

Login Checklist

RQ- 2021-04-19-73

Login Station ID: #5

Shipping Method: RedEx | UPS | HD | Greyhound

Date/Time of Receipt: 4/ 22 /21 @ 9:33

*Cooler Temperature	Samples Frozen?	* All Samples in Cooler Preserved with		Number of Sample Containers in Cooler	Evidence Tape				Waybill Tracking #
					Present?		*Intact?		
	YES	HNO ₃	Formalin		Yes	No	Yes	No	
2.4				8		✓			9293 8009 4573

COOLER CHECK

*All samples received on ice unless otherwise noted. HNO3 and Formalin preserved samples do NOT have a temperature requirement.

- If the temperature of a cooler exceeds 6.0 oC (above 10.0 oC for W-1-4-DIOX and microbiology) or received without ice, identify affected samples in an NCR. NCR# _____
- DOH coolers can exceed 2.0 °C above the standard temperature (6.0 or 10.0 °C for W-1-4-DIOX) without login confirmation from customer for samples requiring ice preservation; identify affected samples in an NCR. NCR# _____
- If cooler or samples are received with a damaged Evidence Seal; identify the affected samples in an NCR. NCR# _____
- If coolers or samples are received late; identify the affected samples in an NCR. NCR# _____

Chain of Custody/ Field Sheet(s) Included?

Yes ✓ No _____

Micro-Biology Overflow Chain of Custody/ Field Sheet(s) Included?

Yes _____ No ✓

CONTAINER CHECK

Evidence Tape on Bottles?

Yes _____ No _____ NA ✓

Evidence Tape intact? Yes _____ No _____

If Criminal, photographs taken?

Yes _____ No _____

Containers intact? Yes ✓ No _____

Caps on tight?

Yes ✓ No _____

Sufficient Sample Volume:

Yes ✓ No _____

If NO is checked above, generate an NCR listing affected sample IDs in its appropriate category. NCR# _____

CHLORINE CHECK REQUIRED? (Blue dot on container)

Yes ✓ No _____ Init: HLK

Chlorine detected? (One container checked per Field ID)

Yes _____ No ✓ Init: HLK

If chlorine is detected, generate an NCR listing affected sample IDs . NCR# _____

PRESERVATION CHECK (EXCLUDING SAMPLES SENT TO OVERFLOW LABS, "OV-")

**Acid Preserved Samples: pH < 2.0?

Yes _____ No _____ NA ✓ Init: HLK

**W-1-4-DIOX (Green dot on container) preserved to pH < 4.0?

Yes _____ No _____ NA ✓ Init: HLK

If samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

Coolers Unpacked/Checked by: HLK

Date: 4/ 22 /21

Event contains NCRs (Y/N)? N

Event ID: bmap tribs
NE-DIST-2021-04-22-03 (SMP)

— Date Received: 22-APR-2021 09:33 —

Login Checklist

Login Preservation Equipment

Lot # & Exp: Date

Additional Comments:

Infrared Thermometer ID:	IR TEMP GUN #5 EXP:10-28-21
pH Test Strips 0 – 6	Lot#213116BV Exp:5-15-21
pH Paper 0 – 3	
pH Paper 0 – 13	Lot#210620 EXP:4-15-23
Chlorine Test Strips	Lot#0261 EXP:5-23

ENCORE SAMPLES

S-VOC-MS samples in Encores must be frozen within 48 hours of collection.

Were Encore samples received? Yes _____ Init: _____

Date and time placed in freezer _____

Were all samples placed in freezer within 48 hours of collection? Yes _____ No _____

If no, were samples shipped on dry ice? Yes _____ No _____

If S-VOC-MS samples were not preserved correctly, generate an NCR listing affected sample IDs . NCR# _____

FOR CLEAN LAB USE ONLY – Total Mercury Preservation Check(W-HG-AF;W-HG-AF-F):

