u>10</u> Left Bank <u>10</u> Secondary Score <u>72</u>	Over 80% of riparian surfaces consist of normal, expected plant community for given sunlight & habitat conditions (e.g., native plants; tree, shrub, and forbs represented, if appropriate). Minimal disturbance. 10 9	> 50% to 80% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). Some disruption in community observed. 8 7 6	25% to 50% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). Disruption obvious. 5 4	Less than 25% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). 3 2 1

112

TOTAL SCORE

Date: <u>11/4/20</u>	Analyst: <u>Brian Davis</u>	Signature: <u>B. Davis</u>
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DEP Form FD 9900-3 Physical/Chemical Characterization Field Sheet

SAMPLE ID _____ ORG ID 21 EIA LATITUDE 10.328
 COUNTY _____ STORET # 1901007E LONGITUDE -82.2307
 DATE _____ TIME 1100 SAMPLING AGENCY DEAR NERO
 SITE NAME Calkins at Turner Cemetery Rd.
 FIELD ID/NAME _____ RECEIVING BODY OF WATER Cedar Creek

RIPARIAN ZONE / STREAM FEATURES

DOMINANT LAND-USE IN WATERSHED (specify relative percent in each category): Forest/Natural ☐ Silviculture ☐ Field/Pasture ☐ Agricultural ☐ Residential ☐ Commercial ☐ Industry ☐ Other (Specify) ☐								Landscape Development Intensity Index (LDI) ☐	
Local Watershed Erosion (select one): ☐ None ☐ Slight ☒ Moderate ☐ Heavy Local Watershed NPS Pollution: ☒ No evidence ☐ Slight ☐ Moderate ☐ Heavy						Typical Width (m) Depth (m)/Velocity (m/sec) Transect 2.5 m wide			
Width of Riparian Vegetation (m) on Each Buffer Side Left Bank: <u>>18</u> Right Bank: <u>>18</u>				Hydrologic Modification Score (per FT 3101) ☐		0.23 m/s			
High Water Mark: <u>0.5</u> + <u>0.5</u> = <u>1.0</u> (m) (above present water level) (present depth) (above bed)				Artificially Impounded ☐ Yes ☒ No		0.5 m deep			
Artificially ☒ NO ☐ Mostly recovered, more sinuous Channelized: ☐ Some recovery ☐ Recent, severe				Canopy Cover % ☐ Open ☐ Lightly Shaded (11-45%) ☒ Heavily Shaded ☐ Moderate Shaded (46-80%)					

