

Biological Analysis Report

NE-DIST-2022-09-16-02

Florida DEP Laboratory
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DOH Accreditation E31780

Overflow Analyses Performed By:
Diversified Environmental Laboratories, Inc.
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DOH Accreditation E821059

Event Description: **LSJR Palatka 2022**
Request ID: **RQ-2022-09-05-37**
Customer: **NE-DIST**
Project ID: **SMP**

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Certified by: Cheryl A. Swanson, Program Administrator

Date Certified: 31-OCT-2022 16:42



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: MILL BR. @ OLD SAN MATEO RD.

Collection Date/Time: 09/15/2022 09:45

Field ID: 20030577

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2357238	SM 10200 H (mod.)	Chlorophyll-a, Corrected	1.8	I	ug/L	P420541	
		Phaeophytin-a	1.8	U	ug/L	P420541	
		Chlorophyll-a, Uncorrected	1.9	I	ug/L	P420541	
2357242	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	920.8		MPN/100 mL		
2357252	SOP-PCR-1.0	HF183-purified-qPCR**	550	UJ	TSC/100 mL	P420676	LCS, SUR
	SOP-PCR-1.4	BacR-purified-qPCR**	1.63E+04	J	TSC/100 mL	P420676	LCS, SUR
	SOP-PCR-1.5	DG3-purified-qPCR**	430	UJ	TSC/100 mL	P420676	LCS, SUR

Ref. Method and Comment:

SM 10200 H (mod.): Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

SOP-PCR-1.0: Precision data unavailable for HF183-purified-qPCR component. Refer to the Lab Analysis Report for an explanation of QC Codes.

SOP-PCR-1.4: Precision data unavailable for BacR-purified-qPCR component. Refer to the Lab Analysis Report for an explanation of QC Codes.

SOP-PCR-1.5: Precision data unavailable for DG3-purified-qPCR component. Refer to the Lab Analysis Report for an explanation of QC Codes.

Sample Location: MILL BRANCH @ McCORMICK RD

Collection Date/Time: 09/15/2022 10:30

Field ID: 20030681

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2357244	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	866.4		MPN/100 mL		

Sample Location: COW CREEK @ E RIVER RD

Collection Date/Time: 09/15/2022 11:15

Field ID: 20030844

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2357245	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	517.2		MPN/100 mL		

Sample Location: Cow Creek at Putnam County Bivd

Collection Date/Time: 09/15/2022 11:35

Field ID: G2NE1143

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2357246	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	1413.6		MPN/100 mL		

Sample Location: Unnamed Ditch at Cowpen Branch Rd

Collection Date/Time: 09/15/2022 12:15

Field ID: G2NE1222

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2357239	SM 10200 H (mod.)	Chlorophyll-a, Corrected	3.0	I	ug/L	P420541	
		Phaeophytin-a	1.8	U	ug/L	P420541	
		Chlorophyll-a, Uncorrected	2.6	I	ug/L	P420541	
2357243	SM 9223 B Quanti-Tray	Escherichia Coli-Quanti-Tray	488.4		MPN/100 mL		

Ref. Method and Comment:

SM 10200 H (mod.): Lab dup result is 2.8 I for Chl-a Corrected, 2.5 I for Chl-a Uncorrected, and 1.8 U for Phaeophytin-a. Precision data is unavailable due to the small amount of analyte in the QC sample.

Quality Assurance Report

Method Blank Results

Quality Assurance Report Method Blank Results

Reference Method: SM 10200 H (mod.)
Batch ID: P420541

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.82	U	ug/L
Chlorophyll-a, Uncorrected	0.60	U	ug/L
Phaeophytin-a	0.90	U	ug/L

Reference Method: SOP-PCR-1.0
Batch ID: P420676

Component	Result	Code	Units
HF183-purified-qPCR	550	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P420676

Component	Result	Code	Units
BacR-purified-qPCR	510	U	TSC/100 mL

Reference Method: SOP-PCR-1.5
Batch ID: P420676

Component	Result	Code	Units
DG3-purified-qPCR	430	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10200 H (mod.)
Batch ID: P420541

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	95.0		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P420676

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-purified-qPCR	101	7.68	P/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P420676

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-purified-qPCR	101	4.26	P/F	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P420676

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
DG3-purified-qPCR	97.7	4.91	P/F	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P420676

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2357252	HF183-purified-qPCR	96.8		P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P420676

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2357252	BacR-purified-qPCR	101		P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P420676

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2357252	DG3-purified-qPCR	103		P	50 - 175

Quality Assurance Report Surrogates

Lab Sample ID: 2357252
Field Sample ID: 20030577

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	8.95	F	50 - 300
SOP-PCR-1.4	SKETA-qPCR	8.45	F	30 - 300
SOP-PCR-1.5	SKETA-qPCR	8.52	F	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery	MS % Recovery	LCS	Precision SMP	MS
SM 10200 H (mod.)	Chlorophyll-a, Corrected	95.0				
SOP-PCR-1.0	HF183-purified-qPCR	7.68 101	96.8			
SOP-PCR-1.4	BacR-purified-qPCR	4.26 101	101			
SOP-PCR-1.5	DG3-purified-qPCR	4.91 97.7	103			

Reference Method Descriptions

Method	Description	Associated Samples
SM 10200 H (mod.)	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2357238, 2357239
SM 9223 B Quanti-Tray	E. coli by Colilert/QT by Diversified Environmental Laboratories in Jacksonville	2357242, 2357243, 2357244, 2357245, 2357246
SOP-PCR-1.0	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2357252
SOP-PCR-1.4	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2357252
SOP-PCR-1.5	Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction	2357252

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SM 10200 H (mod.)	09/16/2022	09/16/2022 13:34	Megan Higbee	10/04/2022 16:22	Carina A. Tull	2357238, 2357239
SM 9223 B Quanti-Tray	09/15/2022	09/15/2022 15:15		09/16/2022 15:15	Diversified-Jacksonville	2357242, 2357243, 2357244, 2357245, 2357246
SOP-PCR-1.0	09/16/2022	09/16/2022 13:25	DeAsia Armster	10/14/2022 09:58	Roberta Tatti	2357252
SOP-PCR-1.4	09/16/2022	09/16/2022 13:25	DeAsia Armster	10/14/2022 12:30	DeAsia Armster	2357252
SOP-PCR-1.5	09/16/2022	09/16/2022 13:25	DeAsia Armster	10/14/2022 09:58	Roberta Tatti	2357252