Samples preserved within 48 hours? Yes _____ No _____

Date and time samples preserved: _____

Florida Department of Environmental Protection
Central Laboratory Submittal Form

Customer: NE-DIST

Requester: Brian Davis

Field Report Prepared By: K. McDonald

Project ID: SMP

Collected By: K. McDonald, B. DavisSampling Agency: FDEP NEROC

Location <u>Little Fishweir @ MacArthur</u>		☐ Comp ☒ Grab	Collection (comp begin or grab) Date <u>4/21/21</u> Time <u>0935</u>		☒ Eastern ☐ Central	Composite end Date <u> </u> Time <u> </u>	Bottle Group(s) (See pg 2)
Field ID <u>LFW2</u>		Matrix (type, e.g., Salt, Fresh, etc) <u>Fresh</u>		Diss. Oxygen <u>5.0</u>	☒ mg/L ☐ % sat	Total Res. Chlorine (mg/L) <u> </u>	<u>A</u>
WIN/STORET #		Temp (C) <u>20.5</u>	pH <u>7.42</u>	Sample Depth <u>0.05</u>	☐ ft ☒ m	Sp. Conductance (umho/cm) <u>428.9</u>	
Latitude <u>30.30231</u>	☒ dd ☐ dms	Longitude <u>-81.71719</u>	☒ dd ☐ dms	Salinity (ppt) <u>0.21</u>	Comments		
Location <u>Stormwater effluent @ Randall</u>		☐ Comp ☒ Grab	Collection (comp begin or grab) Date <u>4/21/21</u> Time <u>1010</u>		☒ Eastern ☐ Central	Composite end Date <u> </u> Time <u> </u>	Bottle Group(s)
Field ID <u>LFW5</u>		Matrix (type, e.g., Salt, Fresh, etc) <u>Fresh</u>		Diss. Oxygen <u>6.75</u>	☒ mg/L ☐ % sat	Total Res. Chlorine (mg/L) <u> </u>	<u>A</u>
WIN/STORET #		Temp (C) <u>20.9</u>	pH <u>7.4</u>	Sample Depth <u>0.05</u>	☐ ft ☒ m	Sp. Conductance (umho/cm) <u>480.1</u>	
Latitude <u>30.302943</u>	☒ dd ☐ dms	Longitude <u>-81.717751</u>	☒ dd ☐ dms	Salinity (ppt) <u>0.23</u>	Comments		
Location <u>Upstream of Randall</u>		☐ Comp ☒ Grab	Collection (comp begin or grab) Date <u>4/21/21</u> Time <u>1030</u> ^{KK}		☒ Eastern ☐ Central	Composite end Date <u> </u> Time <u> </u>	Bottle Group(s)
Field ID <u>LFW6</u>		Matrix (type, e.g., Salt, Fresh, etc) <u>Fresh</u>		Diss. Oxygen <u>2.58</u>	☒ mg/L ☐ % sat	Total Res. Chlorine (mg/L) <u> </u>	<u>A</u>
WIN/STORET #		Temp (C) <u>28.8</u>	pH <u>7.14</u>	Sample Depth <u>0.12</u>	☐ ft ☒ m	Sp. Conductance (umho/cm) <u>394.4</u>	
Latitude <u>30.303090</u>	☒ dd ☐ dms	Longitude <u>-81.177970</u>	☒ dd ☐ dms	Salinity (ppt) <u>0.19</u>	Comments		
Location <u>Rensselaer + Randall manhole</u>		☐ Comp ☒ Grab	Collection (comp begin or grab) Date <u>4/21/21</u> Time <u>1110</u>		☒ Eastern ☐ Central	Composite end Date <u> </u> Time <u> </u>	Bottle Group(s)
Field ID <u>LFW7</u>		Matrix (type, e.g., Salt, Fresh, etc) <u>Fresh</u>		Diss. Oxygen <u>7.73</u>	☒ mg/L ☐ % sat	Total Res. Chlorine (mg/L) <u> </u>	<u>A</u>
WIN/STORET #		Temp (C) <u>21.8</u>	pH <u>7.48</u>	Sample Depth <u>0.10</u>	☐ ft ☒ m	Sp. Conductance (umho/cm) <u>540</u>	
Latitude <u>30.303350</u>	☒ dd ☐ dms	Longitude <u>-81.7110 km</u>	☒ dd ☐ dms	Salinity (ppt) <u>0.26</u>	Comments		

Relinquished By: <u>Kelly McDonald</u>	Date/Time <u>4/21/21 1100</u>	Shipping Method <u>FedEx</u>	Number of Coolers Shipped: <u>1</u>	Received By: <u>KK</u>	Date/Time <u>4.22.21/0933</u>
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* Collection time taken from bottle

CALIBRATION AND VERIFICATION LOG (FDEP SOP FT 1000-FT 1500, FD 1000-FD 4000)

Boldly "X" this box if there are qualified data on this page.