SEDIMENT / SUBSTRATE

Sediment Oils ☒ Absent ☐ Slight ☐ Moderate ☐ Profuse				Sediment Odors ☒ Normal ☐ Sewage ☐ Petroleum ☐ Chemical ☐ Anaerobic ☐ Other (Specify): _____				Smothering of Substrates Sand Smothering: None ☐ Slight ☐ Moderate ☒ Severe ☐ Silt Smothering: None ☒ Slight ☐ Moderate ☐ Severe ☐ Algae Smothering: None ☐ Slight ☒ Moderate ☐ Severe ☐																																																
SUBSTRATE TYPE Assessment Tool: ☒ SCI ☒ RPS ☒ LVS ☐ LCI ☐ BioRecon				WATER QUALITY <table border="1"> <thead> <tr> <th></th> <th>Depth (M)</th> <th>Temp. (°C)</th> <th>pH (SU)</th> <th>D.O. (MG/L)</th> <th>D.O. Sat (%)</th> <th>Cond. (UMHO/CM)</th> <th>Salinity (PPT)</th> <th>SECCHI (M)</th> </tr> </thead> <tbody> <tr> <td>Top:</td> <td>0.4</td> <td>15.0</td> <td>5.78</td> <td>7.27</td> <td>72.1</td> <td>61.5</td> <td>0.03</td> <td>0.4</td> </tr> <tr> <td>Mid:</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>☒ VOB</td> </tr> <tr> <td>Bottom:</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>TOTAL DEPTH</td> </tr> <tr> <td>Meter ID:</td> <td colspan="7">Peri</td> <td>0.4</td> </tr> </tbody> </table>									Depth (M)	Temp. (°C)	pH (SU)	D.O. (MG/L)	D.O. Sat (%)	Cond. (UMHO/CM)	Salinity (PPT)	SECCHI (M)	Top:	0.4	15.0	5.78	7.27	72.1	61.5	0.03	0.4	Mid:								☒ VOB	Bottom:								TOTAL DEPTH	Meter ID:	Peri							0.4
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Bottom:								TOTAL DEPTH																																																
Meter ID:	Peri							0.4																																																
Woody Debris (Snags) <u>2.0</u> <u>7</u> Undercut Banks / Roots <u>2.5</u> <u>7</u> Leaf Packs or Mat <u><1</u> <u>1</u> Aquatic Vegetation Rock or Shell Rubble..... Sand..... <u>5</u> Mud / Muck / Silt Other Other				Water Odors Normal ☒ Petroleum ☐ Sewage ☐ Chemical ☐ Other ☐				Water Surface Oils Normal ☒ Sheen ☐ Globbs ☐ Slick ☐ Other ☐																																																
				Water Sample Taken? ☒ Yes ☐ No Sample Preserved? ☒ Yes ☐ No Lot Number: _____ Exp: _____				Color Tannic ☒ Green (Algae) ☐ Clear ☐ Other (Specify) ☐																																																
				Algae Sample Taken? ☐ Yes ☒ No Sample Preserved? ☐ Yes ☒ No Lot Number: _____ Exp: _____				Clarity Clear ☒ Slightly Turbid ☐ Turbid ☐ Opaque ☐																																																
ABUNDANCE Not Obs. Rare Common Abundant Periphyton: ☐ ☒ ☐ ☐ Fish: ☐ ☐ ☒ ☐ Aquatic Plants: ☒ ☐ ☐ ☐ Iron/Sulfur Bacteria: ☐ ☒ ☐ ☐				System Type: Stream ☒ Lake ☐ Wetland ☐ Estuary ☐ Other (Specify) ☐																																																				
AMBIENT FIELD CONDITIONS / NOTES: <u>Much of Habitat smothered.</u> ☐ The antecedent hydrologic conditions have been met to my best knowledge.								SAMPLING TEAM <u>Brice Davis, Kelly McBeard</u>		SIGNATURE <u>[Signature]</u>		DATE: <u>11/4/20</u>																																												

DEP FORM FD 9000-25 Rapid Benthic Survey Field Sheet

Site: <u>Calkins & Turner Cemetery Rd</u>					County: <u>Baker</u>		Date: <u>11/4/20</u>		Investigators: <u>K McDonald, B Davis</u>	
STORET Station Number: <u>190100716</u>										
Transect (m)	Point 1=right 9=left	Algal Thickness Rank (N-6, X)	Estimated	Canopy Cover	Transect (m)	Point 1=right 9=left	Algal Thickness Rank (N-6, X)	Estimated	Canopy Cover	
0	1	N			60	1	X	✓		
	2					2				
	3					3				
	4	X	✓			4				78
	5	X	✓	92		5				
	6	X	✓			6				
	7	N				7				
	8					8				
	9					9			✓	
10	1	N			70	1	X	✓		
	2	N				2				
	3	N				3				
	4	X	✓			4				
	5		✓	89		5				95
	6					6				
	7					7				
	8					8				
	9					9				
20	1	N			80	1	N			
	2	N				2	X			
	3					3				
	4	X				4				92
	5					5				
	6			86		6				
	7					7				
	8					8				
	9					9			✓	
30	1	N			90	1	N			
	2	N				2				
	3					3				
	4	X				4				
	5			88		5				
	6					6				87
	7					7				
	8		✓			8				
	9	N				9				
40	1	N			100	1	X			
	2	N				2				
	3	X				3				
	4					4				
	5			92		5				27
	6					6				
	7					7				
	8	N				8	N			
	9	N				9	N			
50	1	X	✓		Record "N" and check "Estimated" for points deeper than the Secchi depth; algae are presumed absent. Check "Estimated" if thickness estimated for deep points which can be seen but not reached. Record "X" for points shallower than Secchi depth for which substrate cannot be seen or reached with the hand. No estimated rank. Record canopy cover as the number of small densiometer quadrants (out of 96) that HAVE canopy cover. Measure facing upstream at point 4, 5, or 6. Collect algal mat sample following DEP SOP FS 7240 if the percentage of total sampled points with a thickness rank of 4, 5, or 6 is >20%.					
	2		✓							
	3									
	4									
	5			89						
	6									
	7									
	8		✓							
	9		✓							

Secchi depth 0.4 m
Estimated? No

points ranked 4-6 0
total points assessed 0
% points ranked 4-6 0

Check accompanying data collected at same site/date:
Algal mat sample ☐
Linear Veg Survey ☒
Habitat Assessment ☒
SCI/Biorecon ☒
Water sample ☒

