

Biological Analysis Report

SO-DIST-2023-06-02-01

Florida DEP Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Overflow Analyses Performed By:
Sanders Laboratories
10090 Bavaria Road
Ft. Myers, FL 33913
DOH Accreditation E85457

Event Description: **2023 SMP: Glades Slough**

Request ID: **RQ-2023-05-08-44**

Customer: **SO-DIST**

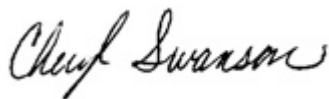
Project ID: **SMP**

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2295 Victoria Ave. Suite 314
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Certified by: Cheryl A. Swanson, Program Administrator

Date Certified: 18-JUL-2023 17:16



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: FENCELINE DRAIN AT BLUE HEAD RANCH

Collection Date/Time: 06/01/2023 10:20

Field ID: G4SD0193

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414175	Colilert-18	Escherichia Coli-Quanti-Tray	816		MPN/100 mL		

Sample Location: GATOR SLOUGH AT US27

Collection Date/Time: 06/01/2023 11:20

Field ID: G4SD0214

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414167	SM 10150 B	Chlorophyll-a, Corrected	50		ug/L	P431061	
		Phaeophytin-a	57		ug/L	P431061	
		Chlorophyll-a, Uncorrected	86		ug/L	P431061	
2414172	Colilert-18	Escherichia Coli-Quanti-Tray	14.8		MPN/100 mL		

Ref. Method and Comment:

SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: BOOTHEELCREEKEASTBRANCH

Collection Date/Time: 06/01/2023 11:45

Field ID: G4SD0105

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414168	SM 10150 B	Chlorophyll-a, Corrected	3.6		ug/L	P431061	
		Phaeophytin-a	3.5		ug/L	P431061	
		Chlorophyll-a, Uncorrected	5.9		ug/L	P431061	
2414173	Colilert-18	Escherichia Coli-Quanti-Tray	488		MPN/100 mL		

Ref. Method and Comment:

SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: RAINEY SLOUGH AT FARABEE RD

Collection Date/Time: 06/01/2023 12:30

Field ID: G4SD0111

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414176	Colilert-18	Escherichia Coli-Quanti-Tray	579		MPN/100 mL		

Sample Location: JOHNHENRYSLOUGH ATCR731

Collection Date/Time: 06/01/2023 12:45

Field ID: G4SD0215

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414170	SM 10150 B	Chlorophyll-a, Corrected	7.6		ug/L	P431061	
		Phaeophytin-a	0.90	U	ug/L	P431061	
		Chlorophyll-a, Uncorrected	8.3		ug/L	P431061	
2414171	SOP-PCR-1.0	HF183-qPCR**	2.1E+03	I	TSC/100 mL	P430325	
	SOP-PCR-1.4	BacR-qPCR**	1.5E+03	U	TSC/100 mL	P430325	
	SOP-PCR-1.5	DG3-qPCR**	1.3E+03	U	TSC/100 mL	P430325	
2414177	Colilert-18	Escherichia Coli-Quanti-Tray	2420	J	MPN/100 mL		

Ref. Method and Comment:

SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Colilert-18: All Quanti-Tray wells were positive. Result may be higher than reported.

Sample Location: GOPHER GULLY AT US27

Collection Date/Time: 06/01/2023 13:15

Field ID: G4SD0216

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2414169	SM 10150 B	Chlorophyll-a, Corrected	6.0	I	ug/L	P431061	
		Phaeophytin-a	3.0	U	ug/L	P431061	
		Chlorophyll-a, Uncorrected	6.6		ug/L	P431061	
2414174	Colilert-18	Escherichia Coli-Quanti-Tray	308		MPN/100 mL		

Ref. Method and Comment:
SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Quality Assurance Report Method Blank Results

Reference Method: SM 10150 B
Batch ID: P431061

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.82	U	ug/L
Chlorophyll-a, Uncorrected	0.60	U	ug/L
Phaeophytin-a	0.90	U	ug/L

Reference Method: SOP-PCR-1.0
Batch ID: P430325

Component	Result	Code	Units
HF183-qPCR	3.3E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P430325

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.5
Batch ID: P430325

Component	Result	Code	Units
DG3-qPCR	2.6E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10150 B
Batch ID: P431061

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	93.1		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P430325

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	96.6	100	P/P	50 - 175

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SOP-PCR-1.4
Batch ID: P430325

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	126	74.7	P/P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P430325

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
DG3-qPCR	94.8	78.8	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P430325

