Assessment Date: September 14, 2011

DOH Laboratory #: E86562

DEP Contract #: LAB035

Laboratory Name: Flowers Chemical Laboratory
571 NW Mercantile Pl, Suite 111
Port St. Lucie, FL 34986

I. Introduction

An onsite evaluation of the laboratory was conducted to verify compliance with the contract requirements of **DEP Contract No. LAB035**, which incorporates adherence to the quality control requirements in Chapter 5 of the NELAC Standards, Standard Methods 20th edition, section 9020 (SM9020), Chapter 62-160, Florida Administrative Code (F.A.C) and continued accreditation with Florida Department of Health under the National Environmental Laboratory Accreditation Program (NELAP) for fecal coliform and enterococci bacteria. The audit consisted of an introductory conference, discussion of laboratory operations, review of selected analytical records for parameters contracted by the FDEP, a walk-through of the laboratory facility, and an exit interview with key laboratory staff. It is noted that a complete review of the sample handling from receipt to data evaluation and delivery to FDEP was conducted.

II. Personnel

The following personnel participated in the laboratory audit:

FDEP Auditor	Title	Contact Information
Daisys Tamayo	Environmental Specialist II	850-245-8162
Michele Reliford	Environmental Supervisor II	850-245-8066

Flowers Chemical Laboratory

Flowers Chemical Laboratory	Title	Contact Information
June Flowers	Quality Systems Director	407-339-5984 x212
Don Braisted	Technical Advisor	772-343-8006

III. Evaluation and Observations

Upon arrival at the lab, FDEP staff met with June Flowers and Don Braisted. Information on the operations of the laboratory was provided by Don Braisted. It was determined that the education and technical experience of laboratory staff are adequate for the microbiological tests being conducted.

The facility, documentation, and procedures were inspected for the following methods covered under the current contract:

Analysis: Fecal Coliforms Method: SM9222D
Analysis: Enterococci Method: EPA 1600

Although the laboratory has been approved for the analyses of enterococci, the laboratory is not currently running this analysis under the current contract. Therefore, specific data were not available for review. However, general procedures, equipment, and logbooks for analysis were reviewed.

The laboratory was clean, uncluttered, and adequate for conducting the contracted parameters. No concerns or deficiencies were noted with respect to laboratory organization and cleanliness.

FDEP staff reviewed tracking records from sample receipt through sample disposal, data analyses and the electronic data deliverable records for FDEP reimbursement. No deficiencies were noted with respect to laboratory records.

Documentation for the following samples analyzed under contract was reviewed for completeness and accuracy. No deficiencies were noted with respect to specific sample data reviewed.

LIMS Job ID	Laboratory Report ID	Date Sampled	Date Analyzed	# of samples	Analysis
TLH-2011-04-18-09	147955	4/14/11	4/14/11	8	
TLH-2011-06-21-45	152436	06/20/2011	06/20/11	3	

IV. Deficiencies Noted and Corrective Action Required

No deficiencies were noted during the laboratory audit and corrective actions are not required. This audit report along with the on-site assessment evaluation checklist comprises the final report. A response to this report is not required.

V. Summary

Based on the information collected and reviewed, the on-site evaluation of the laboratory, and the review of the reports and electronic data deliverables evaluated during the Overflow Laboratory for Microbiological Analyses Audit on September 14, 2011 by the Florida Department of Environmental Protection Biology and QA Staff, Flowers Chemical Laboratory in Port St. Lucie, FL is generally meeting the conditions of the contract. Flowers Chemical Laboratory in Port St. Lucie is meeting the minimum criteria for data quality required by FDEP Contract #LAB035 and data reported under this contract is acceptable for use by FDEP.

Appendix A

Date: 09/14/2011

Lead Auditor:	Daisys Tamayo
Additional Auditors	Michele Reliford

Analysis	Method
Fecal Coliform	SM9222D
Enterococci	EPA 1600

Laboratory Name	Flowers Chemical Laboratories	Certification#	E86562
Primary Contact	June Flowers	Phone Number	(407) 333-5984 x212
Address:	571 NW Mercantile Pl., Suite 111, Port St. Lucie, FL 34986		
Email:	june@flowerslabs.com		

Position Title	Name	Phone	Email	Job Responsibilities
QA Director	June Flowers	(772) 343-8006	june@flowerslabs.com	Responsible for laboratory quality assurance and quality control
Technical Director	Don Braisted	(772) 343-8006	donbraisted@yahoo.com	Supervise microbiological section of laboratory

Reports to Review

Job ID	Report ID	Date Sampled	Date Analyzed	# of samples	Documents reviewed:
TLH-2011-04-18-09	147955	4/14/11	4/14/11	8	✓Custody sheet ✓Sample Receipt Log ✓Raw Data Log ✓Final Report ✓EDD
TLH-2011-06-21-45	152436	06/20/2011	06/20/11	3	✓Custody sheet ✓Sample Receipt Log ✓Raw Data Log ✓Final Report ✓EDD

