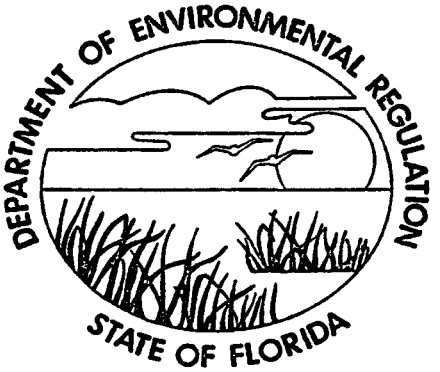


BIOASSAYS OF THE ST. ANDREWS WASTEWATER TREATMENT PLANT PANAMA CITY, BAY COUNTY, FLORIDA

NPDES #FL0020451

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OCTOBER 21, 1985

Biology Section
Division of Environmental Programs

BIOASSAYS OF THE ST. ANDREWS WASTEWATER TREATMENT PLANT PANAMA CITY, BAY COUNTY, FLORIDA NPDES #FL0020451

Biology Section
Bureau of Laboratories and
Special Programs
October 21, 1985

EXECUTIVE SUMMARY

St. Andrews Wastewater Treatment Plant, Panama City, NPDES #FL0020451, on 13 to 14 August, 1985.

The sample of final effluent collected from the St. Andrews Wastewater Treatment Plant proved to be acutely toxic to the test organisms, Daphnia pulex, Notropis leedsi, and Mysidopsis bahia. In the D. pulex and N. leedsi screening tests (100% concentrations only), all of the organisms died within 24 hours, resulting in 24 hour LC_{50}'s of < 100%. In the M. bahia definitive test, a 48 hour LC_{50} of 25.5% was calculated. A portion of this toxicity may be attributed to the residual chlorine (1.0 mg/l HACH and 1.06 mg/l iodometric) measured in the sample.

A second sample of effluent from this facility, in which the chlorine had been neutralized, was not toxic to the same test organisms, D. pulex, N. leedsi, and M. bahia. No significant mortality was noted in these tests. Moderate ammounts of ammounia (4.97 mg/l total and .011 mg/l unionized) were detected in the effluent which may have contributed slightly to the toxicity noted. Chemical analysis indicated the presence of several unidentified organic compounds in this effluent.

INTRODUCTION

The St. Andrews Wastewater Treatment Plant is located at 3601 W. 23rd Street, Panama City, Bay County, Florida (Fig. 1). The facility treats domestic wastewater from a population of approximately 17,000, and industrial wastes from textile mill. The wastewater treatment system consist of biological treatment using activated sludge with diffused air, secondary clarification, and chlorination (Fig. 2). Sludge is treated in aerobic digesters, filtered and dried (NPDES file). The facility has a design flow capacity of 3.5 MGD and discharges to St. Andrews Bay which connects with the Gulf of Mexico.

At the request of DER's Northwest District four screening tests and two 48 hour static acute toxicity bioassays were performed on samples of chlorinated and de-chlorinated effluents from the St. Andrews facility, from 13 to 15 August, 1985. The results of these tests are presented in this report. Those involved in performing the tests and in preparation of this report were: R. B. Frydenborg, M. L. Moultrie, and M. L. Roll.