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2412133	HF183-qPCR	54.2	82.9	P	50 - 175
2412134	HF183-qPCR	89.0		P/P	50 - 175
2412247	HF183-qPCR	75.9	82.9	P	50 - 175
2412540	HF183-qPCR	89.8		P	50 - 175
2412798	HF183-qPCR	94.2	82.9	P	50 - 175
2412799	HF183-qPCR	82.6		P	50 - 175
2412859	HF183-qPCR	89.2	82.9	P	50 - 175
2412860	HF183-qPCR	77.7		P	50 - 175
2412950	HF183-qPCR	93.8	82.9	P	50 - 175
2414171	HF183-qPCR	87.1		P	50 - 175
2414608	HF183-qPCR	93.6	82.9	P	50 - 175
2414962	HF183-qPCR	108		P	50 - 175
2415132	HF183-qPCR	94.3		P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P430325

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2412133	BacR-qPCR	80.2	90.6	P	50 - 175
2412134	BacR-qPCR	97.1		P/P	50 - 175
2412247	BacR-qPCR	82.1	90.6	P	50 - 175
2412540	BacR-qPCR	84.2		P	50 - 175
2412798	BacR-qPCR	69.4	90.6	P	50 - 175
2412799	BacR-qPCR	77.3		P	50 - 175
2412859	BacR-qPCR	92.6	90.6	P	50 - 175
2412860	BacR-qPCR	85.0		P	50 - 175
2412950	BacR-qPCR	95.4	90.6	P	50 - 175
2414171	BacR-qPCR	89.6		P	50 - 175
2414608	BacR-qPCR	72.9	90.6	P	50 - 175
2414962	BacR-qPCR	90.3		P	50 - 175
2415132	BacR-qPCR	96.9		P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P430325

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
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Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.5
Batch ID: P430325

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2412133	DG3-qPCR	69.5	82.0	P	50 - 175
2412134	DG3-qPCR	84.9		P/P	50 - 175
2412247	DG3-qPCR	79.6		P	50 - 175
2412540	DG3-qPCR	87.9		P	50 - 175
2412798	DG3-qPCR	83.8		P	50 - 175
2412799	DG3-qPCR	81.7		P	50 - 175
2412859	DG3-qPCR	86.6		P	50 - 175
2412860	DG3-qPCR	78.2		P	50 - 175
2412950	DG3-qPCR	86.6		P	50 - 175
2414171	DG3-qPCR	87.9		P	50 - 175
2414608	DG3-qPCR	92.0		P	50 - 175
2414962	DG3-qPCR	86.8		P	50 - 175
2415132	DG3-qPCR	87.2		P	50 - 175

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P430325

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2412134	HF183-qPCR	7.10	Spike	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P430325

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2412134	BacR-qPCR	6.92	Spike	P	0 - 25

Reference Method: SOP-PCR-1.5
Batch ID: P430325

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2412134	DG3-qPCR	3.42	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.

Quality Assurance Report Surrogates

Lab Sample ID: 2414171
Field Sample ID: G4SD0215

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	94.7	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	104	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	87.0	P	50 - 300

Quality Assurance Report

Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			Precision	MS
							SMP	
SM 10150 B	Chlorophyll-a, Corrected	93.1						
SOP-PCR-1.0	HF183-qPCR	100	96.6	94.3	54.2	89.0		7.10
				82.9				
SOP-PCR-1.4	BacR-qPCR	74.7	126	80.2	97.1	90.6		6.92
				82.1				
SOP-PCR-1.5	DG3-qPCR	78.8	94.8	69.5	84.9	82.0		3.42
				79.6				

Reference Method Descriptions

Method	Description	Associated Samples
Colilert-18	E. coli by Colilert-18/QT by Sanders in Fort Myers	2414172, 2414173, 2414174, 2414175, 2414176, 2414177
SM 10150 B	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2414167, 2414168, 2414169, 2414170
SOP-PCR-1.0	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2414171
SOP-PCR-1.4	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2414171
SOP-PCR-1.5	Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction	2414171

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
Colilert-18	06/01/2023	06/01/2023 16:05		06/02/2023 10:30	Sanders-Fort Myers	2414172, 2414173, 2414174, 2414175, 2414176, 2414177
SM 10150 B	06/02/2023	06/02/2023 13:08	Keanu Thijm	06/07/2023 13:18	Callie Williams	2414167, 2414168, 2414169, 2414170
SOP-PCR-1.0	06/02/2023	06/02/2023 14:30	Claudia Castro	06/15/2023 13:35	Jennifer Mihalic	2414171
SOP-PCR-1.4	06/02/2023	06/02/2023 14:30	Claudia Castro	06/16/2023 11:46	Jennifer Mihalic	2414171
SOP-PCR-1.5	06/02/2023	06/02/2023 14:30	Claudia Castro	06/16/2023 14:43	Jennifer Mihalic	2414171