

Login Checklist

RQ- 2020- 11 - 30 - 38

Login Station ID: # 3

Shipping Method: FedEx | UPS | HD | Greyhound

Date/Time of Receipt: 12-01-2020 / 09:35

*Cooler Temperature	Samples Frozen?	* All Samples in Cooler Preserved with		Number of Sample Containers in Cooler	Evidence Tape				Waybill Tracking #
					Present?		*Intact?		
	YES	HNO ₃	Formalin		Yes	No	Yes	No	
3.1C				9		x			9293 8006 8749

COOLER CHECK

*All samples received on ice unless otherwise noted. HNO₃ and Formalin preserved samples do NOT have a temperature requirement.

- If the temperature of a cooler exceeds 6.0 oC (above 10.0 oC for W-1-4-DIOX and microbiology) or received without ice, identify affected samples in an NCR. NCR# _____
- DOH coolers can exceed 2.0 °C above the standard temperature (6.0 or 10.0 °C for W-1-4-DIOX) without login confirmation from customer for samples requiring ice preservation; identify affected samples in an NCR. NCR# _____
- If cooler or samples are received with a damaged Evidence Seal; identify the affected samples in an NCR. NCR# _____
- If coolers or samples are received late; identify the affected samples in an NCR. NCR# _____

Chain of Custody/ Field Sheet(s) Included?

Yes X No _____

Micro-Biology Overflow Chain of Custody/ Field Sheet(s) Included?

Yes _____ No X

CONTAINER CHECK

Evidence Tape on Bottles?

Yes _____ No _____ NA X

Evidence Tape intact? Yes _____ No _____

If Criminal, photographs taken?

Yes _____ No _____

Containers intact? Yes X No _____

Caps on tight?

Yes X No _____

Sufficient Sample Volume:

Yes X No _____

If NO is checked above, generate an NCR listing affected sample IDs in its appropriate category. NCR# _____

CHLORINE CHECK REQUIRED? (Blue dot on container)

Yes _____ No X Init: ABS

Chlorine detected? (One container checked per Field ID)

Yes _____ No _____ Init: ABS

If chlorine is detected, generate an NCR listing affected sample IDs . NCR# _____

PRESERVATION CHECK (EXCLUDING SAMPLES SENT TO OVERFLOW LABS, "OV-")

**Acid Preserved Samples: pH < 2.0?

Yes X No _____ NA _____ Init: ABS

**W-1-4-DIOX (Green dot on container) preserved to pH < 4.0?

Yes _____ No _____ NA X Init: ABS

If samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

Coolers Unpacked/Checked by: ABS

Date: 12-01-2020

Event contains NCRs (Y/N)? N

Event ID:

NE - DIST - 2020 - 12 - 01 - 0

Login Checklist

Login Preservation Equipment

Lot # & Exp: Date

Additional Comments:

Infrared Thermometer ID:	IRTG # 3 EXP:10/28/2021
pH Test Strips 0.0 – 6.0	213316 BV EXP:05/15/21
pH Paper 0.0 – 3.0	XX XX XX XX
pH Paper 0.0 – 13.0	231019 EXP:11/01/2022
Chlorine Test Strips 0.-10.0	0129 / 2023 / 03

ENCORE SAMPLES

S-VOC-MS samples in Encores must be frozen within 48 hours of collection.

Were Encore samples received? Yes _____ Init: ABS

Date and time placed in freezer _____

Were all samples placed in freezer within 48 hours of collection? Yes _____ No _____

If no, were samples shipped on dry ice? Yes _____ No _____

If S-VOC-MS samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

FOR CLEAN LAB USE ONLY – Total Mercury Preservation Check(W-HG-AF;W-HG-AF-F):

Samples preserved within 48 hours? Yes _____ No _____

Date and time samples preserved: _____

FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION
CHAIN OF CUSTODY FORM

Date: 11/30/2020
Customer: NE-DIST
RQ: RQ-2020-11-30-38
Collected By: Jeremy Parrish, Brian Davis
Sampling Agency: FDEP
Project ID: SMP

PLEASE RETURN THE ORIGINAL OF THIS FORM WITH SAMPLE SUBMITTAL
FORMS AND THE COOLER PACKING WORKSHEET TO THE LAB

Number of Coolers Shipped: 1

FIELD ID (Labels) submitted under this RQ for this collection date.

Site Location	Field ID/WIN ID
Mills Creek At Us 301/us1	 19020048
Unnamed Branch Above Alligator Cr.	 G4NE0288
Cushing Cr @ Stratton Rd.	 NE400002

RELINQUISHED BY(SIGNATURE):

Jeremy Parrish

DATE/TIME:

11/30/20

THIS SECTION TO BE COMPLETED BY LABORATORY

Rec'd/Inspected By: **A.SHAIK**

Rec'd/Inspected Date: **12-01-2020**



RQ-2020-11-30-38

Customer: WQAP

Project ID: SMP

Collected By: Jeremy Parrish, Brian Davis

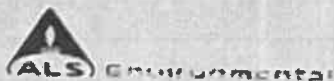
Field Report Prepared By: Jeremy Parrish

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to contract laboratory				
WIN ID/Field ID:										
		NE400002								
Location:		Cushing Cr @ Stratton Rd.								
Matrix: Fresh						(Check one) ☒ GRAB ☐ COMPOSITE				
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp COND (µmhos/cm)	Salinity (PPT _h)
11/30/2020	12:00 EST	4.55	49	18.8	7.1	0.3	0.5 ☐ VOB (S)	0.6	224.6	0.11
Central Lab please use top row for login purposes										
	EST									
Parameter Suite		Parameter Methods				Preservation		pH Check	# Bottles sent to Lab	Bottle Group
Preserved Nutrients (P-500ML)		☒ W-NH3 / W-NO2NO3 / W-TKN / W-TOC / W-S-A-TP				☒ Ice ☒ H2SO4*		☒ <2	1	A
		Lot # Sa0038110		Exp: 2/7/21						
Metals (P-500ML)		☐ W-ICPMS ☐ W-ICP ☐ W-HARD				☐ Ice ☐ HNO3*		☐ <2		
		Lot #		Exp:						
Filtered Nutrients (P125ML)		☒ W-P04-F ☒ Field Filtered 0.45 µm Filter				☒ Ice			1	A
		Filter Lot # R9na98003								
		Syringe Lot #								
Chlorophyll (BP-1L)		☒ CHLSUITE-W				☒ Ice			1	A
Physical/Aggregate (Fresh) (P-1L)		☒ Alkalinity / W-F / W-TSS / TURBIDITY / W-CH-IC / W-COLOR / W-SO4-IC / W-TDS				☒ Ice			1	A
Physical/Aggregate (Marine) (P-1L)		☐ Alkalinity / W-BR-IC / W-F / W-TSS / TURBIDITY				☐ Ice				
Macroinvertebrates (PJ-2L)		☐ MI-FW-QLDC				☐ Formalin				
Periphyton (PT-50ML)		☐ ALGAE-ID				☐ Ice				
Microbiology (Contract Lab Bottle)		☒ E Coli ☐ F Coli ☐ Enterococci ☒ Contract Lab				☒ Ice			1	A
Molecular (QPCR-P-500ML)		☐ PCR-HF183 ☐ PCR-BacR ☐ PCR-DG3 ☐ PCR-GULL2 ☐ PCR-GFD ☐				☐ Ice				
Tracers (BG-500ML)		☐ W-E8321-DI ☐ W-E8321-MS				☐ Ice				
Pesticides (BG-500ML) (BG-1L)		☐ W-E8321-DI ☐ W-E8321-MS ☐ W-PSNP-TQ ☐ W-CT-CL-R ☐ W-PCL-SQ-R ☐ W-CARB-AA (5 ml of MCAA** Buffer Solution)				☐ Ice				
						☐ Ice				
Name: Jeremy Parrish Brian Davis				Signature: 				Date: 11/30/20		

*2.0 ml of Preservative used unless noted otherwise.