Meter ID: 54126 RQ: 2021-04-19-73 Project: LFV Source Tracking

- Notes: (1) Always wait for meter to stabilize before recording any readings.
 (2) Report all digits displayed. Do not round before reporting measurements. (See special instructions for depth).
 (3) For Calibrations, record calibrated meter reading. Do not record initial meter reading before calibration.

Temperature (Quarterly) FT 1400

Date of Last Temperature Verification 2/15/21

DO DEP SOP FT 1500	Name	Date	Time CT-ET	Temp °C	Baro-meter mmHg	D.O. Chart mg/L	Meter D.O. mg/L	% DO	Probe Charge	Probe Gain	Pass / Fail	Lab / Field
Calibr.	<u>Brian Davis</u>	<u>4/21/21</u>	<u>735</u>	<u>22.6</u>	<u>760.9</u>	<u>8.7</u>	<u>8.58</u>	<u>100.1</u>	<u>✓</u>	<u>1.02</u>	<u>P/F</u>	<u>L/F</u>
ICV			<u>738</u>	<u>22.8</u>	<u>760.8</u>	<u>8.6</u>	<u>8.58</u>	<u>99.9</u>	<u>✓</u>	<u>1.02</u>	<u>P/F</u>	<u>L/F</u>
CCV			<u>1540</u>	<u>24.5</u>	<u>758.9</u>	<u>8.3</u>	<u>8.24</u>	<u>98.9</u>	<u>✓</u>	<u>1.02</u>	<u>P/F</u>	<u>L/F</u>
CCV											<u>P/F</u>	<u>L/F</u>

DO Acceptance criteria from Table ± 0.3 mg/L.

Rapid-Pulse Sensors: DO Gain Range 0.7 to 1.4; DO Charge Range 25-75.

Optical: DO gain range 0.85 to 1.15 (Pro DSS 0.75 to 1.50); DO charge N/A. Steady-state & Galvanic Sensors: DO Gain & Charge N/A.

Spec. Cond. FT 1200	Name	Date	Time CT-ET	Lot #	Expir. Date	Standard μ hos/cm	Meter Reading μ hos/cm	Pass / Fail	Lab / Field
Calibr.	<u>Brian Davis</u>	<u>4/21/21</u>	<u>740</u>	<u>201222B</u>	<u>1/22</u>	<u>1000</u>	<u>1000</u>	<u>P/F</u>	<u>L/F</u>
ICV			<u>742</u>	<u>201222A</u>	<u>1/22</u>	<u>100</u>	<u>103.3</u>	<u>P/F</u>	<u>L/F</u>
CCV			<u>1543</u>	<u>200902C</u>	<u>9/21</u>	<u>1413</u>	<u>1407</u>	<u>P/F</u>	<u>L/F</u>
CCV								<u>P/F</u>	<u>L/F</u>

Conductivity Acceptance criteria $\pm 5\%$

pH DEP SOP FT 1100	Name	Date	Time CT-ET	Lot #	Expir. Date	pH Buffer SU	Temp °C	Meter reading SU	mV	Pass / Fail	Lab / Field
Calibr.	<u>Brian Davis</u>	<u>4/21/21</u>	<u>746</u>	<u>2010F27</u>	<u>10/22</u>	<u>7.01</u>	<u>23.3</u>	<u>7.01</u>	<u>-20.1</u>	<u>P/F</u>	<u>L/F</u>
Calibr.			<u>749</u>	<u>2010C31</u>	<u>9/22</u>	<u>4.00</u>	<u>23.2</u>	<u>4.00</u>	<u>157.0</u>	<u>P/F</u>	<u>L/F</u>
Calibr.			<u>752</u>	<u>2012797</u>	<u>5/22</u>	<u>10.02</u>	<u>23.3</u>	<u>10.02</u>	<u>-192.2</u>	<u>P/F</u>	<u>L/F</u>
ICV			<u>754</u>	<u>2010F27</u>	<u>10/22</u>	<u>7.01</u>	<u>23.3</u>	<u>7.05</u>	<u>-20.7</u>	<u>P/F</u>	<u>L/F</u>
CCV			<u>1545</u>	<u>2012797</u>	<u>5/22</u>	<u>10.02</u>	<u>23.2</u>	<u>10.02</u>	<u>-192.2</u>	<u>P/F</u>	<u>L/F</u>
CCV										<u>P/F</u>	<u>L/F</u>