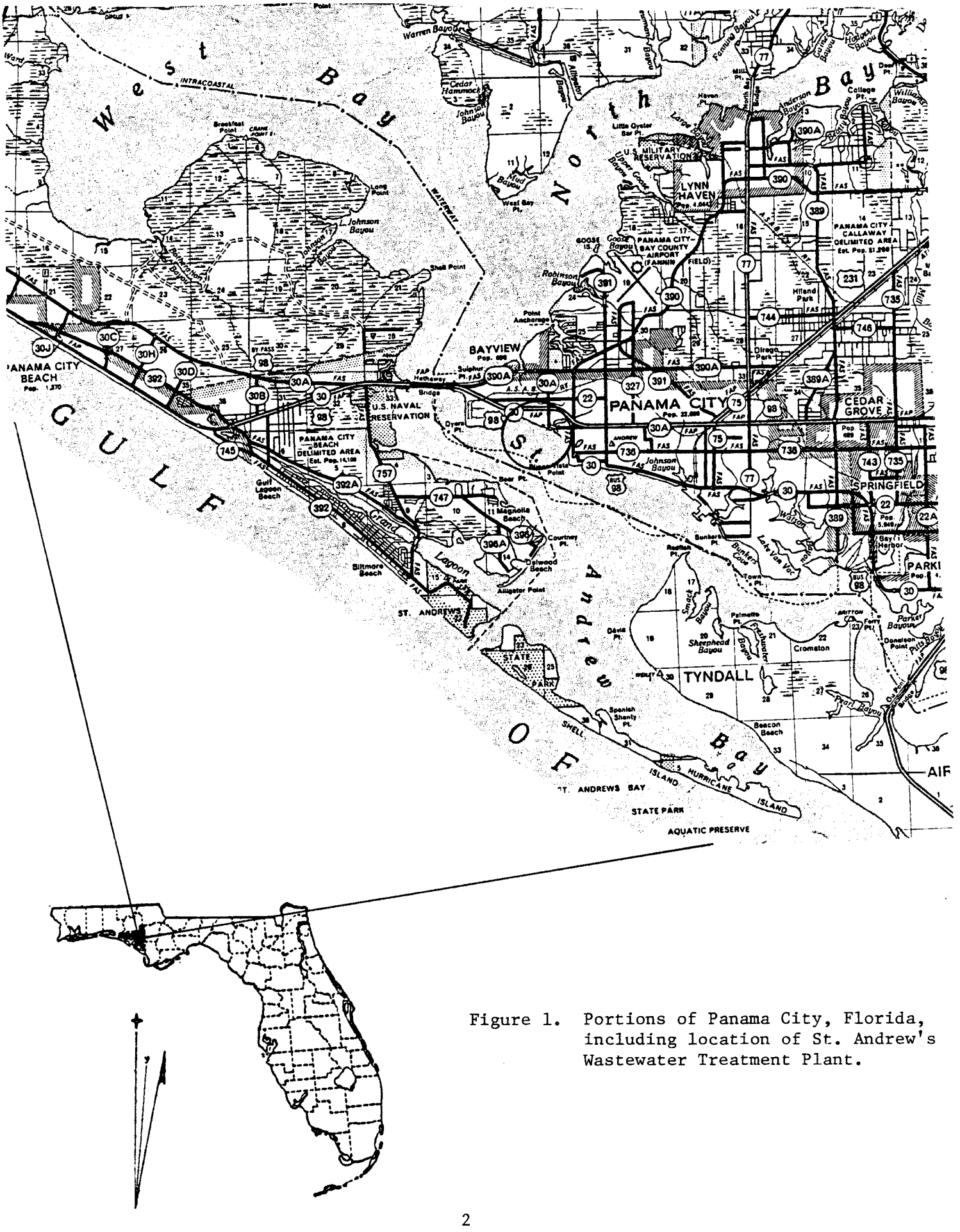


Figure 1. Portions of Panama City, Florida, including location of St. Andrew's Wastewater Treatment Plant.