**MCAA: Monochloroacetic acid buffer solution



CHAIN OF CUSTODY / LABORATORY ANALYSIS REQUEST FORM

9143 Phillips Highway, Suite 200, Jacksonville, FL 32256
Phone (904) 739-2277 / 800-695-7222 x06 / FAX (904) 739-2011

www.alsglobal.com

SR #

ALS Contact

Page ____ of ____

Project Name Nassau Mckay		Project Number 2020-11-30-38		ANALYSIS REQUESTED (Include Method Number and Container Preservative)															
Report To 7EP-OverFlow Micro @ day state		Report CC Jeremy Pellish		Preservative															
Company/Address 8800 Bynedon Way W suite 100, Jacksonville FL 32256		Phone # 904-256-1700		FAX #		NUMBER OF CONTAINERS Each													
Sampler's Signature B-P		Sampler's Printed Name Brian Davis		Preservative Key															
CLIENT SAMPLE ID		LAB ID		SAMPLING DATE TIME		Matrix		REMARKS											
NE400002				11/30/20 1200		SW		FLOSL											
19020048				↓ 940		↓		↓											
64NE0288				↓ 1030		↓		↓											
Special Instructions/Comments:																			
TURNAROUND REQUIREMENTS																			
RUSH (SURCHARGES APPLY) ☒ STANDARD																			
REQUESTED FAX DATE																			
REQUESTED REPORT DATE																			
REPORT REQUIREMENTS																			
I. Results Only																			
II. Results + QC Summaries																			
III. Results + QC and Calibration Summaries																			
IV. Data Validation Report with Raw Data																			
Edata Yes No																			
INVOICE INFORMATION																			
P.O. #																			
Bill to:																			
Signature																			
Printed Name																			
Firm																			
Date/Time																			
SAMPLE RECEIPT: CONDITION/COOLER TEMP.				CUSTODY SEALS: Y N				Relinquished By				Received By							
Signature B-P				Signature Brian Davis				Signature Valerie Spears				Signature ALS							
Printed Name Brian Davis				Printed Name Valerie Spears				Printed Name ALS				Printed Name							
Firm FDEP DEAR				Firm ALS				Firm				Firm							
Date/Time 11/30/20 1302				Date/Time 11/30/20 1302				Date/Time				Date/Time							



RQ-2020-11-30-38

Customer: WQAP

Project ID: SMP

Collected By: Jeremy Parrish, Brian Davis

Field Report Prepared By: Jeremy Parrish

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to contract laboratory				
WIN ID/Field ID:  G4NE0288										
Location: Unnamed Branch Above Alligator Cr.										
Matrix: Fresh						(Check one) ☒ GRAB ☐ COMPOSITE				
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp COND (µmhos/cm)	Salinity (PPT _h)
11/30/2020	10:30 EST	4.81	51.6	18.7	7.17	0.3	0.4 ☐ VOB (S)	0.5	326	0.16
Central Lab please use top row for login purposes										
EST										
Parameter Suite		Parameter Methods				Preservation		pH Check	# Bottles sent to Lab	Bottle Group
Preserved Nutrients (P-500ML)		☒ W-NH3 / W-NO2NO3 / W-TKN / W-TOC / W-S-A-TP				☒ Ice ☒ H2SO4*		☒ <2	1	B
		Lot # Sa0038110		Exp: 2/7/21						
Metals (P-500ML)		☒ W-ICPMS ☒ W-ICP ☒ W-HARD				☒ Ice ☒ HNO3*		☒ <2	1	B
		Lot # Sa9354060		Exp: 12/20/20						
Filtered Nutrients (P125ML)		☒ W-PO4-F ☒ Field Filtered 0.45 µm Filter				☒ Ice			1	B
		Filter Lot # Rna98003								
		Syringe Lot #								
Chlorophyll (BP-1L)		☒ CHLSUITE-W				☒ Ice			1	B
Physical/Aggregate (Fresh) (P-1L)		☒ Alkalinity / W-F / W-TSS / TURBIDITY / W-CH-IC / W-COLOR / W-SO4-IC / W-TDS				☒ Ice			1	B
Physical/Aggregate (Marine) (P-1L)		☐ Alkalinity / W-BR-IC / W-F / W-TSS / TURBIDITY				☐ Ice				
Macroinvertebrates (PJ-2L)		☐ MI-FW-QLDC				☐ Formalin				
Periphyton (PT-50ML)		☐ ALGAE-ID				☐ Ice				
Microbiology (Contract Lab Bottle)		☒ E Coli ☐ F Coli ☐ Enterococci ☒ Contract Lab				☒ Ice			1	B
Molecular (QPCR-P-500ML)		☐ PCR-HF183 ☐ PCR-BacR ☐ PCR-DG3 ☐ PCR-GULL2 ☐ PCR-GFD ☐				☐ Ice				
Tracers (BG-500ML)		☐ W-E8321-DI ☐ W-E8321-MS				☐ Ice				
Pesticides (BG-500ML) (BG-1L)		☐ W-E8321-DI ☐ W-E8321-MS ☐ W-PSNP-TQ ☐ W-CT-CL-R ☐ W-PCL-SQ-R ☐ W-CARB-AA (5 ml of MCAA** Buffer Solution)				☐ Ice				
						☐ Ice				
Name: Jeremy Parrish Brian Davis				Signature: 				Date: 11/30/20		

*2.0 ml of Preservative used unless noted otherwise.

**MCAA: Monochloroacetic acid buffer solution



FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION STRATEGIC MONITORING PROGRAM FIELD SHEET

January 2020

Meter Passed End

of Day CCV

Yes / No

WIN Station Name: Cushing Cr @ Stratton Rd.