pH Acceptance criteria ± 0.2 SU; mV pH 7 Range 0 ± 50 ; mV pH 4 Range $+180 \pm 50$; mV pH 10 Range -180 ± 50 ;

If mV are recorded: slope from 7 to 10 _____, slope from 4 to 7 _____ (both must be between 165 and 180 mV)

Does meter have a depth sensor that will be used to measure total depth & sample depth? YES / NO / NA (not surf. water project)

If YES, complete daily Calibr. & ICV below and list date of last quarterly depth verification: _____

If NO, what will be used? (circle one) Secchi Disk Line / Sonar Unique ID: _____; Date of last verification: _____

Depth Sensor (Daily Calibration & ICV)	Name	Date	Time CT-ET	Calibrated Value (0.00 or Offset), meters	ICV Value, meters	Pass / Fail	Lab / Field
Pressure mode in air	<u>Brian Davis</u>	<u>4/11/21</u>	<u>755</u>	<u>0.00</u>	<u>0.00</u>	<u>P/F</u>	<u>L/F</u>

Report two decimal places. Round numbers ≤ 4 down, ≥ 5 up. ICV acceptance criteria $\pm 5\%$ or ± 0.05 m, whichever is greater.

COMMENTS:

Group: A Bottle Type: QPCR-P-500ML # of Bottles: 4 Preservative: ICE Enter Number of Bottles Sent to Lab 4

PCR-DG3

PCR-HF183

Group: A Bottle Type: BG-500ML # of Bottles: 4 Preservative: ICE Enter Number of Bottles Sent to Lab 4

W-E8321-DI

W-E8321-MS

Date of Request: 20-APR-2021
Created By: DAVIS_B On: 20-APR-2021
Modified By: JASROTIA_P On: 21-APR-2021
Customer: NE-DIST
Project: SMP
Division:
District: Northeast District
Event Description: bmap tribs

Send Coolers To:

Phone: 904-256-1700
8800 Baymeadows Way West
Suite 100
Jacksonville, FL 32256-7577
Attn: Jeremy Parrish
Cooler Ship EMail: Jeremy.M.Parrish@dep.state.fl.us

Priority: 3
Event Status: A
Criminal Investigation: NO
Chemistry Request Reviewed By: WATTS_J On: 21-APR-2021
Biology Request Reviewed By: JASROTIA_P On: 21-APR-2021
Sampling Kit Required: NO Ship on:
Sampling Kit Shipped:
Sampling Kit Packed By:
Preservatives Needed: NO
Date to Receive Samples: 20-APR-2021
Logged In By:

Send Final Report To:

8800 Baymeadows Way West
Suite 100
Jacksonville, FL 32256-7577
Attn: Jeremy Parrish
Report Notification EMail: Jeremy.M.Parrish@dep.state.fl.us

Comments: Group A: Source Track for Tracers and qPCR

Bottle Group A (Water) With 4 Samples:

Bottle Type: QPCR-P-500ML	Number of Bottles: 4	Preserved With: ICE
PCR-DG3	Template: DEFAULT	(SOP-PCR-1.5) Detection and quantification of canine-specific DG3 Bacteroides genetic marker with quantitative polymerase chain reaction
PCR-HF183	Template: DEFAULT	(SOP-PCR-1.0) Detection and quantification of human-specific HF183 Bacteroides genetic marker with quantitative polymerase chain reaction
Bottle Type: BG-500ML	Number of Bottles: 4	Preserved With: ICE
W-E8321-DI	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS
W-E8321-MS	Template: DEFAULT	(EPA 8321B) Pesticides, herbicides, and other chemicals in water matrices by HPLC/MS/MS

* - The laboratory is not NELAP certified for this analyte/method, or certification is not applicable.