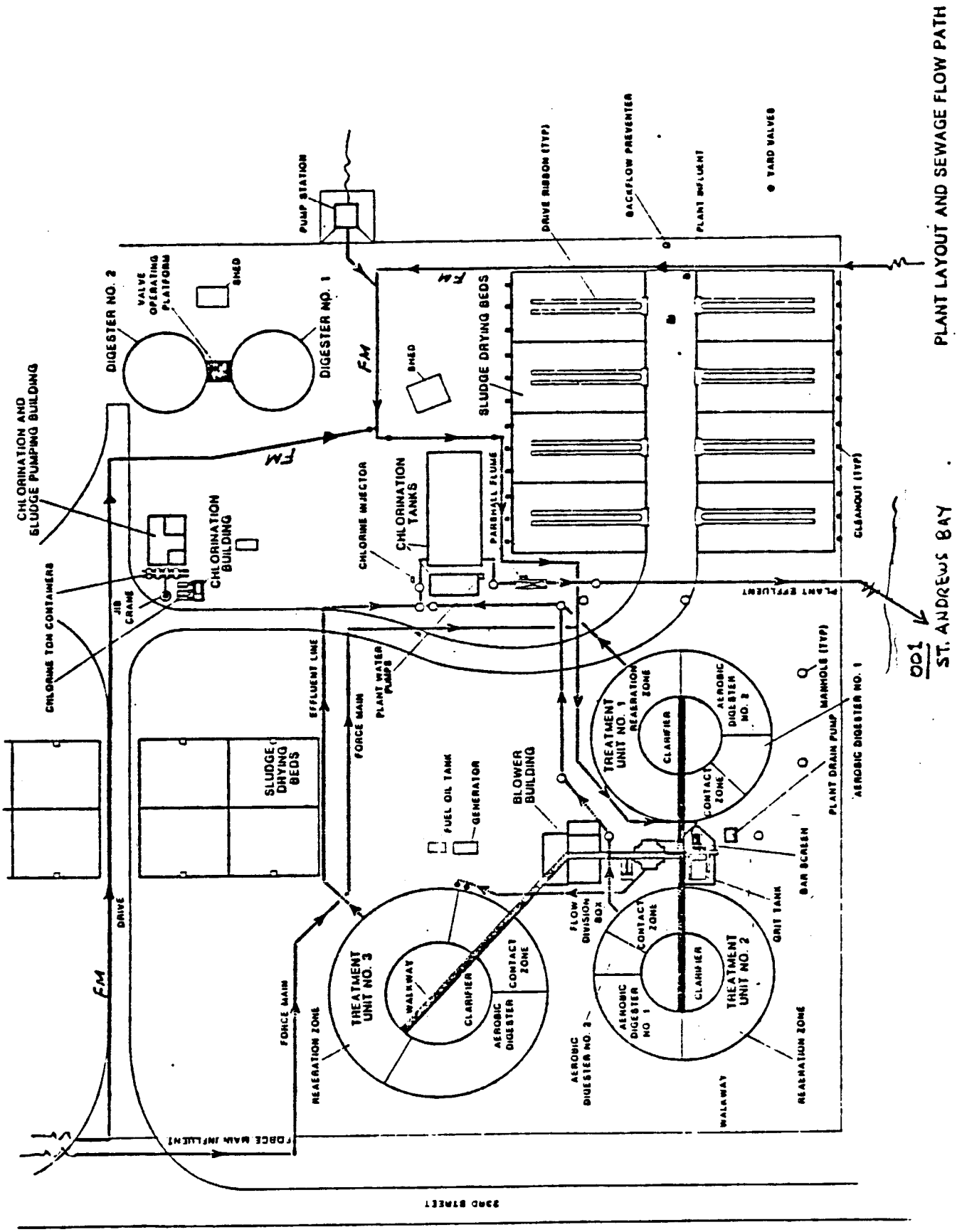


Figure 2. Schematic Diagram of St. Andrews Wastewater Treatment Plant.

METHODS AND MATERIALS

Test methods used to conduct these static bioassays are extensively described by Peltier and Weber (1985) and the FDER Biology Section (1985). The organisms selected for this study include species recommended by Peltier and Weber (1985), the U.S. EPA (1978), and the FDER Biology Section (1985). Additionally, the Biology Section maintains cultures of several species of organisms indigenous to the State of Florida. One of these indigenous species was also selected for testing. Life histories of these test organisms and maintenance procedures used to culture them are detailed below.

History and Maintenance of Bioassay Organisms

Daphnia pulex (water flea) - A breeding culture of Daphnia pulex was obtained from the EPA laboratory in Athens, Georgia, in March, 1979. This culture has been maintained since its arrival in 2 1/2 gallon all-glass aquaria, filled with unchlorinated DER well water, under a 16:8 hour light-dark cycle. Organisms are fed a liquified blend of Purina Trout Chow® and Cerophyl® twice daily, once daily on weekends, and are kept at a constant temperature of 22° ± 2°C. The aquaria are cleaned weekly by siphoning off half the water along with the detritus and excess organisms, and then refilling with fresh aerated well water. Additional measures are taken and/or new cultures started in the event of abnormal circumstances such as: declining populations, ephippia formation, or excessive fungal or bacterial growth.

Notropis leedsi (bannerfin shiner) - Approximately 40 adult bannerfin shiners were obtained from an indigenous population in the upper Suwannee River during October, 1981. The breeding stock is maintained in 30 and 125 gallon all-glass aquaria equipped with a flow-through water exchange system. Water for this system is unchlorinated and is pumped from the DER deep well. The bannerfin shiners are reared at 22° ± 2°C under a 16:8 hour light-dark cycle and are fed Tetra-Min® and frozen Artemia (brine shrimp) twice daily, and once daily on weekends. Breeding and egg hatching are encouraged at 25°C. The eggs are laid between stacked red ceramic tiles separated by a 3mm bead of silicone. Most of the eggs adhere to the tiles and are easily removed to heated, aerated hatching tanks. The narrow space between the tiles eliminates egg predation by adult fish. New hatching tanks are started regularly to segregate fry into even age groups. Newly hatched fry are fed a liquefied blend of Purina Trout Chow® and Cerophyl® twice daily. After one week, feeding is augmented with newly hatched Artemia nauplii and finely crushed Tetra Min®. The fish used in this test were 3 to 4 weeks old at the time of the test.

Mysidopsis bahia (mysid shrimp) - In September, 1978, approximately 300 adult mysid shrimp were obtained from the EPA laboratory in Gulf Breeze, Florida. The mysids are cultured under a 16:8 hour light-dark cycle in 10 and 15 gallon all-glass aquaria equipped with undergravel filtration and aeration. The mysid shrimp are cultured over a wide range of salinities (15-30 ppt) to minimize acclimation periods to low or high salinity effluents. Seawater, used for culturing the test organisms and as diluent in the bioassays, is prepared by mixing artificial sea salts (Forty Fathoms®) and well water to obtain a final salinity of 20-30 ppt. Freshwater, for dilution of the seawater as necessary, is obtained from a deep well near the DER laboratory. The mysids are fed twice daily and once daily on weekends with freshly hatched Artemia (brine shrimp) and are kept at a constant temperature of 23° ± 2°C. Adult or large juvenile mysids were used in this test.

To ensure the uniform sensitivity of these test organisms to toxic substances, the Biology Section conducts monthly standard reference toxicant bioassays using sodium lauryl sulfate as the reference toxicant.