Date: 11/30/2020

WIN/Field ID: NE400002

WBID: 2162

ENR:

Latitude: 30.5682

County: NASSAU

Org ID: 21FLA

Longitude: -81.8212

Receiving Body of Water: Nassau - St. Marys

RQ: 2020-11-30-38

Sampling Team Members		WQ Measurements	Sample Collection	Documentation	Preservation	Preservation Check
☒	Jeremy Parrish	☒	☒	☒	☒	☒
☒	Brian Davis	☒	☒	☒	☒	☒
☐		☐	☐	☐	☐	☐
☐		☐	☐	☐	☐	☐

Sampling Equipment	
☐ Sample Container	☐ Carboy
☐ Dipper	☐ Van Dorn
☐ Syringe	☒ Pole
Depth measured with: Meter	
Secchi Equipment ID:	
Collection Method	
☐ Wading	☐ Canoe/Kayak
☒ From Shore	☐ Electric Motor
☐ Other	☐ Gasoline Motor

Water Level: ☒ Normal ☐ Low ☐ High ☐ Dry ☐ Pooled ☐ Flooded		Meter ID# DESI	
Photos: No	Flow: FLOW	Tide:	Tide Times: High: Low:
Biological Samples Collected: ☒ None ☐ HA ☐ SCI ☐ RPS ☐ Algae ID ☐ LVS ☐ LVI Other:			
SBIO ID Numbers:			
Notes:			

Duplicate Sample

Time Zone: EST	Time: N/A	Field ID: N/A	(WIN ID_DUP)
WIN DMT Activity ID:			

Blank

Time Zone:	Time: N/A	Field ID: N/A	(Blank Type_DDMYYYY)
DI Source: N/A	Location: N/A		
DI Type: N/A	Equipment ID: N/A		

LIMS EVENT ID: WIN Import ID:

Enter Field Parameters, Verify Station ID In LIMS DMT: (Initials/Date) QC Station ID, Field Parameters In LIMS DMT: (Initials/Date)

Scan/Save Field Sheets (Initials/Date) Migrate Data to WIN: (Initials/Date) Verify Data In WIN: (Initials/Date)



RQ-2020-11-30-38

Customer: WQAP

Project ID: SMP

Collected By: Jeremy Parrish, Brian Davis

Field Report Prepared By: Jeremy Parrish

Sampling Agency: FDEP

Place Label Here						Comments: Microbiology sample(s) submitted to contract laboratory				
WIN ID/Field ID:										
		19020048								
Location:		Mills Creek At Us 301/us1								
Matrix: Fresh						(Check one) ☒ GRAB ☐ COMPOSITE				
Date	Time	DO (mg/L)	DO (%SAT)	Temp. (C°)	pH	Sample Depth (m)	Secchi Disk (m)	Total Depth (m)	Sp COND (µmhos/cm)	Salinity (PPT)
11/30/2020	9:40 EST	2.97	32	18.8	5.6	0.1	0.15 ☒ VOB (S)	0.15	88.5	0.04
Central Lab please use top row for login purposes										
	EST									
Parameter Suite		Parameter Methods				Preservation		pH Check	# Bottles sent to Lab	Bottle Group
Preserved Nutrients (P-500ML)		☐ W-NH3 / W-NO2NO3 / W-TKN / W-TOC / W-S-A-TP				☐ Ice ☐ H2SO4*		☐ <2		
		Lot #			Exp:					
Metals (P-500ML)		☐ W-ICPMS ☐ W-ICP ☐ W-HARD				☐ Ice ☐ HNO3*		☐ <2		
		Lot #			Exp:					
Filtered Nutrients (P125ML)		☐ W-P04-F ☐ Field Filtered 0.45 µm Filter				☐ Ice				
		Filter Lot #								
		Syringe Lot #								
Chlorophyll (BP-1L)		☐ CHLSUITE-W				☐ Ice				
Physical/Aggregate (Fresh) (P-1L)		☐ Alkalinity / W-F / W-TSS / TURBIDITY / W-CH-C / W-COLOR / W-SO4-IC / W-TDS				☐ Ice				
Physical/Aggregate (Marine) (P-1L)		☐ Alkalinity / W-BR-IC / W-F / W-TSS / TURBIDITY				☐ Ice				
Macroinvertebrates (PJ-2L)		☐ MI-FW-QLDC				☐ Formalin				
Periphyton (PT-50ML)		☐ ALGAE-ID				☐ Ice				
Microbiology (Contract Lab Bottle)		☒ E Coli ☐ F Coli ☐ Enterococci ☒ Contract Lab				☒ Ice			1	C
Molecular (QPCR-P-500ML)		☐ PCR-HF183 ☐ PCR-BacR ☐ PCR-DG3 ☐ PCR-GULL2 ☐ PCR-GFD ☐				☐ Ice				
Tracers (BG-500ML)		☐ W-E8321-DI ☐ W-E8321-MS				☐ Ice				
Pesticides (BG-500ML) (BG-1L)		☐ W-E8321-DI ☐ WE8321-MS ☐ W-PSNP-TQ ☐ W-CT-CL-R ☐ W-PCL-SQ-R ☐ W-CARB-AA (5 ml of MCAA** Buffer Solution)				☐ Ice				
						☐ Ice				
Name: Jeremy Parrish Brian Davis				Signature: <i>Jeremy Parrish</i>				Date: 11/30/20		

*2.0 ml of Preservative used unless noted otherwise.

**MCAA: Monochloroacetic acid buffer solution



FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION STRATEGIC MONITORING PROGRAM FIELD SHEET

January 2020

Meter Passed End
of Day CCV
☒ Yes / ☐ No

WIN Station Name: Unnamed Branch Above Alligator Cr. **Date:** 11/30/2020
WIN/Field ID: G4NE0288 **WBID:** 2156 **ENR:** **Latitude:** 30.5767
County: NASSAU **Org ID:** 21FLA **Longitude:** -81.8245
Receiving Body of Water: Nassau - St. Marys **RQ-** 2020-11-30-38

Sampling Team Members		WQ Measurements	Sample Collection	Documentation	Preservation	Preservation Check
☒	Jeremy Parrish	☒	☒	☒	☒	☒
☒	Brian Davis	☒	☒	☒	☒	☒
☐		☐	☐	☐	☐	☐
☐		☐	☐	☐	☐	☐

Sampling Equipment	
☒ Sample Container	☐ Carboy
☐ Dipper	☐ Van Dorn
☐ Syringe	☐ Pole
Depth measured with: Meter	
Secchi Equipment ID: <u></u>	
Collection Method	
☒ Wading	☐ Canoe/Kayak
☐ From Shore	☐ Electric Motor
☐ Other <u></u>	☐ Gasoline Motor

Water Level: ☒ Normal ☐ Low ☐ High ☐ Dry ☐ Pooled ☐ Flooded		Meter ID# <u>DESI</u>	
Photos: <u>No</u>	Flow: <u>FLOW</u>	Tide: <u>N/A</u>	Tide Times: High: <u></u> Low: <u></u>
Biological Samples Collected: ☐ None ☒ HA ☐ SCI ☒ RPS ☐ Algae ID ☒ LVS ☐ LVI Other: <u></u>			
S BIO ID Numbers: <u></u>			
Notes: 			

Duplicate Sample	
Time Zone: <u>EST</u> Time: <u>N/A</u> Field ID: <u>N/A</u> (WIN ID_DUP)	
WIN DMT Activity ID: <u></u>	

Blank	
Time Zone: <u></u> Time: <u>N/A</u> Field ID: <u>N/A</u> (Blank Type_DDMMYYYY)	
DI Source: <u>N/A</u> Location: <u>N/A</u>	
DI Type: <u>N/A</u> Equipment ID: <u>N/A</u>	

LIMS EVENT ID: **WIN Import ID:**
Enter Field Parameters, Verify Station ID in LIMS DMT: (Initials/Date) **QC Station ID, Field Parameters in LIMS DMT:** (Initials/Date)
Scan/Save Field Sheets (Initials/Date) **Migrate Data to WIN:** (Initials/Date) **Verify Data in WIN:** (Initials/Date)

CALIBRATION AND VERIFICATION LOG (FDEP SOP FT 1000-FT 1500, FD 1000-FD 4000)

Boldly "X" this box if there are qualified data on this page.