Sample Receipt (NELAC 5.5.8)	
✓	Sample Acceptance Policy made available to sample collectors.
✓	Chlorine check performed on appropriate samples.
✓	Samples stored under refrigeration if not analyzed immediately.
Documentation:	
✓	Date/Time Received
✓	Sample Container Check
✓	Temperature Check (≤10 °C)
✓	Improperly preserved samples are “Y” qualified.
Additional comments:	

Laboratory Facility (NELAC D.3.8, SM9020B)	
✓	Walls, floors, and work surfaces are non-absorbent, and easy to clean and disinfect.
✓	Lab well ventilated and free of dust and drafts.
✓	Monitor air density (15 colonies/plate/15 min) and bench surface contamination.
✓	Lab space clean and free of clutter.
✓	Plants, food, and drinks are prohibited in the lab.
Additional comments:	

Media (NELAC 3.1, 3.6, SM9020B)	
✓	Sterility check run for each batch of prepared media or each lot of ready-to-use media.
✓	Positive control run for each batch of prepared media or each lot of ready-to-use media.
✓	Negative control run for each batch of prepared media or each lot of ready-to-use media.
✓	Open media bottles stored away from moisture and used within 6 months from open date.
✓	Expiration date (2 weeks for agar plates, loose cap tubes; 96 hrs for broth in screw cap flasks, 3 months for screw cap tubes.
Documentation:	
✓	pH check of media
✓	Media bottles used and amount
Additional comments:	

Equipment and supplies (NELAC D.3.8, SM9020B)	
Autoclave	
✓	Record contents, temp, sterilization time, total run time, and analyst for each cycle.
✓	Chemical steam indicator used for each cycle (autoclave tape).
✓	Biological (*Geobacillus stearothermophilus*) check performed monthly.
✓	Timer checked quarterly.
Waterbaths	
✓	Temp maintained at 44.5 ± 0.2° C for fecal coliforms. (Checked twice daily).
✓	Cover and/or circulation pump used to prevent evaporation and maintain even temp.
✓	Racks/weights used to prevent samples from floating.
✓	Waterbath emptied and cleaned periodically.
Incubators	
✓	Temp maintained at 41.0 ±0.5 °C for enterococci, 35.0± 0.5°C and 45.0 ±0.5°C for verification of fecal coliforms and enterococci.
✓	Thermometer submerged in water or glycerin.
✓	Samples not overcrowded and provided sufficient airflow when placed in incubator.
✓	Incubator cleaned and sterilized periodically.
pH meter	
✓	Meter graduated in 0.1 pH units or less and includes temperature compensation.
✓	Calibrated with 2 buffers that bracket pH being measured.
UV lamps	
NA	Lamps cleaned monthly with ethanol.
NA	Tested quarterly with a light meter (output >70%) or by biologicals (99% reduction).
Thermometers	
✓	Calibration checked at least annually with NIST certified thermometer.
✓	Calibration correction factors marked on thermometer as needed.
✓	Thermometer has appropriate graduated increments for use.
✓	Thermometer in good working condition and free of separations in liquid column.
✓	Temperature checked twice daily at least four hours apart for working incubators/water baths.
Glassware and Supplies	
✓	Check pH with bromothymol blue for each batch of washed glassware.
✓	Inhibitory residue test performed annually or after a change in detergent lot or washing procedure. (SM9020 requires test only with initial use of compound or change in procedure if BTB pH check is done consistently.
✓	Sterility checks performed with each lot of sample container, membrane filters and pads, and dilution water bottles (also require pH and volume check).
Laboratory Water	
✓	Run the following water quality checks (See Table 1):
Additional comments:	

Membrane Filtration (NELAC 3.1, SM9020B, SM9050C, SM9222D, EPA 1600)	
✓	Three dilutions analyzed per sample.
✓.	Control Blanks analyzed at beginning and end of each filtration series.
✓	Control blank run after every ten samples, unless UV sterilized.
✓	Duplicates analyzed on 10% of samples, at least one every batch.
✓	Phosphate buffered water used for rinse, dilutions, laboratory blanks. PBS may be used for EPA 1600.
Documentation:	
✓	Filtration time
✓	Incubation Start Time
✓	Incubation End Time
Additional comments:	

Verification (NELAC D.3.4, SM9020B, SM9222D, EPA 1600)	
✓	Verify at least 10 colonies from a positive sample per month (per Table 3).
✓	Verify atypical or ambiguous colonies of different morphological types to check for false negatives.
✓	Adjust colony count based on percentage of verified colonies.
✓	For samples that are subject to verification process as "Verified"
Additional comments:	

Demonstration of Capability (NELAC D.3.2, SM9020B)	
✓	Participates in a proficiency testing program (analyze PT's 2x/yr).
✓	Each analyst has documentation of demonstration of capability (initial and annually).
✓	Comparison of colony counts performed monthly. (<10% difference btwn 2 analysts, <5% for 1 analyst)
Additional comments:	