Test Methods

Methods used to perform acute toxicity bioassays are described by Peltier and Weber (1985) and the FDER Biology Section (1985). Some of the methods described below are more specific than those of Peltier and Weber. These will be noted in more detail.

Sampling Methods and Test Dates

A total of four screening tests and two definitive 48 hour static acute toxicity tests were performed on chlorinated and de-chlorinated samples of effluent collected from the St. Andrew's Wastewater Treatment Plant. The tests were performed from 1010 hours on 13 August to 1020 hours on 15 August, 1985.

The sample was collected by Mr. Robert Reining Jr. of DER's Panama City Branch office at 1200 hours on 12 August, 1985, using standard wastewater sampling procedures. Separate samples were obtained for chemical and biological testing. Samples for metal analyses were preserved with HNO_3. Samples for organic chemical analyses and for the bioassays were unpreserved. The samples were iced and transported to the DER laboratory in Tallahassee for analyses within 24 hours of collection.

Test Temperature

Upon arrival of the iced (4°C) bioassay samples in the Tallahassee laboratory, the containers were placed in warm water and the temperature of the samples allowed to rise to 20° $\pm$ 2°C. Although this temperature may be different than the temperature of the sample during collection, it is the temperature range to which the test organisms are acclimatized and is the range recommended in the above referenced bioassay protocols. This avoids stressful effects caused by exposure to unusually cold or warm samples.

Salinity Adjustment

Since this particular bioassay sample is discharged to a marine environment, the Biology Section used a saltwater test organism in the definitive tests to evaluate toxicity. Therefore, artificial sea salts (Forty Fathoms®) were added to portions of the chlorinated and de-chlorinated effluents until salinities of approximately 18 ppt were obtained. This salinity is well within the salinity tolerance of the test organism. A Control was also employed, consisting of artificial seawater at a similar salinity of approximately 20 ppt. Additional information concerning salinity adjustment and saltwater tests is given below.

pH Adjustment

Since the toxicity, or lack of toxicity, in a bioassay sample may be dependent on the sample's pH, the Biology Section prefers, and recommends, that the pH of a sample at the start of the bioassay be similar to the pH measured during sampling. In this particular case, the salinity adjusted samples used in the M. bahia definitive tests were adjusted with an acid (0.1N HCl). This was necessary due to the pH having increased between sampling and initiating the test. This pH increase was likely due to the salinity adjustment. The pH of the saltwater samples were adjusted to ± 0.3 s.u. of the value measured at sampling. This procedure provides the fairest test possible considering the varied chemical substances in complex wastewaters, the toxicities of which are dependent on pH.

Total Residual Chlorine

Since many wastewaters are chlorinated, especially those originating from sewage treatment facilities, it is often desirable to differentiate between the toxicity resulting from chlorine and the toxicity associated with other components of the wastewater. To accomplish this, the Biology Section performed concurrent bioassays on a sample of chlorinated wastewater in which the chlorine had been neutralized. De-chlorination was accomplished by adding a calculated amount of 0.025N sodium thiosulfate. The total residual chlorine concentrations of the tested effluents are given on the bioassay data forms. A comparison of the results of these tests will distinguish between the toxicity due solely to residual chlorine and the toxicity inherently contained in the wastewater.

Freshwater Screening Tests

Freshwater for use in the controls was obtained from the same DER deep well that is used to culture the test organisms.

Two replicates, of 100% effluent and a control, were used to test with the water flea, Daphnia pulex. Flint glass jars of 250 ml capacity were used as test chambers, and 200 ml of the appropriate test concentration was placed in each chamber. Daphnia were siphoned into a large culture dish to isolate the proper size and age organisms (neonates). To accomplish this, the water fleas were siphoned through a stainless steel screen. The trapped adults were then discarded. These neonates were then transferred by glass pipette, examined for injury, and were loaded 10 per chamber, resulting in 20 per test concentration. Water fleas were fed during these tests.

Two replicates, of 100% effluent and a control, were used for the Notropis leedsi (bannerfin shiner) bioassays. The fish were transferred individually or by twos, into wide mouth quart jars containing 500 ml each of the appropriate test concentration, and were loaded 5 per chamber, resulting in 10 per concentration. Fish were not fed during these tests.

Saltwater Definitive Tests

Saltwater for dilution of the effluent samples was prepared from artificial sea salts (Forty Fathoms®), and adjusted with DER well water until a salinity of approximately 20 ppt was achieved. As discussed previously, the samples used for these tests were adjusted with the same artificial sea salts until salinities of 18 ppt were achieved. Therefore, the final salinities in these tests ranged from approximately 18 ppt to 20 ppt.