Meter ID: Desi RQ- 2020-11-20-38 Project: Nassau Mckay

- Notes: (1) Always wait for meter to stabilize before recording any readings.
 (2) Report all digits displayed. Do not round before reporting measurements. (See special instructions for depth).
 (3) For Calibrations, record calibrated meter reading. Do not record initial meter reading before calibration.

Temperature (Quarterly) FT 1400

Date of Last Temperature Verification 10/14/20

DO DEP SOP FT 1500	Name	Date	Time CT-ET	Temp. °C	Baro- meter mmHg	D.O. Chart mg/L	Meter D.O. mg/L	% DO	Probe Charge	Probe Gain	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/30/20	812	22.8	755.5	8.6	✓	99.4	✓	1.06	(P) F	(L) F
ICV			814	22.8	755.4	8.6	8.56	99.5	✓	1.06	(P) F	(L) F
CCV			1328	19.0	755.8	9.2	9.31	100.4	✓	1.06	(P) F	(L) F
CCV											P / F	L / F

DO Acceptance criteria from Table ± 0.3 mg/L.

Rapid-Pulse Sensors: DO Gain Range 0.7 to 1.4; DO Charge Range 25-75.

Optical: DO gain range 0.85 to 1.15 (Pro DSS 0.75 to 1.50); DO charge N/A. Steady-state & Galvanic Sensors: DO Gain & Charge N/A.

Spec. Cond. FT 1200	Name	Date	Time CT-ET	Lot #	Expir. Date	Standard μ hos/cm	Meter Reading μ hos/cm	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/10/20	819	200902B	9/21	1000	1000	(P) F	(L) F
ICV			820	200902A	9/21	100	102.7	(P) F	(L) F
CCV			1331	200902B	9/21	1000	997	(P) F	(L) F
CCV								P / F	L / F

Conductivity Acceptance criteria $\pm 5\%$

pH DEP SOP FT 1100	Name	Date	Time CT-ET	Lot #	Expir. Date	pH Buffer SU	Temp °C	Meter Reading SU	mV	Pass / Fail	Lab / Field
Calibr.	Brian Davis	11/30/20	824	2001C31	1/22	7.01	23.2	7.04	-25.1	(P) F	(L) F
Calibr.			826	2006B06	5/22	4.0	23.2	4.0	17.6	(P) F	(L) F
Calibr.			828	2002911	8/21	10.02	23.2	10.02	78.7	(P) F	(L) F
ICV			830	2001C31	1/22	7.01	23.2	7.04	-25.9	(P) F	(L) F
CCV			1332	2002911	8/21	10.02	23.2	10.15	-26.6	(P) F	(L) F
CCV										P / F	L / F

pH Acceptance criteria ± 0.2 SU; mV pH 7 Range 0 ± 50 ; mV pH 4 Range $+180 \pm 50$; mV pH 10 Range -180 ± 50 ;

If mV are recorded: slope from 7 to 10 _____, slope from 4 to 7 _____ (both must be between 165 and 180 mV)

Does meter have a depth sensor that will be used to measure total depth & sample depth? YES / NO / NA (not surf. water project)

If YES, complete daily Calibr. & ICV below and list date of last quarterly depth verification: _____

If NO, what will be used? (circle one) Secchi Disk Line / Sonar Unique ID: _____; Date of last verification: _____

Depth Sensor (Daily Calibration & ICV)	Name	Date	Time CT-ET	Calibrated Value (0.00 or Offset), meters	ICV Value, meters	Pass / Fail	Lab / Field
Pressure mode in air	Brian Davis	11/30/20	832	0.00	0.00	(P) F	(L) F

Report two decimal places. Round numbers ≤ 4 down, ≥ 5 up. ICV acceptance criteria $\pm 5\%$ or ± 0.05 m, whichever is greater.

COMMENTS:



FLORIDA DEPARTMENT OF ENVIRONMENTAL PROTECTION STRATEGIC MONITORING PROGRAM FIELD SHEET

January 2020

Meter Passed End

of Day CCV

Yes / No

WIN Station Name: Mills Creek At Us 301/us1

Date: 11/30/2020

WIN/Field ID: 19020048

WBID: 2120B

ENR:

Latitude: 30.6389

County: NASSAU

Org ID: 21FLA

Longitude: -81.8631

Receiving Body of Water: Nassau - St. Marys

RQ- 2020-11-30-38

Sampling Team Members		WQ Measurements	Sample Collection	Documentation	Preservation	Preservation Check
☒	Jeremy Parrish	☒	☒			
☒	Brian Davis		☒			
☐						
☐						

Sampling Equipment	
☒ Sample Container	☐ Carboy
☐ Dipper	☐ Van Dorn
☐ Syringe	☐ Pole
Depth measured with: Meter	
Secchi Equipment ID:	
Collection Method	
☐ Wading	☐ Canoe/Kayak
☒ From Shore	☐ Electric Motor
☐ Other	☐ Gasoline Motor

Water Level: ☒ Normal ☐ Low ☐ High ☐ Dry ☐ Pooled ☐ Flooded		Meter ID# DESI	
Photos: NO	Flow: FLOW	Tide:	Tide Times: High: Low:
Biological Samples Collected: ☒ None ☐ HA ☐ SCI ☐ RPS ☐ Algae ID ☐ LVS ☐ LVI Other:			
SBIO ID Numbers:			
Notes:			

Duplicate Sample

Time Zone: EST	Time: N/A	Field ID: N/A	(WIN ID_DUP)
WIN DMT Activity ID:			

Blank

Time Zone:	Time: N/A	Field ID: N/A	(Blank Type_DDMMYYYY)
DI Source: N/A	Location: N/A		
DI Type: N/A	Equipment ID: N/A		

LIMS EVENT ID: WIN Import ID:

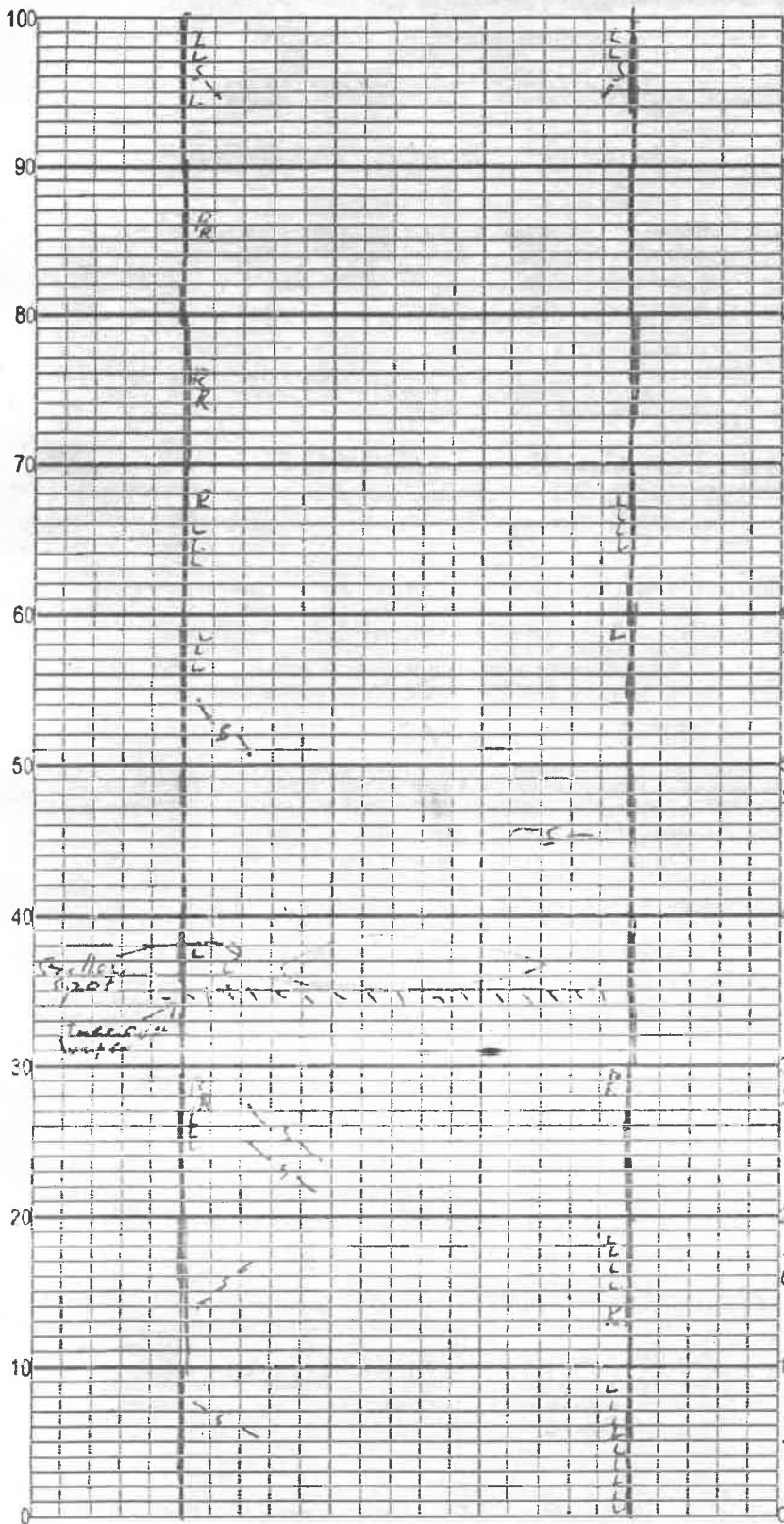
Enter Field Parameters, Verify Station ID In LIMS DMT: (Initials/Date) QC Station ID, Field Parameters In LIMS DMT: (Initials/Date)

Scan/Save Field Sheets (Initials/Date) Migrate Data to WIN: (Initials/Date) Verify Data In WIN: (Initials/Date)

DEP Form FD 9000-4 Stream/River Habitat Assessment Sketch Sheet



Left Bank



S=0.5
L=0.4

Location: Unshaded Branch Above Alligator Creek

Date: 11/30/20

Sampler: Brian Davis

R=0.2

100 m: Latitude _____

Longitude _____

0 m: Latitude _____

Longitude _____

Substrates: Code key, draw proportionate habitat abundance.

☐ Snags <1

☐ Roots/undercut banks <1

☐ Leaf Packs (or mats) <1

☐ Aquatic Macrophytes _____

☐ _____

☐ _____

☐ Pools total # observed = 0
range expected = 2

Average width 4 m

Average depth 0.5 m

Velocity measured at 10/0 meter mark.

WQ & chemistry taken at 0 meter mark.

Habitat Smothering.

Note areas (on map) where sand or silt is smothering substrates, limiting habitability.

Bank Stability:

Note areas (on map) with unstable, eroding banks.

Riparian Buffer Width:

Note areas (on map) where natural vegetation is altered or eliminated.

Plants observed / other notes:

S=0.2

L=0.1

R=0.15

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

S=0.2

L=0.4

L=2.0

R=0.8

S=1.5

0.1 m/s
0.5 m deep

Length of grid represents 100 m of stream (not linear meters).
(Horizontal scale is double vertical scale, draw proportionately).

DEP Form FD 9000-5 Stream/River Habitat Assessment Field Sheet

SAMPLING AGENCY: <u>FDEP DEAR</u>		STORET STATION NUMBER: <u>64NE0288</u>	DATE (MM/DD/YY): <u>11/30/20</u>	RECEIVING BODY OF WATER: <u>Aligator Creek</u>
REMARKS:	COUNTY: <u>Nassau</u>	LOCATION: <u>Ungrazed Branch above Aligator Creek</u>		FIELD ID/NAME:

Habitat Parameter	Optimal	Suboptimal	Marginal	Poor
Primary Habitat Components	Four or more major productive habitats present (snags, tree roots, aquatic vegetation, leaf packs (partially decayed), rock)	Three major productive habitats present. Adequate habitat. Some substrates may be new fall (fresh leaves or snags)	Two major productive habitats present. Less than desirable habitat, frequently disturbed or removed.	One or less major productive habitat. Lack of habitat is obvious, substrates unstable or smothered
Substrate Diversity <u>4</u>	20 19 18 17 16	15 14 13 12 11	10 9 8 7 6	5 <u>4</u> 3 2 1
Substrate Availability <u>4</u>	Greater than 30% major productive habitat present at site 20 19 18 17 16	16% to 30% major productive habitat, by aerial extent 15 14 13 12 11	6% to 15% major productive habitat 10 9 8 7 6	Less than 5% major productive habitat 5 <u>4</u> 3 2 1
Water Velocity <u>10</u>	Max. observed at typical transect: > 0.25 m/sec. But < 1 m/sec ≥0.33 0.31 0.29 0.27 >0.25 20 19 18 17 16	Max. observed at typical transect: >0.1 to 0.25 m/sec 0.25 0.21 0.17 0.13 >0.1 15 14 13 12 11	Max. observed at typical transect: 0.05 to 0.1 m/sec 0.1 0.09 0.07 0.06 0.05 <u>10</u> 9 8 7 6	Max. observed at typical transect: <0.05 m/sec; or spate occurring: > 1 m/sec <0.05 0.04 0.03 0.01 <0.01 5 4 3 2 1
Habitat Smothering <u>4</u> Primary Score <u>22</u>	Adequate number of stable pools (1-2 per 12 times width) and <25% of habitats affected by sand, silt, or algae 20 19 18 17 16	Adequate number of stable pools (1-2 per 12 times width) and >25% of habitats affected by sand, silt, or algae. 15 14 13 12 11	Does not have required number of stable pools (1-2 per 12 x width) and/or has shallow pools (<2 x prevailing depth). 10 9 8 7 6	Stable pools are absent. Most habitats affected by sand, silt, or algae accumulation. 5 4 3 2 1
Secondary Habitat Components Artificial Channelization <u>6</u>	Expected sinuosity given the stream width. No evidence of dredging or artificial straightening. No spoil banks. 20 19 18 17 16	Good sinuosity within old channelized area. Evidence of dredging or straightening in the past (>25 yrs) but mostly recovered. 15 14 13 12 11	Straightened with trapezoidal cross section, but has a small degree of sinuosity developed within channelized area. 10 9 8 7 <u>6</u>	Straightened or engineered by dredging, has trapezoidal or box cut cross section, lacks required pools. May have spoil banks. 5 4 3 2 1
Bank Stability Right Bank <u>5</u> Left Bank <u>5</u>	Bankfull > 60% of bank height. Slope of bank ≤ 60° from bankfull to top of bank. Bankfull is within or above the woody root zone with few raw, eroded areas. 10 9	Only meets 2 of the 3 requirements for optimal bank stability 8 7 6	Only meets 1 of the 3 requirements for optimal bank stability. 5 4	Bankfull < 60% of bank height. Slope of bank > 60°. Bankfull is below the woody root zone with raw, eroded areas. 3 2 1
Riparian Buffer Zone Width Right Bank <u>10</u> Left Bank <u>10</u>	Width of vegetation greater than 18 m <u>10</u> 9	Width of vegetation >12 to 18m 8 7 6	Width of vegetation 6 to 12 m human activities close to system 5 4	Less than 6 m of buffer zone due to intensive human activities 3 2 1
Riparian Zone Vegetation Quality Right Bank <u>6</u> Left Bank <u>6</u> Secondary Score <u>50</u>	Over 80% of riparian surfaces consist of normal, expected plant community for given sunlight & habitat conditions (e.g., native plants; tree, shrub, and forbs represented, if appropriate). Minimal disturbance. 10 9	>50% to 80% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). Some disruption in community observed. <u>8</u> 7 <u>6</u>	25% to 50% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). Disruption obvious. 5 4	Less than 25% of riparian zone is undisturbed (normal, expected plant community for given sunlight & habitat conditions). 3 2 1

72 TOTAL SCORE

Date: <u>11/30/20</u>	Analyst: <u>Barich Davis</u>	Signature: <u>B. Davis</u>
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DEP FORM FD 9000-32: LINEAR VEGETATION SURVEY FIELD SHEET

Waterbody: <u>Wm. Creek at Allegan</u>	Date: <u>1/30/20</u>	County: <u>Nash</u>	Store Number: <u>64AE0288</u>
Analyst: _____	Signature: <u>[Signature]</u>		

Signature: _____

Instructions: Note taxa growing in the water with a checkmark if present, "D" if dominant, and "C" if codominant (dominance only if 1 m² of taxa present). Note in the boxes below if no dominant are selected, and note total macrophyte abundance rank for each section. Only the most common taxa are listed, so use blank spaces for additional taxa names.

HERBACEOUS SPECIES	Meter Mark										Specimen collected (check)		Meter Mark										Specimen collected (check)	
	0-10	10-20	20-30	30-40	40-50	50-60	60-70	70-80	80-90	90-100			0-10	10-20	20-30	30-40	40-50	50-60	60-70	70-80	80-90	90-100		
<i>Alternanthera philoxeroides</i>													<i>Micranthemum glomeratum</i>											
<i>Bacopa caroliniana</i>													<i>Micranthemum umbrosum</i>											
<i>Bacopa monnieri</i>													<i>Myriophyllum aquaticum</i>											
<i>Bidens mitis</i>													<i>Najas guadelupensis</i>											
<i>Boehmeria cylindrica</i>													<i>Nuphar luteum</i>											
<i>Centella asiatica</i>													<i>Orontium aquaticum</i>											
<i>Ceratophyllum demersum</i>													<i>Panicum hemitomon</i>											
<i>Chara</i>													<i>Panicum repens</i>											
<i>Cicuta maculata</i>													<i>Panicum rigidulum</i>											
<i>Cladium jamaicense</i>													<i>Pistia stratioides</i>											
<i>Colocasia esculenta</i>													<i>Polygonum glabra</i>											
<i>Commelina diffusa</i>													<i>Polygonum hydropiperoides</i>											
<i>Commelina virginica</i>													<i>Polygonum punctatum</i>											
<i>Crinum americanum</i>													<i>Pontedaria cordata</i>											
<i>Diodia virginiana</i>													<i>Potamogeton illinoensis</i>											
<i>Eichhornia crassipes</i>													<i>Ruellia simplex</i>											
<i>Eleocharis (wispy viviparus)</i>													<i>Sacciolepis striata</i>											
<i>Hydrilla verticillata</i>													<i>Sagittaria kurziana</i>											
<i>Hydrocotyle</i>													<i>Sagittaria lancifolia</i>											
<i>Hygrophila polysperma</i>													<i>Sagittaria latifolia</i>											
<i>Hymenocallis</i>													<i>Salvinia minima</i>											
<i>Lachnanthes caroliniana</i>													<i>Samolus parviflora</i>											
<i>Landoltia punctata</i>													<i>Saururus cernuus</i>											
<i>Lemna</i>													<i>Sparganium americanum</i>											
<i>Limnophila sessiliflora</i>													<i>Typha</i>											
<i>Ludwigia leptocarpa</i>													<i>Vallisneria americana</i>											
<i>Ludwigia palustris</i>													<i>Zizania aquatica</i>											
<i>Ludwigia peruviana</i>																								
<i>Ludwigia repens</i>																								

DEP Form FD 9000-3 Physical/Chemical Characterization Field Sheet

SAMPLE ID _____ ORG ID 21FLA LATITUDE _____
 COUNTY Nassau STORET # 64NE0288 LONGITUDE _____
 DATE 11/30/20 TIME 1030 SAMPLING AGENCY FDEP DEARVEROC
 SITE NAME Unnamed creek abt. Alligator Creek
 FIELD ID/NAME _____ RECEIVING BODY OF WATER Allig. creek

RIPARIAN ZONE / STREAM FEATURES

PREDOMINANT LAND-USE IN WATERSHED (specify relative percent in each category): Forest/Natural ☐ Silviculture ☐ Field/Pasture ☐ Agricultural ☐ Residential ☐ Commercial ☐ Industry ☐ Other (Specify) ☐								Landscape Development Intensity Index (LDI) ☐
Local Watershed Erosion (select one): ☐ None ☒ Slight ☐ Moderate ☐ Heavy Local Watershed NPS Pollution: ☐ No evidence ☒ Slight ☐ Moderate ☐ Heavy								Typical Width (m) Depth (m)/Velocity (m/sec) Transect _____ m wide _____ m/s _____ m/s _____ m/s
Width of Riparian Vegetation (m) on Each Buffer Side Left Bank: <u>>18</u> Right Bank: <u>>18</u>				Hydrologic Modification Score (per FT 3101) _____				_____ m deep _____ m deep _____ m deep
High Water Mark: <u>0.3</u> + <u>0.5</u> = <u>0.8</u> (m) (above present water level) (present depth) (above bed)				Artificially Impounded ☐ Yes ☐ No				_____ m deep _____ m deep _____ m deep
Artificially ☐ No ☐ Mostly recovered, more sinuous Channelized: ☒ Some recovery ☒ Recent, severe				Canopy Cover % ☐ Open ☐ Lightly Shaded (11-45%) ☒ Heavily Shaded ☐ Moderate Shaded (46-80%)				

