DETERMINATION OF TRUE COLOR IN WATER USING THE AQ2 OR AQ400 DISCRETE ANALYZER

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1. SCOPE AND APPLICATION

The test covered by this SOP is W-COLOR. This SOP is based on our TNI accredited method, "DEP SOP NU-094-1.14/UV-Vis (2022)" and is applicable to waters and wastes that have color characteristics derived from naturally occurring materials.

Calibration range	0 – 150 PCU
Method Detection Limit (MDL)	2.0 PCU
Practical Quantitation Limit (PQL)	6.0 PCU

Samples containing true color levels higher than 150 PCU are diluted prior to analysis.

2. SUMMARY OF THE METHOD

2.1. True sample color is determined spectrophotometrically at a wavelength of 450 nm. Particulate matter is removed prior to color determination. Platinum cobalt standards are used for quantification. One unit of color is that produced by 1 mg/L platinum in the form of the chloroplatinate ion.

2.2. The basic principles of this method are consistent with SM 2120 C-2011.

2.3. Interferences: Turbidity and the presence of colloidal and suspended particles will interfere with this analysis. All samples analyzed are lab filtered before true color is measured. The results are pH dependent. Highly colored industrial wastes may have hues far different from those of the platinum-cobalt standards. Their color characteristics may not be derived from naturally occurring materials such as humic acids.

3. EQUIPMENT AND SUPPLIES

3.1. Instrumentation

Component	Manufacturer	Model
Discrete Analyzer	SEAL Analytical	AQ2
Discrete Analyzer	SEAL Analytical	AQ400
pH Meter	Fisher Scientific or equivalent	Accumet AE150 or equivalent

3.2. Equipment

Equipment	Manufacturer/Vendor[1]	Model/Catalog Number
Adjustable Pipette, 100–1000 µL	Eppendorf/Fisher Scientific	Reference 2, 4924000088
Adjustable Pipette, 500-5000 µL	Eppendorf/Fisher Scientific	Research Plus, 3123000071
Pipette, Volumetric, Glass, 0.5 mL	Fisher Scientific (SR)	13-650-2A
Pipette, Volumetric, Glass, 3 mL	Fisher Scientific (SR)	13-650-2D
Pipette, Volumetric, Glass, 5 mL	Fisher Scientific (SR)	13-650-2F
Pipette, Volumetric, Glass, 10 mL	Fisher Scientific (SR)	13-650-2L
Pipette, Volumetric, Glass, 20 mL	Fisher Scientific (SR)	16-650-SH

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Pipette, Volumetric, Glass, 40 mL	Fisher Scientific (SR)	13-650-2R
Pipette, Volumetric, Glass, 75 mL	Fisher Scientific (SR)	13-650-2T
Volumetric Flask, Class A, Assorted	Fisher Scientific (SR)	10-212B, 10-210-5D, 10-212BB, 10-212A, 10-240B
Or Equivalent		
[1]**Note:** (SR) designates equipment is typically ordered through the in-house stockroom		

3.3. Supplies/Consumables

Supplies/Consumables	Vendor[1]	Catalog Number
Cup, Sample, 1.2	SEAL Analytical (SR)	5385
Reaction Segments	SEAL Analytical (SR)	5000
Pipette tips. Eppendorf, 1000 µL	Fisher Scientific (SR)	05-403-43
Pipette tips, Eppendorf, 5000 µL	Fisher Scientific	05-403-62
Bottle, Wash, 500 mL	Fisher Scientific (SR)	03-409-22C
Sample Beakers, Polystyrene, 20 mL	Fisher Scientific (SR)	08-732-120
Paper, pH, 0.0-6.0	Fisher Scientific (SR)	14-850-114
Paper, pH, 5.0-9.0	Fisher Scientific (SR)	14-850-113
Reagent Containers	SEAL Analytical	5010
Pipette, Transfer, Polyethylene, 5 mL	Fisher Scientific (SR)	13-711-5AM
Syringe, Polypropylene, 5 cc	Fisher Scientific (SR)	22-204-500
Filters, Syringe, 25mm (0.45 µm Filters)	Fisher Scientific (SR)	05-713-404A
Cleaning Swabs (20/pk)	SEAL Analytical	AQCSW-20
Lube Pen	SEAL Analytical	AQLP
Peristaltic Pump Tubing	SEAL Analytical	5140
Or Equivalent		
[1]**Note:** (SR) designates supply is typically ordered through the in-house stockroom		