The concentration series selected for this study (Table 1) is an abbreviated version of that recommended by APHA (1980). Two replicates of each test concentration and of the control were used to test with Mysidopsis bahia. Immediately prior to the bioassays, a suitable number of mysids were transferred by net from the appropriate salinity aquarium to a culture dish for observation and selection of individuals for the tests. Individual mysid shrimp were siphoned using a glass tube, carefully examined for injury, and transferred to wide mouth quart jars, each containing 500 ml of the appropriate concentration. For each concentration, the mysid shrimp were pre-acclimated to within 2 ppt salinity. Five organisms were loaded per test chamber, giving ten per concentration. Based on the recommendations of EPA (1978), the mysids were fed brine shrimp (Artemia) nauplii at the rate of approximately 10 to 20 nauplii per mysid per day to minimize cannibalism.

Dissolved oxygen (YSI Model 57 oxygen meter), pH (Corning Model 7 pH meter with Orion Ross® combination electrode), temperature (NBS calibrated glass thermometer), and the number of live organisms were recorded for each test concentration at 0 hours, 24 hours, and 48 hours. Total alkalinity and total hardness (measured as $CaCO_3$ using HACH Chemical Company reagents) were recorded at the beginning of the test for the controls and the 100% effluent concentration. The specific conductance or salinity (YSI Model 33 S-C-T meter) of each test concentration was recorded at the beginning and end of the test. All instruments were calibrated daily according to manufacturer's recommendations. Ammonia concentrations, including the total and unionized fraction (expressed in mg/l as N and as NH_3, respectively), were recorded in the controls and the 100% effluent test concentrations at the beginning of the test. Total ammonia concentrations were determined using an Orion® specific ion probe and an Orion Ionalyzer®. The unionized fraction was then calculated based on the total ammonia concentration, and the pH, temperature, and salinity of the sample. These values were recorded on the bioassay data forms and are presented in Tables 2 through 7.

Table 1. Concentration Series and Volumes Used for These Static Bioassays.

Daphnia pulex (water flea) Screening test only

Concentration	Dilution Water	Effluent	Total Volume
Control	200.0 ml	--------	200 ml
100.0%	--------	200.0 ml	200 ml

Notropis leedsi (bannerfin shiner) Screening test only

Concentration	Dilution Water	Effluent	Total Volume
Control	500 ml	--------	500 ml
100.0%	-------	500 ml	500 ml

Mysidopsis bahia (mysid shrimp)

Concentration	Dilution Water	Effluent	Total Volume
Control	500 ml	--------	500 ml
10.0%	450 ml	50 ml	500 ml
32.0%	340 ml	160 ml	500 ml
56.0%	220 ml	280 ml	500 ml
100.0%	------	500 ml	500 ml

Daphnia pulex

Table 2. St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida, NPDES #FL0020451 on 13 to 14 August, 1985.
Chlorinated sample[a]

Concentration or %	Number of Live Organisms			Dissolved Oxygen (mg/l)			pH			Temperature (°C)			Conductivity (umhos)		
	0	24	48	0	24	48	0	24	48	0	24	48	0		48
Control	10	10		7.9	8.0		7.5	7.7		22.0	19.7				
Control	10	10		8.0	8.0		7.5	7.7					210		
100.0	10	0		8.1	8.6		6.6	7.4		22.5	19.5				
100.0	10	0		8.1	8.7		6.6	7.4					425		

Control Water

NH_3(as NH_3)(mg/l) 0

Total Alkalinity 92 (mg/l $CaCO_3$)

Total ammonia (as N)(mg/l) 0

Total Hardness 105 (mg/l $CaCO_3$)

100% Effluent

NH_3(as NH_3)(mg/l) .011

Total Alkalinity 45 (mg/l $CaCO_3$)

Total ammonia (as N)(mg/l) 4.97

Total Hardness 75 (mg/l $CaCO_3$)

[a] Total Chlorine residual: 1.0 mg/l (HACH) and 1.06 mg/l (Iodometric).

Notropis leedsi

Table 3. St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida NPDES #FL0020451 on 13 to 14 August, 1985.

Chlorinated sample[a]

Concentration or %	Number of Live Organisms 0	24	48	Dissolved Oxygen (mg/1) 0	24	48	pH 0	24	48	Temperature (°C) 0	24	48	Conductivity (umhos) 0	48
Control	5	5		8.2	8.3		7.5	7.7		22.2				
Control	5	5		8.2	8.3		7.5	7.7					210	
100.0	5	0		8.4	8.7		6.6	7.3		22.8				
100.0	5	0		8.5	8.7		6.6	7.3					419	

Control Water

NH_3(as NH_3)(mg/l): 0

Total ammonia (as N)(mg/l): 0

Total Alkalinity (mg/l $CaCO_3$): 92

Total Hardness (mg/l $CaCO_3$): 105

100% Effluent

NH_3(as NH_3)(mg/l): .011

Total ammonia (as N)(mg/l): 4.97

Total Alkalinity (mg/l $CaCO_3$): 45

Total Hardness (mg/l $CaCO_3$): 75

[a] Total Chlorine residual: 1.0 mg/l (HACH) and 1.06 mg/l (Iodometric).