SEDIMENT / SUBSTRATE

Sediment Oils ☐ Absent ☒ Slight ☐ Moderate ☐ Profuse			Sediment Odors ☐ Normal ☐ Sewage ☐ Petroleum ☐ Chemical ☒ Anaerobic ☐ Other (Specify): _____			Smothering of Substrates Sand Smothering: None ☐ Slight ☒ Moderate ☐ Severe ☐ Silt Smothering: None ☐ Slight ☐ Moderate ☐ Severe ☒ Algae Smothering: None ☒ Slight ☐ Moderate ☐ Severe ☐		
SUBSTRATE TYPE Assessment Tool: ☐ SCI ☒ RPS ☒ LVS ☐ LCI ☐ BioRecon INVERT PERI % Coverage # Times Sampled # Times Sampled			WATER QUALITY Depth (M) Temp (°C) pH (SU) D.O. (MG/L) D.O. Sat (%) Cond. (UMHC/CM) Salinity (PPT) SECCHI (M) Top: _____ 18.7 _____ 4.55 49.0 _____ _____ 0.4 Mid: <u>0.3</u> <u>18.7</u> <u>7.17</u> <u>4.81</u> <u>51.6</u> <u>326</u> <u>0.16</u> ☐ VOB Bottom: _____ Meter ID: <u>Pesi</u> TOTAL DEPTH <u>0.5</u>					
Woody Debris (Snags) _____ Undercut Banks / Roots <u>1</u> Leaf Packs or Mat <u>1</u> Aquatic Vegetation _____ Rock or Shell Rubble _____ Sand _____ Mud / Muck / Silt <u>98</u> Other _____ Other _____			Water Odors Normal ☒ Petroleum ☐ Sewage ☐ Chemical ☐ Other ☒			Water Surface Oils Normal ☒ Sheen ☐ Clobs ☐ Slick ☐ Other ☐ Color Tannic ☒ Green (Algae) ☐ Clear ☐ Other (Specify) ☐		
ABUNDANCE Not Obs. Rare Common Abundant Periphyton: ☒ ☐ ☐ ☐ Fish: ☐ ☒ ☐ ☐ Aquatic Plants: ☒ ☐ ☐ ☐ Iron/Sulfur Bacteria: ☐ ☒ ☐ ☐			Water Sample Taken? ☒ Yes ☐ No Sample Preserved? ☒ Yes ☐ No Lot Number: _____ Exp: _____ Algae Sample Taken? ☐ Yes ☒ No Sample Preserved? ☐ Yes ☐ No Lot Number: _____ Exp: _____			Clarity Clear ☐ Slightly Turbid ☐ Turbid ☒ Opaque ☐		
System Type: ☒ Stream ☐ Lake ☐ Wetland ☐ Estuary ☐ Other (Specify) ☐			AMBIENT FIELD CONDITIONS / NOTES: ☐ The antecedent hydrologic conditions have been met to my best knowledge.					

SAMPLING TEAM Jeremy Pank / Brian Peris SIGNATURE: Jeremy Pank DATE: 11/30/20

DEP FORM FD 9000-25 Rapid Periphyton Survey Field Sheet

Site: <u>Unnamed abv. Alisaka cr.</u>					County: <u>Nassau</u>		Date: <u>11/30/20</u>		Investigator: <u>J. Smith</u>		
STORET Station Number: <u>GYNE0288</u>											
Transect t (m)	Point 1=right 9=left	Algal Thickness Rank (N-6, X)	Estimated	Canopy Cover	Transect t (m)	Point 1=right 9=left	Algal Thickness Rank (N-6, X)	Estimated	Canopy Cover		
0	1	N	X	80	60	1	N	N	92		
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										
10	1	N	N	46	70	1	N	N	92		
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										
20	1	N	N	92	80	1	N	N	96		
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										
30	1	N	N	92	90	1	N	N	92		
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										
40	1	N	N	92	100	1	N	N	94		
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										
50	1	N	N	94		1	N	N			
	2										
	3										
	4										
	5										
	6										
	7										
	8										
	9										

Record "N" and check "Estimated" for points deeper than the Secchi depth; algae are presumed absent. Check "Estimated" if thickness estimated for deep points which can be seen but not reached.

Record "X" for points shallower than Secchi depth for which substrate cannot be seen or reached with the hand. No estimated rank.

Record canopy cover as the number of small densiometer quadrants (out of 96) that HAVE canopy cover. Measure facing upstream at point 4, 5, or 6.

Collect algal mat sample following DEP SOP FS 7240 if the percentage of total sampled points with a thickness rank of 4, 5, or 6 is >20%.

Secchi depth 0.4
Estimated? No

points ranked 4-6 9
total points assessed 9
% points ranked 4-6 100

Check accompanying data collected at same site/date:
Algal mat sample ☐
Linear Veg Survey ☐
Habitat Assessment ☐
SCI/Biorecon ☐
Water sample ☐
RQ- 2020.11.30.38

Algal Thickness Rank	
N	rough, no algae, slimy, algae up to 1mm
3	>1mm - 6mm
4	>6mm - 20mm
5	>20mm - 10 cm
6	>10 cm

Date of Request: 16-NOV-2020
 Created By: PARRISH_J On: 16-NOV-2020
 Modified By: JASROTIA_P On: 18-NOV-2020
 Customer: NE-DIST
 Project: SMP
 Division: Division of Environmental Assessment and Restoration
 District: Northeast District
 Event Description: Callahan makeup

Send Coolers To:

Phone: 904-256-1700
 8800 Baymeadows Way West
 Suite 100
 Jacksonville, FL 32256
 Attn: Jeremy Parrish
 Cooler Ship EMail: Jeremy.M.Parrish@dep.state.fl.us

Priority: 3
 Event Status: R
 Criminal Investigation: NO
 Chemistry Request Reviewed By: WATTS_J On: 17-NOV-2020
 Biology Request Reviewed By: JASROTIA_P On: 18-NOV-2020
 Sampling Kit Required: NO Ship on:
 Sampling Kit Shipped:
 Sampling Kit Packed By:
 Preservatives Needed: NO
 Date to Receive Samples: 30-NOV-2020
 Logged In By: SHAIK_A On: 01-DEC-2020

Send Final Report To:

8800 Baymeadows Way West
 Suite 100
 Jacksonville, FL 32256
 Attn: Jeremy Parrish
 Report Notification EMail: Jeremy.M.Parrish@dep.state.fl.us