4. REAGENTS AND STANDARDS (SEE DEP SOP NU-042)

Note: Before preparing standards or running samples, please read the safety notes in the safety/hazardous waste section of this SOP and the MSDS for the chemicals used in this analysis.

Note: Standard preparations must be documented in the Laboratory Information Management System (LIMS) 'Standard Preparation Tracker' module. Unique ID numbers for each standard are generated in the module and documented in the Reagent and Standard Logsheet (NU-RGTSTD). The NU-RGTSTD template is located on the network at \\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RunData\MetaData\Color\Forms\. This excel sheet needs to be updated and submitted as part of the analysis report folder. If available, a Certificate of Analysis for each purchased standard or reagent is also maintained in the MetaData\CertificatesofAnalysis folder. All containers holding standards or reagents must be labeled with their unique ID and expiration date.

Note: The volumes prepared for the reagents and standards may be altered by the analyst as necessary. The final concentrations of the constituents of a standard or reagent must not be changed.

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4.1. Chemicals

	Chemical	Alias	Manufacturer[1]	Catalog Number	Purity	Storage
4.1.1.	Color Standard Solution 500 Platinum Cobalt Units	Stock	Hach	141453	See CoA	Room Temp in the dark
4.1.2.	Color,WP CRM (Water Pollution Certified Reference Material)	ALTQC, ERA	Environmental Resource Associates	1070C	See CoA	Room Temp
4.1.3.	Deionized (DI) Water	H_2O	In House	N/A	N/A	N/A
4.1.4.	Cuvette Cleaning Solution		SEAL Analytical	5676		Room Temp
4.1.5.	Bleach, Generic	NaOCl, For Cleaning	Store Brand	N/A	No additives, ~6%	Room Temp
4.1.6.	Sodium Hydroxide	NaOH	Fisher Chemical (SR)	S318-3	ACS, ≥97%	Room Temp
4.1.7.	Ethylenediamine Tetraacetic Acid Disodium Salt, EDTA	$C_{10}H_{14}N_2Na_2O_8$ *H_2O	Fisher Chemical	S311-500	Certified ACS	Room Temp
Or Equivalent						

Deionized water is replaced daily. All other purchased chemicals are given the expiration date from the CoA and/or chemical container. If there is not an expiration date listed, the expiration date is designated by following the protocol in SOPs NU-041 and NU-042.

4.2. Calibration Standard Preparation

4.2.1. Calibration (Top) standard (150 PCU) –

Color Stock Standard PCU (4.1.1.)	75 mL
DI Water (4.1.3.)	to 250 mL

To a 250 mL volumetric flask containing approx. 100 mL of DI water, add 75 mL of 500 PCU stock standard and bring to volume.

Store in the dark when not in use. Holding time, 30 days.

4.2.2. The remaining standards are prepared from the 150 PCU calibration standard (4.2.1.). Using either a class-A glass volumetric pipette or a mechanical pipette, place the appropriate volume of top standard in a volumetric flask partially filled with DI water, and dilute to the mark with DI water. Use the table below as a guide.

Store in the dark when not in use. Holding time, 30 days.

Concentration (PCU)	QC Standard, Alias	Final Volume (mL)	Vol of Top Standard (4.2.1.) added (mL)
120		25	20
90		50	30
60	CCV	200	80
45	LFB	200	60
30		50	10
15		50	5
10.5		50	3.5
6.0	PQL	100	4

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4.3. Bleach Cleaning Solution –

Bleach (4.1.5.)	100 mL
DI water (4.1.1.)	to 500 mL

To a 500 mL volumetric flask, add 100 mL of bleach and bring to volume with DI water.