Mysidopsis bahia

Table 4. St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida, NPDES #FL0020451 on 13 to 15 August, 1985.

Chlorinated sample[a]

Concentration or %	Number of Live Organisms			Dissolved Oxygen (mg/l)			pH			Temperature (°C)			Salinity (ppt)	
	0	24	48	0	24	48	0	24	48	0	24	48	0	48
Control	5	4	4	8.4	6.8	6.0	8.5	8.5	8.3	21.3	19.9	19.3		
Control	5	5	5	8.4	6.8	6.2	8.5	8.5	8.3				20.0	20.5
10.0%	5	3	3	8.4	7.0	6.6	8.4	8.5	8.3	21.3	19.4	19.0		
10.0%	5	5	5	8.4	7.2	6.7	8.4	8.5	8.3				20.0	20.0
32.0%	5	2	2	8.4	7.5	6.7	8.1	8.3	8.2	21.8	19.3	18.9		
32.0%	5	5	4	8.4	7.5	6.7	8.1	8.3	8.2				19.5	19.5
56.0%	5	0	-	8.4	7.6	-[b]	7.4	8.0	-[b]	22.1	19.1	-[b]		
56.0%	5	0	-	8.4	7.7	-[b]	7.4	8.0	-[b]				19.0	-[b]
100.0%	5	0	-	8.4	7.8	-[b]	6.6	7.7	-[b]	23.0	19.4	-[b]		
100.0%	5	0	-	8.4	7.7	-[b]	6.6	7.7	-[b]				18.0	-[b]

Control Water	
NH_3(as NH_3)(mg/l) 0	Total Alkalinity ____ (mg/l $CaCO_3$)
Total ammonia (as N)(mg/l) 0	Total Hardness ____ (mg/l $CaCO_3$)

100% Effluent	
NH_3(as NH_3)(mg/l) 0.10	Total Alkalinity ____ (mg/l $CaCO_3$)
Total ammonia (as N)(mg/l) 4.97	Total Hardness ____ (mg/l $CaCO_3$)

[a] Total Chlorine residual: 1.0 mg/l (HACH) and 1.06 mg/l (Iodometric).

[b] Remaining parameters were not recorded since all organisms were dead by 24 hours

Daphnia pulex

Table 5. St. Andrews Wastewater Treatment Plant, Panamy City, Bay County, Florida, NPDES #FL0020451 on 13 to 15 August, 1985.

De-Chlorinated sample[a]

Concentration or %	Number of Live Organisms			Dissolved Oxygen (mg/l)			pH			Temperature (°C)			Conductivity (umhos)	
	0	24	48	0	24	48	0	24	48	0	24	48	0	48
Control	10	10	10	7.9	8.0	7.8	7.5	7.7	7.9	22.0	19.7	19.1		
Control	10	10	10	8.0	8.0	7.8	7.5	7.7	7.9				210	210
100.0%	10	10	10	7.6	7.6	7.3	6.8	7.5	7.5	21.5	20.0	19.0		
100.0%	10	10	10	7.5	7.6	7.3	6.8	7.5	7.5				425	428

Control Water

NH_3(as NH_3)(mg/l) 0

Total Alkalinity 92 (mg/l $CaCO_3$)

Total ammonia (as N)(mg/l) 0

Total Hardness 105 (mg/l $CaCO_3$)

100% Effluent

NH_3(as NH_3)(mg/l) 0.17

Total Alkalinity 45 (mg/l $CaCO_3$)

Total ammonia (as N)(mg/l) 4.97

Total Hardness 75 (mg/l $CaCO_3$)

[a] Total Residual Chlorine: < 0.04 mg/l (HACH).

Notropis leedsi

Table 6. St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida, NPDES #FL0020451 on 13 to 15 August, 1985.
De-Chlorinated sample[a]

Concentration or %	Number of Live Organisms			Dissolved Oxygen (mg/1)			pH			Temperature (°C)			Conductivity (umhos)	
	0	24	48	0	24	48	0	24	48	0	24	48	0	48
Control	5	5	4	8.2	8.3	b	7.5	7.7	7.8	22.2	19.7	19.0		
Control	5	5	5	8.2	8.3	b	7.5	7.7	7.8				210	210
100.0%	5	5	5	7.6	7.6	7.3	6.7	7.4	7.5	21.5	19.7	19.9		
100.0%	5	4	4	7.8	7.8	7.3	6.7	7.4	7.5				430	432

Control Water

NH_3(as NH_3)(mg/1): 0

Total Alkalinity (mg/1 $CaCO_3$): 92

Total ammonia (as N)(mg/1): 0

Total Hardness (mg/1 $CaCO_3$): 105

100% Effluent

NH_3(as NH_3)(mg/1): 0.13

Total Alkalinity (mg/1 $CaCO_3$): 45

Total ammonia (as N)(mg/1): 4.97

Total Hardness (mg/1 $CaCO_3$): 75

[a] Total Residual Chlorine: < 0.04 mg/1(HACH)

[b] Data not recorded

Mysidopsis bahia

Table 7. St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida NPDES #FL0020451 on 13 to 15 August, 1985.