Comments: Group A: (Surf-Fresh) Nutrients, Bacteria
 Group B: (Surf-Fresh) Nutrients, Metals, Bacteria, tracers
 Group C: (Surf-Fresh) Bacteria

NE-ALS-ECQ

Packing Notes:
 None

Bottle Group A (Water) With 1 Samples:

Bottle Type: P-1L	Number of Bottles: 1	Preserved With: ICE
ALKALINITY	Template: DEFAULT	(SM 2320 B-2011) Alkalinity in aqueous matrices as mg CaCO ₃ /L
TURBIDITY	Template: DEFAULT	(EPA 180.1 Rev. 2.0) Turbidity in aqueous matrices
W-CL-IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Chloride in aqueous matrices
W-COLOR	Template: DEFAULT	(SM 2120 B-2011) True Color measured at 450 nm (not valid for NPDES monitoring)
Color (true)		
W-F	Template: DEFAULT	(SM 4500 F-C-2011) Fluoride in aqueous matrices, without distillation (not valid for NPDES monitoring)
W-SO ₄ -IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Sulfate in aqueous matrices
W-TDS	Template: DEFAULT	(SM 2540 C-2011) Total Dissolved Solids in aqueous matrices
W-TSS	Template: DEFAULT	(SM 2540 D-2011) Total Suspended Solids in aqueous matrices using a maximum of 250 mL of sample
Bottle Type: BP-1L	Number of Bottles: 1	Preserved With: ICE
CHLSUITE-W	Template: DEFAULT	(SM 10200 H (mod.)) Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry
Bottle Type: P-250ML-WI-THI	Number of Bottles: 1	Preserved With: ICE
NE-ALS-ECQ	Template: DEFAULT	(SM 9223 B Quanti-Tray) E. coli by Colilert/QT by ALS in Jacksonville
Bottle Type: P-500ML	Number of Bottles: 1	Preserved With: HNO ₃
W-HARD	Template: DEFAULT	(SM 2340B) Hardness by calculation, in aqueous matrices, reported as mg/L CaCO ₃
Hardness		
Bottle Type: P-500ML	Number of Bottles: 1	Preserved With: ICE And H ₂ SO ₄
W-NH ₃	Template: DEFAULT	(EPA 350.1 Rev. 2.0) Ammonia in aqueous matrices as mg N/L
W-NO ₂ NO ₃	Template: DEFAULT	(EPA 353.2 Rev. 2.0) Nitrite/Nitrate in aqueous matrices as mg N/L
W-S-A-TP	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Total Phosphorus in aqueous matrices as mg P/L

Bottle Group A (Water) With 1 Samples:

Bottle Type: P-500ML	Number of Bottles: 1	Preserved With: ICE And H2SO4
W-TKN	Template: DEFAULT	(EPA 351.2 Rev. 2.0) Total Kjeldahl Nitrogen in aqueous matrices
W-TOC	Template: DEFAULT	(SM 5310 B-2011) Nonpurgeable Organic Carbon in aqueous matrices
Bottle Type: P-125ML	Number of Bottles: 1	Preserved With: FILTER-ICE
W-PO4-F	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Ortho-phosphate, dissolved, in filtered, aqueous matrices as mg P/L

Bottle Group B (Water) With 1 Samples:

Bottle Type: BG-500ML	Number of Bottles: 1	Preserved With: ICE
W-E8321-DI	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS
W-E8321-MS	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS
Bottle Type: P-500ML	Number of Bottles: 1	Preserved With: HNO3
W-HARD	Template: DEFAULT	(SM 2340B) Hardness by calculation, in aqueous matrices, reported as mg/L CaCO3
Hardness		
W-ICP	Template:	(EPA 200.7 Rev. 4.4) Total Recoverable Metals analysis using ICP emission spectroscopy for aqueous samples supporting Clean Water Act Projects
Barium		
Calcium		
Iron		
Magnesium		
Potassium		
Sodium		
Zinc		
W-ICPMS	Template:	(EPA 200.8 Rev. 5.4) Total Recoverable Metals analysis using ICP-MS for aqueous samples supporting Clean Water Act Projects
*Boron		
Aluminum		
Antimony		
Arsenic		
Cadmium		
Chromium		
Copper		
Lead		
Nickel		
Selenium		
Silver		
Thallium		
Bottle Type: P-500ML	Number of Bottles: 1	Preserved With: ICE And H2SO4
W-NH3	Template: DEFAULT	(EPA 350.1 Rev. 2.0) Ammonia in aqueous matrices as mg N/L
W-NO2NO3	Template: DEFAULT	(EPA 353.2 Rev. 2.0) Nitrite/Nitrate in aqueous matrices as mg N/L
W-S-A-TP	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Total Phosphorus in aqueous matrices as mg P/L
W-TKN	Template: DEFAULT	(EPA 351.2 Rev. 2.0) Total Kjeldahl Nitrogen in aqueous matrices
W-TOC	Template: DEFAULT	(SM 5310 B-2011) Nonpurgeable Organic Carbon in aqueous matrices
Bottle Type: P-125ML	Number of Bottles: 1	Preserved With: FILTER-ICE
W-PO4-F	Template: DEFAULT	(EPA 365.1 Rev. 2.0) Ortho-phosphate, dissolved, in filtered, aqueous matrices as mg P/L
Bottle Type: P-1L	Number of Bottles: 1	Preserved With: ICE
ALKALINITY	Template: DEFAULT	(SM 2320 B-2011) Alkalinity in aqueous matrices as mg CaCO3/L
TURBIDITY	Template: DEFAULT	(EPA 180.1 Rev. 2.0) Turbidity in aqueous matrices
W-CL-IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Chloride in aqueous matrices
W-COLOR	Template: DEFAULT	(SM 2120 B-2011) True Color measured at 450 nm (not valid for NPDES monitoring)
Color (true)		
W-F	Template: DEFAULT	(SM 4500 F-C-2011) Fluoride in aqueous matrices, without distillation (not valid for NPDES monitoring)
W-SO4-IC	Template: DEFAULT	(EPA 300.0 Rev. 2.1) Sulfate in aqueous matrices
W-TDS	Template: DEFAULT	(SM 2540 C-2011) Total Dissolved Solids in aqueous matrices
W-TSS	Template: DEFAULT	(SM 2540 D-2011) Total Suspended Solids in aqueous matrices using a maximum of 250 mL of sample
Bottle Type: BP-1L	Number of Bottles: 1	Preserved With: ICE
CHLSUITE-W	Template: DEFAULT	(SM 10200 H (mod.)) Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry
Bottle Type: P-250ML-WI-THI	Number of Bottles: 1	Preserved With: ICE
NE-ALS-ECQ	Template: DEFAULT	(SM 9223 B Quanti-Tray) E. coli by Colilert/QT by ALS in Jacksonville

Bottle Group C (Water) With 1 Samples:

Bottle Type: P-250ML-WI-THI

Number of Bottles: 1

Preserved With: ICE

NE-ALS-ECQ

Template: DEFAULT

(SM 9223 B Quanti-Tray) E. coli by Colilert/QT by ALS in Jacksonville

* - The laboratory is not NELAP certified for this analyte/method, or certification is not applicable.