Data Handling (Chapter 62-160 F.A.C., SM9020B, SM9222D, EPA 1600)	
✓	Samples that are exceed holding time requirements are "Q" qualified.
✓	Samples that exceed incubation time limits are "J" qualified.
✓	Samples that are associated with a failed QC are qualified (blanks, replicate failures).
✓	Results are calculated per method and contract requirements.
✓	Ideal colony count range is 20-60 (fecal and enterococci) per membrane.
✓	Samples with no dilution in ideal range are "B" qualified.
✓	Calculation of MDL (dilution factor or min. reporting limit) based on acceptable dilutions.
✓	Control charts maintained for lab replicate samples and used to determine precision criteria.
✓	Data entry process includes a verification step to minimize transcription errors.
✓	Electronic data deliverable meets contract requirements.
Additional comments:	

Table 1. Laboratory Water Requirements

Analysis	Frequency	Exceptable Limits
pH	with each use (not required for Online Edition -05)	5.5-7.5
Conductivity	with each use	< 2.0 umho/cm
TRC	monthly	< 1.0 mg/L
HPC	monthly	<1000 CFU/mL
NH3, Organic N	monthly (not required for Online Edition -05)	<0.1 mg/L
TOC	monthly	< 1.0 mg/L
Use test	quarterly or for new water source	student's t < 2.78
Metals	Cd, Cr, Cu, Ni, Pb, Zn annually	each <0.05 mg/L, total <0.1 mg/L
Bacteriological ratio	annually (not needed for type II (medium) water quality or better, see Table 2)	0.8 - 3.0

Table 2. Water types per SM1080C

	High	Medium
Type	I	II
Resistivity (megaohm-cm)	> 10	> 1
Conductivity (umhos/cm)	< 0.1	< 1
SiO2, mg/L	< 0.05	< 0.1

Table 3: Verification Tests

Method **Verify with the following:**

Enterococci EPA 1600	BHI Agar Slant 35 ± 0.5°C 48 ± 3 h (use growth for gram staining)	BHI 35 ± 0.5°C 24 ± 2 h (turbidity)	Gram stain (gram positive cocci)	BHI 45 ± 0.5°C 48 ± 3 h (turbidity)	BEA 35 ± 0.5°C 48 ± 3 h (growth w/ black/brown precipitate)	BHI w/6.5% NaCl 35 ± 0.5°C 48 ± 3 h (turbidity)
Fecal Coliforms SM9222D	LTB 35 ± 0.5°C 48 ± 3 h (turbidity and gas)	EC 44.5 ± 0.2°C 24 ± 2 h (turbidity and gas)				
( ) Positive result for each test in parenthesis.						

Table 1

Reporting Requirements:

Field	Format	Required	Comments
Contract Lab Identifier	Varchar2(20)	X	DOH Certification Number of Contract Laboratory
LIMS Request ID	Varchar2(20)		Not used by our matching algorithm but potentially useful for reference
Field ID	Varchar2(50)	X	Bar-coded information (eventually). Might be Field-ID, sequestial number or a combination of both.
Site Location	Varchar2(80)		Sample collection location
STORET Station	Varchar2(20)		STORET Station Identifier
Contract Lab Sample ID	Varchar2(x)	X	Contract Lab Sample ID - not useful for us but potentially useful for refernce. No character limitation.
Date Sampled	mm/dd/yyyy	X	Date sample was collected.
Time Sampled	hh24:mm	X	Time of sample collection, in military format.
Date Received	mm/dd/yyyy	X	Date the sample was received by the contract laboratory.
Time Received	hh24:mm	X	Time received by the contract laboratory, in military format.
Analysis Start Date	mm/dd/yyyy	X	Date of placement of the processed sample into or on the applicable growth medium.
Analysis start Time	hh24:mm	X	The time of placement of the processed sample into or on the applicable growth medium, in military format.
Analysis End Date	mm/dd/yyyy	X	Date the sample was read by the contract laboratory.
Analysis End Time	hh24:mm	X	Time read by the contract laboratory, in military format.
Contract Lab Test ID	Varchar2(80)	X	Overflow lab's test ID - will be mapped to B/Labs Test ID
Contract Lab Compound	Varchar2(80)	X	Overflow laboratory's analyte name - will be mapped to B/Labs component name.
Units	Varchar2(10)	X	Overflow laboratory's units
Result	Varchar2(20)	X	Must be a numeric expression (no<, >, etc); Stored as test to preserve significant figures.
Qualifier		if appropriate	Must match those in the QA Rule (Chapter 62-160 F.A.C.); multiple qualifiers concatenated with no spaces or characters in between.
Dilution Factor	Varchar2(10)		Must be numeric expression.
MDL	Varchar2(10)		Must be numeric expression.
PQL	Varchar2(10)		Must be numeric expression.
Comments	Varchar2(80)		

Electronic Data Deliverables:

The Contractor must report the data in the electronic format described above unless both parties mutually agree otherwise.

The Electronic Data Deliverable must be a flat file with the field delimited by tabs. For the items that are not required and not available, the delimiters must still be included.