

Biological Analysis Report

SO-DIST-2023-12-13-01

Florida DEP Laboratory
2600 Blair Stone Road
Tallahassee, FL 32399-2400
DOH Accreditation E31780

Overflow Analyses Performed By:
Sanders Laboratories
10090 Bavaria Road
Ft. Myers, FL 33913
DOH Accreditation E85457

Event Description: **2023 SMP: Felda Boys**

Request ID: **RQ-2023-11-27-21**

Customer: **SO-DIST**

Project ID: **SMP**

Send Reports to:
FL Dept. of Environmental Protection
FLDEP South District Office
2295 Victoria Ave. Suite 314
Fort Myers, FL 33901-3381
Attn: Nicole Reistad

For additional information please contact
Cheryl Swanson - Administrator
Sarah Menz, Ph.D. - Bench Biology & Aquatic
Toxicity
Puja Jasrotia, Ph.D. - Molecular Biology & Taxonomy
Thekkekalathil Chandrasekhar, Ph.D, QA Officer
Phone (850) 245-8177

Certified by: Sarah Menz, Environmental Administrator

Date Certified: 29-JAN-2024 10:19

A handwritten signature in dark ink, appearing to be 'Sarah Menz', written in a cursive style.

Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical groups are included in this report: Chlorophyll/Grain Size/BOD, Molecular and Overflow.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example: 1.3E+03, which is equivalent to 1.3×10^3 or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (**) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited by the Florida Department of Health Environmental Laboratory Certification Program or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: L 61W

Collection Date/Time: 12/12/2023 10:20

Field ID: G4SD0194

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2456643	SM 10150 B	Chlorophyll-a, Corrected	6.2		ug/L	P439318	
		Phaeophytin-a	0.90	U	ug/L	P439318	
		Chlorophyll-a, Uncorrected	6.8		ug/L	P439318	
2456655	Colilert-18	Escherichia Coli-Quanti-Tray	210		MPN/100 mL		

Ref. Method and Comment:

SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Nine Mile Canal at SR 80

Collection Date/Time: 12/12/2023 11:05

Field ID: G3SD0088

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2456656	Colilert-18	Escherichia Coli-Quanti-Tray	13.2		MPN/100 mL		

Sample Location: TOWNSEND CANAL AT CHURCH RD

Collection Date/Time: 12/12/2023 12:05

Field ID: G3SD0221

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2456644	SM 10150 B	Chlorophyll-a, Corrected	30		ug/L	P439318	
		Phaeophytin-a	8.9		ug/L	P439318	
		Chlorophyll-a, Uncorrected	36		ug/L	P439318	
2456657	Colilert-18	Escherichia Coli-Quanti-Tray	14.4		MPN/100 mL		

Ref. Method and Comment:

SM 10150 B: Precision data is not available for at least one component due to the small amount of analyte in the QC sample. Refer to QA report for available precision data.

Sample Location: Cow Slough

Collection Date/Time: 12/12/2023 13:05

Field ID: IMKFSHCK

Matrix: W-SURF-FRH

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
2456647	SOP-PCR-1.0	HF183-qPCR**	1.7E+03	U	TSC/100 mL	P438902	
	SOP-PCR-1.4	BacR-qPCR**	1.04E+05	A	TSC/100 mL	P438902	
	SOP-PCR-1.5	DG3-qPCR**	1.3E+03	U	TSC/100 mL	P438902	
2456658	Colilert-18	Escherichia Coli-Quanti-Tray	1120		MPN/100 mL		

Quality Assurance Report Method Blank Results

Reference Method: SM 10150 B

Batch ID: P439318

Component	Result	Code	Units
Chlorophyll-a, Corrected	0.82	U	ug/L
Chlorophyll-a, Uncorrected	0.60	U	ug/L
Phaeophytin-a	0.90	U	ug/L

Quality Assurance Report Method Blank Results

Reference Method: SOP-PCR-1.0
Batch ID: P438902

Component	Result	Code	Units
HF183-qPCR	3.3E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.4
Batch ID: P438902

Component	Result	Code	Units
BacR-qPCR	3.0E+03	U	TSC/100 mL

Reference Method: SOP-PCR-1.5
Batch ID: P438902

Component	Result	Code	Units
DG3-qPCR	2.6E+03	U	TSC/100 mL

Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SM 10150 B
Batch ID: P439318

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
Chlorophyll-a, Corrected	98.2		P	84 - 116

Reference Method: SOP-PCR-1.0
Batch ID: P438902

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
HF183-qPCR	96.3	73.6	P/P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P438902

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
BacR-qPCR	97.4	76.9	P/P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P438902

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
DG3-qPCR	91.1	74.7	P/P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P438902

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2456647	HF183-qPCR	88.5	91.6	P/P	50 - 175
2457024	HF183-qPCR	113		P	50 - 175
2457050	HF183-qPCR	104		P	50 - 175

Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR-1.0
Batch ID: P438902

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2457077	HF183-qPCR	105		P	50 - 175
2457241	HF183-qPCR	89.5		P	50 - 175
2457935	HF183-qPCR	90.2		P	50 - 175
2458096	HF183-qPCR	98.6		P	50 - 175
2458181	HF183-qPCR	95.5		P	50 - 175
2458438	HF183-qPCR	94.4		P	50 - 175
2458601	HF183-qPCR	84.8		P	50 - 175
2459071	HF183-qPCR	93.3		P	50 - 175

Reference Method: SOP-PCR-1.4
Batch ID: P438902

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2456647	BacR-qPCR	83.7	85.6	P/P	50 - 175
2457024	BacR-qPCR	78.1		P	50 - 175
2457050	BacR-qPCR	84.4		P	50 - 175
2457077	BacR-qPCR	77.5		P	50 - 175
2457241	BacR-qPCR	73.9		P	50 - 175
2457935	BacR-qPCR	61.1		P	50 - 175
2458096	BacR-qPCR	97.9		P	50 - 175
2458181	BacR-qPCR	73.3		P	50 - 175
2458438	BacR-qPCR	85.5		P	50 - 175
2458601	BacR-qPCR	78.4		P	50 - 175
2459071	BacR-qPCR	87.2		P	50 - 175

Reference Method: SOP-PCR-1.5
Batch ID: P438902

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
2456647	DG3-qPCR	94.3	98.7	P/P	50 - 175
2457024	DG3-qPCR	89.4		P	50 - 175
2457050	DG3-qPCR	99.9		P	50 - 175
2457077	DG3-qPCR	104		P	50 - 175
2457241	DG3-qPCR	93.1		P	50 - 175
2457935	DG3-qPCR	88.1		P	50 - 175
2458096	DG3-qPCR	104		P	50 - 175
2458181	DG3-qPCR	99.1		P	50 - 175
2458438	DG3-qPCR	101		P	50 - 175
2458601	DG3-qPCR	98.5		P	50 - 175
2459071	DG3-qPCR	104		P	50 - 175

Quality Assurance Report Precision

Reference Method: SM 10150 B
Batch ID: P439318

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2456725	Chlorophyll-a, Corrected	2.56	Sample	P	0 - 25
2456725	Chlorophyll-a, Uncorrected	1.18	Sample	P	0 - 25

Quality Assurance Report Precision

Reference Method: SOP-PCR-1.0
Batch ID: P438902

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2456647	HF183-qPCR	3.36	Spike	P	0 - 25

Reference Method: SOP-PCR-1.4
Batch ID: P438902

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2456647	BacR-qPCR	2.19	Spike	P	0 - 25

Reference Method: SOP-PCR-1.5
Batch ID: P438902

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
2456647	DG3-qPCR	4.52	Spike	P	0 - 25

* Sample, spike and/or laboratory control sample precision (LCS) is reported.

Quality Assurance Report Surrogates

Lab Sample ID: 2456647
Field Sample ID: IMKF5HCK

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR-1.0	SKETA-qPCR	99.0	P	50 - 300
SOP-PCR-1.0	SKETA-qPCR	100	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	107	P	50 - 300
SOP-PCR-1.4	SKETA-qPCR	112	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	108	P	50 - 300
SOP-PCR-1.5	SKETA-qPCR	102	P	50 - 300

Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			LCS	Precision SMP	MS
SM 10150 B	Chlorophyll-a, Corrected	98.2						2.56	
	Chlorophyll-a, Uncorrected							1.18	
SOP-PCR-1.0	HF183-qPCR	73.6	96.3	91.6	113	104			3.36
				105					
SOP-PCR-1.4	BacR-qPCR	97.4	76.9	83.7	85.6	78.1			2.19
				84.4					
SOP-PCR-1.5	DG3-qPCR	74.7	91.1	94.3	98.7	89.4			4.52
				99.9					

Reference Method Descriptions

Method	Description	<u>Associated Samples</u>
Colilert-18	E. coli by Colilert-18/QT by Sanders in Fort Myers	2456655, 2456656, 2456657, 2456658
SM 10150 B	Phytoplankton chlorophyll-a corrected, uncorrected, and phaeophytin by spectrophotometry	2456643, 2456644
SOP-PCR-1.0	Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction	2456647
SOP-PCR-1.4	Detection and quantification of Ruminant specific Bacteroidetes BacR genetic marker with quantitative polymerase chain reaction	2456647
SOP-PCR-1.5	Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction	2456647

Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
Colilert-18	12/12/2023	12/12/2023 15:17		12/13/2023 09:57	Sanders-Fort Myers	2456655, 2456656, 2456657, 2456658
SM 10150 B	12/13/2023	12/13/2023 13:44	Callie Williams	12/20/2023 10:04	Tynan A Branch	2456643, 2456644
SOP-PCR-1.0	12/13/2023	12/13/2023 13:56	Madeline Gruver	01/17/2024 08:58	Roberta Tatti	2456647
SOP-PCR-1.4	12/13/2023	12/13/2023 13:56	Madeline Gruver	01/18/2024 09:02	Roberta Tatti	2456647
SOP-PCR-1.5	12/13/2023	12/13/2023 13:56	Madeline Gruver	01/18/2024 09:57	Roberta Tatti	2456647