RQ- 2020-11-02-08

Algal Thickness Rank	
N	rough, no algae, slimy, algae up to 1mm
3	>1mm - 6mm
4	>6mm - 20mm
5	>20mm - 10 cm
6	>10 cm

8/16
-18
78

20 sweeps
3 major + 1 minor
7 snag
1 root
6 sand (3ea)

CHAIN OF CUSTODY / LABORATORY ANALYSIS REQUEST FORM

©143 Philips Highway, Suite 200 Jacksonville, FL 32256 / P:904 739-2277 / FAX (904) 719-2011

522

Alicia Gonzalez Mandy Sullivan

Page 1 of 1

[illegible]

AIN OF CUSTODY / LABORATORY ANALYSIS REQUEST FORM

ps Highway, Suite 200 Jacksonville, FL 32256 / Ph (904) 739-2277 / FAX (904) 739-2011

SR #

ALS Contact: Mandy Sullivan

Page 1 of 1

Subject Number Q-2020- 11-02-08		ANALYSIS REQUESTED (Include Method Number and Container Preservative)																			
Report CC		Preservat	8	8															Preservative Key 0. None 1. HCL 2. HNO3 3. H2SO4 4. NaOH 5. Zn. Acetate 6. MeOH 7. NaHSO4 8. ICE REMARKS		
AX #		NUMBER OF CONTAINERS	E. Coli	Enterococci																	
Sampler's Printed Name Kelly A McDonald																					
LAB ID	SAMPLING DATE TIME		Matrix																FRESH		
	11/4	1100	SW	1	X																
ep-Overflowmicro@dep.state.fl.us				TURNAROUND REQUIREMENTS				REPORT REQUIREMENTS				INVOICE INFORMATION									
				RUSH (SURCHARGES APPLY) X STANDARD				I. Results Only II. Results + QC Summaries (LCS, DUP, MS/MSD as required) III. Results + QC and Calibration Summaries IV. Data Validation Report with Raw Data				P.O. # <u>210799</u> Bill to:									
3.9				REQUESTED FAX DATE N/A				REQUESTED REPORT DATE N/A				Edata Yes No									
Received By Signature		Relinquished By Signature		Received By Signature		Relinquished By Signature		Received By Signature		Relinquished By Signature		Received By Signature									
Printed Name		Printed Name		Printed Name		Printed Name		Printed Name		Printed Name		Printed Name									
Firm		Firm		Firm		Firm		Firm		Firm		Firm									
Date/Time		Date/Time		Date/Time		Date/Time		Date/Time		Date/Time		Date/Time									

Received By
[Signature]
 Name
 Kelly A McDonald
 Date
 11/4/20

Date of Request: 05-OCT-2020
 Created By: PARRISH_J On: 05-OCT-2020
 Modified By: JASROTIA_P On: 07-OCT-2020
 Customer: NE-DIST
 Project: SMP
 Division: Division of Environmental Assessment and Restoration
 District: Northeast District
 Event Description: Nassau SCI's

Send Coolers To:

Phone: 904-256-1700
 8800 Baymeadows Way West
 Suite 100
 Jacksonville, FL 32256
 Attn: Jeremy Parrish
 Cooler Ship Email: Jeremy.M.Parrish@dep.state.fl.us

Priority: 3
 Event Status: R
 Criminal Investigation: NO
 Chemistry Request Reviewed By: WATTS_J On: 06-OCT-2020
 Biology Request Reviewed By: JASROTIA_P On: 07-OCT-2020
 Sampling Kit Required: YES Ship on: 23-OCT-2020
 Sampling Kit Shipped: YES On: 20-OCT-2020
 Sampling Kit Packed By: KLUG_K
 Preservatives Needed: NO
 Date to Receive Samples: 02-NOV-2020
 Logged In By: REYNOLDS_T On: 05-NOV-2020

Send Final Report To:

8800 Baymeadows Way West
 Suite 100
 Jacksonville, FL 32256
 Attn: Jeremy Parrish
 Report Notification Email: Jeremy.M.Parrish@dep.state.fl.us

Comments: Group A: Nutrients, Metals, Tracers, Q-PSNP-TQ, Bacteria
 Group B: SCI, Algae_ID

NE-ALS-ECQ

Packing Notes:
 none

Bottle Group A (Water) With 4 Samples:

Bottle Type: P-1L	Number of Bottles: 4	Preserved With: ICE
ALKALINITY	Template: DEFAULT	(SM 2320 B-2011) Alkalinity in aqueous matrices as mg CaCO ₃ /L
TURBIDITY	Template: DEFAULT	(EPA 180.1 Rev. 2.0) Turbidity in aqueous matrices
W-CL-IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Chloride in aqueous matrices
W-COLOR	Template: DEFAULT	(SM 2120 B-2011) True Color measured at 450 nm (not valid for NPDES monitoring)
Color (true)		
W-F	Template: DEFAULT	(SM 4500 F-C-2011) Fluoride in aqueous matrices, without distillation (not valid for NPDES monitoring)
W-SO ₄ -IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Sulfate in aqueous matrices
W-TDS	Template: DEFAULT	(SM 2540 C-2011) Total Dissolved Solids in aqueous matrices
W-TSS	Template: DEFAULT	(SM 2540 D-2011) Total Suspended Solids in aqueous matrices using a maximum of 250 mL of sample
Bottle Type: BP-1L	Number of Bottles: 4	Preserved With: ICE
CHLSUITE-W	Template: DEFAULT	(SM 10200 H (mod.)) Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry
Bottle Type: P-250ML-WI-THI	Number of Bottles: 4	Preserved With: ICE
NE-ALS-ECQ	Template: DEFAULT	(SM 9223 B Quanti-Tray) E. coli by Colilert/QT by ALS in Jacksonville
Bottle Type: BG-500ML	Number of Bottles: 4	Preserved With: ICE
W-E8321-DI	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS
W-E8321-MS	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS
Bottle Type: P-500ML	Number of Bottles: 4	Preserved With: HNO ₃
W-HARD	Template: DEFAULT	(SM 2340B) Hardness by calculation, in aqueous matrices, reported as mg/L CaCO ₃
Hardness		
W-ICP	Template:	(EPA 200.7 Rev. 4.4) Total Recoverable Metals analysis using ICP emission spectroscopy for aqueous samples supporting Clean Water Act Projects

Bottle Group A (Water) With 4 Samples:

Bottle Type: P-500ML	Number of Bottles: 4	Preserved With: HNO3
W-ICP	Template:	(EPA 200.7 Rev. 4.4) Total Recoverable Metals analysis using ICP emission spectroscopy for aqueous samples supporting Clean Water Act Projects
Barium		
Calcium		
Iron		
Magnesium		
Potassium		
Sodium		
Zinc		
W-ICPMS	Template:	(EPA 200.8 Rev. 5.4) Total Recoverable Metals analysis using ICP-MS for aqueous samples supporting Clean Water Act Projects
*Boron		
Aluminum		
Antimony		
Arsenic		
Beryllium		
Cadmium		
Chromium		
Copper		
Lead		
Nickel		
Selenium		
Silver		
Thallium		

Bottle Type: P-500ML	Number of Bottles: 4	Preserved With: ICE And H2SO4
W-NH3	Template: DEFAULT	(EPA 350.1 Rev. 2.0) Ammonia in aqueous matrices as mg N/L
W-NO2NO3	Template: DEFAULT	(EPA 353.2 Rev. 2.0) Nitrite/Nitrate in aqueous matrices as mg N/L
W-S-A-TP	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Total Phosphorus in aqueous matrices as mg P/L
W-TKN	Template: DEFAULT	(EPA 351.2 Rev. 2.0) Total Kjeldahl Nitrogen in aqueous matrices
W-TOC	Template: DEFAULT	(SM 5310 B-2011) Nonpurgeable Organic Carbon in aqueous matrices
Bottle Type: P-125ML	Number of Bottles: 4	Preserved With: FILTER-ICE
W-PO4-F	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Ortho-phosphate, dissolved, in filtered, aqueous matrices as mg P/L
Bottle Type: BG-1L	Number of Bottles: 10	Preserved With: ICE
W-PSNP-TQ	Template: MASTERLIST	(EPA 8270E) Organonitrogen and phosphorus pesticides in water matrices by GC/MS/MS

Bottle Group B (Biological) With 4 Samples:

Bottle Type: PJ-2L	Number of Bottles: 8	Preserved With: Formalin
MI-FW-QLDC	Template: DEFAULT	(SOP-IZ06) No. Taxa of FW macroinverts, qual samp, 20 dipnets.
Bottle Type: PT-50ML	Number of Bottles: 4	Preserved With: ICE
ALGAL_ID	Template: DEFAULT	(SOP-AB05) Qualitative assessment of algal taxa present without counts

* - The laboratory is not NELAP certified for this analyte/method, or certification is not applicable.