De-Chlorinated sample[a]

Concentration or %	Number of Live Organisms			Dissolved Oxygen (mg/l)			pH			Temperature (°C)			Salinity (ppt)		
	0	24	48	0	24	48	0	24	48	0	24	48	0	48	
Control	5	5	5	8.4	6.8	7.0	8.5	8.5	8.4	21.1	19.6	19.2			Control Water
Control	5	5	5	8.4	6.8	7.0	8.5	8.5	8.4				20.0	20.0	NH_3(as NH_3)(mg/l) 0 — Total Alkalinity (mg/l $CaCO_3$)
10.0%	5	5	5	8.4	6.8	7.2	8.4	8.4	8.3	21.0	19.3	19.0			
10.0%	5	5	5	8.5	6.8	7.2	8.4	8.4	8.3				20.0	20.0	
32.0%	5	5	5	8.4	6.6	6.6	8.1	8.2	8.2	21.1	19.2	19.0			Total ammonia (as N)(mg/l) 0 — Total Hardness (mg/l $CaCO_3$)
32.0%	5	5	5	8.3	6.8	6.6	8.1	8.2	8.2				19.5	19.3	
56.0%	5	5	5	8.4	6.6	6.5	7.5	7.9	8.0	21.4	19.2	18.9			
56.0%	5	5	5	8.4	6.6	6.5	7.5	7.9	8.0				19.0	19.0	
100.0%	5	4	4	8.1	6.8	6.5	6.6	7.6	7.8	22.0	19.3	19.3			
100.0%	5	5	5	8.1	6.9	6.5	6.6	7.6	7.8				18.0	18.0	100% Effluent
															NH_3(as NH_3)(mg/l) .008 — Total Alkalinity (mg/l $CaCO_3$)
															Total ammonia (as N)(mg/l) 4.43 — Total Hardness (mg/l $CaCO_3$)

[a] Total Residual Chlorine: < 0.04 mg/l (HACH)

Table 8. LC_{50} Data for Bioassays of St. Andrews Wastewater Treatment Plant, Panama City, Bay County, Florida, NPDES #FL0020451 on 13 to 15 August, 1985.

Organism	Method of Data Analysis	24 Hour LC_{50} and 95% Confidence Intervals	48 Hour LC_{50} and 95% Confidence Intervals
Chlorinated sample:			
Dapnnia pulex		100.0%[a]	———
Notropis leedsi		100.0%[a]	———
Mysidopsi bahia	Binomial	36.3% (0 and 56)	34.3% (0 and 56
	Moving Average	27.9% (18.9 and 41.4)	25.5% (16.9 and 37.0), b
	Probit	25.4% (0 and 00)	23.5% (13.6 and 34.2)
De-chlorinated sample:			
Daphnia pulex		No Mortality	No Mortality
Notropis leedsi		No Mortality	No Mortality
Mysidopsis bahia	Binomial	c	c
	Moving Average		
	Probit		

[a] Since all organisms were dead in the 100% screening test, the LC_{50} is < 100.0%.

[b] This LC_{50} is reported in the text since it is the most statistically conservative value of the data set.

[c] Due to an insufficient number of dead organisms, or to the irregular distribution of mortality, an LC_{50} value cannot be calculated using this method of statistical analysis.

Table 9. Results of Chemical Analyses Performed on Effluent from the St. Andrews Wastewater Treatment Plant, NPDES #FL0020451, collected on 12 August, 1985.

Organics

Acid Extractables:	N.D.
Pesticide Extractables:	Unidentified Compounds
Base Neutral Extractables:	N.D.

Metals

Antimony	N.D.
Arsenic	< 7 ug/l[a]
Beryllium	N.D.
Cadmium	N.D.
Chromium	N.D.
Copper	N.D.
Lead	N.D.
Mercury	N.D.
Nickel	N.D.
Selenium	N.D.
Silver	< 0.5 ug/l[a]
Thallium	N.D.
Zinc	____[b]

N.D. = None Detected
[a] Actual value is known to be less than value given.
[b] Zinc analysis was invalidated.

RESULTS AND DISCUSSIONS

The sample of final effluent collected from the St. Andrews Wastewater Treatment Plant proved to be acutely toxic to the test organisms, Daphnia pulex, Notropis leedsi, and Mysidopsis bahia. In the D. pulex and N. leedsi screening tests (100% concentrations only) all of the organisms died by 24 hours, giving LC_{50}'s of 100% (Table 2 and 3). In the M. bahia definitive test, a 48 hour LC_{50} of 25.5% was calculated (Table 4). The 96 hour LC_{50}'s for the above organisms would therefore be less than or equal to the 24 and 48 hour LC_{50} values given. A portion of this toxicity may be attributed to the residual chlorine (1.0 mg/l HACH and 1.06 mg/l Iodometric) measured in the sample.

