

Biological Analysis Report

SO-DIST-2023-10-04-01

Florida DEP Laboratory
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DOH Accreditation E31780

Overflow Analyses Performed By:
Sanders Laboratories
10090 Bavaria Road
Ft. Myers, FL 33913
DOH Accreditation E85457

Event Description: **2023 SMP: Felda Boys**

Request ID: **RQ-2023-10-02-63**

Customer: **SO-DIST**

Project ID: **SMP**

Send Reports to:
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FLDEP South District Office
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Date Certified: 14-NOV-2023 14:09



Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: L 61W

Collection Date/Time: 10/03/2023 09:40

Field ID: G4SD0194

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2442505	SM 10150 B	Chlorophyll-a, Corrected	4.2	I	ug/L	P436732	
		Phaeophytin-a	2.5	I	ug/L	P436732	
		Chlorophyll-a, Uncorrected	5.9		ug/L	P436732	
2442516	Colilert-18	Escherichia Coli-Quanti-Tray	148		MPN/100 mL		

Ref. Method and Comment:
 SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: NINE MILE CANAL AT SR 80

Collection Date/Time: 10/03/2023 11:05

Field ID: G3SD0088

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2442517	Colilert-18	Escherichia Coli-Quanti-Tray	98.5		MPN/100 mL		

Sample Location: TOWNSEND CANAL AT CHURCH RD

Collection Date/Time: 10/03/2023 12:05

Field ID: G3SD0221

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2442506	SM 10150 B	Chlorophyll-a, Corrected	8.4		ug/L	P436732	
		Phaeophytin-a	2.2	I	ug/L	P436732	
		Chlorophyll-a, Uncorrected	10		ug/L	P436732	
2442518	Colilert-18	Escherichia Coli-Quanti-Tray	48.0		MPN/100 mL		

Ref. Method and Comment:
 SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: COW SLOUGH

Collection Date/Time: 10/03/2023 13:00

Field ID: IMKFSHCK

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2442507	SOP-PCR-1.0	HF183-qPCR**	1.7E+03	U	TSC/100 mL	P435503	LCS
	SOP-PCR-1.4	BacR-qPCR**	3.1E+03	I	TSC/100 mL	P435503	
	SOP-PCR-1.5	DG3-qPCR**	1.3E+03	U	TSC/100 mL	P435503	
2442519	Colilert-18	Escherichia Coli-Quanti-Tray	120		MPN/100 mL		

Quality Assurance Report Method Blank Results

Reference Method: SM 10150 B
Batch ID: P436732

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.96	U	ug/L
Chlorophyll-a, Uncorrected	0.70	U	ug/L
Phaeophytin-a	1.0	U	ug/L

Quality Assurance Report Method Blank Results

Reference Method: SOP-PCR-1.0
Batch ID: P435503

Component	Result	Code	Units
HF183-qPCR	3.3E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P435503

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.5
Batch ID: P435503

Component	Result	Code	Units
DG3-qPCR	2.6E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10150 B
Batch ID: P436732

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	95.8		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P435503

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	67.3	13.9	P/F	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P435503

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	102	62.9	P/P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P435503

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
DG3-qPCR	107	65.4	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P435503

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2440404	HF183-qPCR	98.9	105	P/P	50 - 175
2441149	HF183-qPCR	78.9		P	50 - 175
2441337	HF183-qPCR	81.4		P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P435503

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2441586	HF183-qPCR	67.9		P	50 - 175
2441587	HF183-qPCR	97.9		P	50 - 175
2442197	HF183-qPCR	88.2		P	50 - 175
2442198	HF183-qPCR	76.7		P	50 - 175
2442312	HF183-qPCR	97.2		P	50 - 175
2442410	HF183-qPCR	70.6		P	50 - 175
2442507	HF183-qPCR	87.1		P	50 - 175
2443119	HF183-qPCR	77.3		P	50 - 175
2443140	HF183-qPCR	84.3		P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P435503

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2440404	BacR-qPCR	90.4	92.9	P/P	50 - 175
2441149	BacR-qPCR	69.3		P	50 - 175
2441337	BacR-qPCR	88.1		P	50 - 175
2441586	BacR-qPCR	75.2		P	50 - 175
2441587	BacR-qPCR	59.3		P	50 - 175
2442197	BacR-qPCR	60.3		P	50 - 175
2442198	BacR-qPCR	79.4		P	50 - 175
2442312	BacR-qPCR	69.3		P	50 - 175
2442410	BacR-qPCR	78.0		P	50 - 175
2442507	BacR-qPCR	53.3		P	50 - 175
2443119	BacR-qPCR	90.2		P	50 - 175
2443140	BacR-qPCR	84.0		P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P435503

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2440404	DG3-qPCR	99.1	96.1	P/P	50 - 175
2441149	DG3-qPCR	85.5		P	50 - 175
2441337	DG3-qPCR	85.2		P	50 - 175
2441586	DG3-qPCR	71.7		P	50 - 175
2441587	DG3-qPCR	91.4		P	50 - 175
2442197	DG3-qPCR	93.8		P	50 - 175
2442198	DG3-qPCR	95.7		P	50 - 175
2442312	DG3-qPCR	99.3		P	50 - 175
2442410	DG3-qPCR	90.6		P	50 - 175
2442507	DG3-qPCR	70.3		P	50 - 175
2443119	DG3-qPCR	88.8		P	50 - 175
2443140	DG3-qPCR	87.4		P	50 - 175

Quality Assurance Report Precision

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P435503

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2440404	HF183-qPCR	6.00	Spike	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P435503

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2440404	BacR-qPCR	2.75	Spike	P	0 - 25

Reference Method: SOP-PCR-1.5
Batch ID: P435503

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2440404	DG3-qPCR	3.15	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.

Quality Assurance Report Surrogates

Lab Sample ID: 2442507
Field Sample ID: IMKF5HCK

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	65.0	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	77.4	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	65.6	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10150 B	Chlorophyll-a, Corrected	95.8							
SOP-PCR-1.0	HF183-qPCR	13.9	67.3	98.9	105	78.9			6.00
				81.4					
SOP-PCR-1.4	BacR-qPCR	62.9	102	90.4	92.9	69.3			2.75
				88.1					
SOP-PCR-1.5	DG3-qPCR	65.4	107	99.1	96.1	85.5			3.15
				85.2					

Reference Method Descriptions

Method	Description	Associated Samples
Colilert-18	E. coli by Colilert-18/QT by Sanders in Fort Myers	2442516, 2442517, 2442518, 2442519
SM 10150 B	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2442505, 2442506

Reference Method Descriptions

Method	Description	<u>Associated Samples</u>
SOP-PCR-1.0	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2442507
SOP-PCR-1.4	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2442507
SOP-PCR-1.5	Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction	2442507

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
Colilert-18	10/03/2023	10/03/2023 14:40		10/04/2023 09:24	Sanders-Fort Myers	2442516, 2442517, 2442518, 2442519
SM 10150 B	10/04/2023	10/04/2023 13:45	Tynan A Branch	10/18/2023 08:55	Keanu Thijm	2442505, 2442506
SOP-PCR-1.0	10/04/2023	10/04/2023 14:45	Claudia Castro	10/25/2023 08:32	Roberta Tatti	2442507
SOP-PCR-1.4	10/04/2023	10/04/2023 14:45	Claudia Castro	10/26/2023 08:41	Roberta Tatti	2442507
SOP-PCR-1.5	10/04/2023	10/04/2023 14:45	Claudia Castro	10/26/2023 09:34	Roberta Tatti	2442507