A second sample of effluent from this facility, in which the chlorine had not been neutralized, was not toxic to the same test organisms (D. pulex, N. leedsi, and M. bahia). No significant mortality was noted in these tests. Moderate amounts of ammonia (4.97 mg/l total and .011 mg/l unionized) were detected in the effluent, which may have contributed only slightly to the toxicity noted. Chemical analyses indicated the presence of several unidentified organic compounds in this effluent (Table 9).

Discussion of the Toxicity Criteria

The toxicity standards of Chapters 17-3 and 17-4, F.A.C., are complex. Because of this complexity, they will be briefly reviewed here so that these test results may be more readily compared with the standards. For legal purposes, however, the language of the Florida Administrative Code should be consulted directly.

The two basic concepts utilized in the standards are acute and chronic toxicity (defined in 17-3.021(1) and (4), respectively). Simplifying the actual F.A.C. language somewhat, acute toxicity is defined as the presence of a substance in a concentration one-third as great as that concentration which kills one-half of the test organisms in 96 hours, i.e., one-third of the 96 hour LC_{50}. Chronic toxicity is similarly defined as one-twentieth of the 96 hour LC_{50}. Therefore, if a test solution kills one-half or more of a group of test organisms in 96 hours, it must contain at least three times the defined "acutely toxic" concentration and at least twenty times the "chronically toxic" concentration of some substance or substances. If a test solution causes that same mortality in less than 96 hours, (for example, in 24 or 48 hours), it is at least as toxic as the example given above, and actually is probably considerably more toxic.

The particular restrictions applied to a given discharger depend upon the existence of a mixing zone, whetheras a specific part of the permit or based on the existence of a currently valid permit predating the language in Chapter 17-4 requiring mixing zone designation. In the absence of a mixing zone, the general criteria for surface waters [17-3.061] are applicable at all points, beginning at the point of discharge. This section prohibits the discharge of substances in concentrations which are chronically toxic [17-3.061(2)(p)]; that is, the concentrations cannot exceed one-twentieth of the 96 hour LC_{50} as discussed above. If a mixing zone does exist, then this chronic toxicity standard is only applicable from the edge of the mixing zone and beyond. Within the mixing zone, the less stringent acute toxicity standard of the minimum criteria for all waters [17-3.051(1)(d)] applies: one-third of the 96 hour LC_{50}. The acute toxicity standard does not apply, however, to each point within a mixing zone not designated for dredge and fill permitting, but rather to the average of a number of points within such a mixing zone, based on the provisions of the surface water mixing zones section of Chapter 17-4 [17-4.244(4)]. At the same time, though, this same rule prohibits waste concentrations within such a mixing zone (i.e., at any point within the mixing zone), from exceeding the 96 hour LC_{50}. Therefore, at any point within such a mixing zone, including the point of discharge, one-half of the test organisms cannot be killed in 96 hours. Within a mixing zone designated for dredge and fill permitting, the acute toxicity standard (one-third of the 96 hour LC_{50}) applies at all points, including the point of discharge.

From the above discussion, it should be clear that if the least stringent toxicity standard, the 96 hour LC_{50}, is exceeded at any point, from the point of discharge outwards, by any discharger, then a violation of the standards has occurred. This is an important consideration since the test procedure here reported involves the measurement of the 50% lethal concentration, and not some fraction of that concentration. Any discharger required to meet the one-twentieth of the 96 hour LC_{50} standard is therefore being tested very leniently. Any reduction in the test period from 96 hours provides an added degree of leniency to a discharger.

A number of different dilutions were examined in order to determine by how much, if any, the discharge exceeds the 96 hour LC_{50}. For example, if the sample, after two-fold dilution with diluent (a 33% concentration), kills one-half of the test organisms after 96 hours, then the sample is three times as toxic as is allowed by that standard. Similarly, if a 5% solution of the sample kills 50% of the organisms, the sample is twenty times as toxic as is allowed. Toxicities are reported here in that manner, i.e., in terms of the percentage dilution of the test sample. The smaller the percentage which is reported means a correspondingly greater toxicity.

Note that even if a discharger receives a very large mixing zone not designated for dredge and fill permitting, the regulations require that the maximum toxicity at any point within the mixing zone be restricted to the 96 hour LC_{50}. The results here reported are based on actual effluent samples, representative of the effluent concentration which enters waters of the state at the point of discharge. If it is demonstrated that, under the conditions existing at the point of discharge, effluent concentrations will be less than at the actual sampling point (if the two points are different), the reported bioassay results will reflect the reduced discharge concentration.

With respect to the species of organisms used in these tests, the language of the F.A.C. indicates that they should be "significant to the indigenous aquatic community". All species used in these tests are significant to the indigenous aquatic community since they are sensitive to toxic substances which may be expected to impact the indigenous community.

The toxicity reported here is in excess of allowable limits.

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