Store at room temperature. Used for cleaning only.

4.4. Alkaline EDTA Rinse (Cuvette Wash)

Sodium EDTA (4.1.7.)	10 g
NaOH (4.1.6.)	20 g
DI water (4.1.1.)	to 1000 mL

In a 1 L volumetric flask containing approx. 600 mL of DI water, dissolve 10 g of sodium EDTA and 20 g of NaOH. Dilute to volume with DI water and invert to mix.

Store at room temperature. Holding time, 60 days.

5. SAMPLE PRESERVATION, PREPARATION AND HANDLING

5.1. Since biological activity may change the color characteristics of a sample, the color determination should be made as soon as possible. Samples are stored refrigerated ≤ 6°C (not frozen) and must be analyzed within 48 hours of sampling. Allow samples to come to room temperature or use a warm water bath.

5.2. Invert the sample bottle well. Single use sterile syringes may be put directly into the sample bottle to obtain a representative aliquot. If the syringes are not sterile, the sample must be poured into a disposable beaker. Filter samples using the 0.45 µm filter and syringe apparatus into **labeled** disposable beakers.

Note: If filtration with a 0.45 µm filter does not remove all particulate matter from a sample, or if the Turbidity result for the sample is ≥15 NTU (see DEP SOP NU-103), the result is reported J qualified with the comment “The result may be biased high because of turbidity”.

Note: In some rare cases precipitation may occur over time in the already filtered aliquot of the sample. Samples should be analyzed as soon as possible after filtration. Always document visible changes of the sample appearance on the tray report/bench sheet.

6. SAMPLE ANALYSIS AND QUANTITATION (SEE DEP SOP NU-033)

Note: Observe routine maintenance as appropriate to keep the instrument in good working condition and enter any maintenance performed in the maintenance logbook.

6.1. Initial Preparation: Remove standards from their designated storage area and check the volumes and expiration dates. Discard any expired standards and prepare, if necessary. Make sure there is enough volume of all standards to complete the analysis.

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6.2. Calibration:

6.2.1. The AQ2 and AQ400 are manually calibrated at least every 30 days using the calibration standards in Section 4.2. It is recommended to calibrate the instrument upon the preparation of new standards.

6.2.2. The standards in the calibration are as follows:

STD	Conc. (PCU)	Source
S1	0.0	DI water
S2	6.0 (PQL)	See 4.2.2.
S3	10.5	See 4.2.2.
S4	15.0	See 4.2.2.
S5	30.0	See 4.2.2.
S6	45.0 (LFB)	See 4.2.2.
S7	60.0 (CCV)	See 4.2.2.
S8	90.0	See 4.2.2.
S9	120.0	See 4.2.2.
S10	150.0	Top Standard (4.2.1.)

6.2.3. The instrument can be auto calibrated using the high calibration standard placed into a reagent wedge in position 5 and select “Auto Calibrate” in the Options dialog box. This option is used only rarely. Please contact a supervisor for guidance. All calibration acceptance criteria must be met.

Note: The analyst can request in the software for a carryover calculation to be performed. The software will then automatically add an “S0” point to the end of the calibration sequence. Carryover calculations cannot be performed on manually poured calibrations.

6.3. Controls and Calibration Standard Loading:

6.3.1. Following the calibration or at the start of a new run, the following QC is required: Continuing calibration verification standard (CCV), continuing calibration blank (CCB), initial calibration verification (second source standard, ALTQC) and a PQL.

6.3.2. CCV and CCB should be poured individually on the sample tray and backet each batch of samples as shown in 6.4.5.

6.3.3. The reagent tray, with plastic wedge-shaped containers, is located on the right side of the injector arm. For the AQ2, cuvette cleaning solution is in position 1 and DI water is in position 15. For the AQ400, the cuvette cleaning solution is in position 1 and DI water is in position 20. The sample tray is loaded starting at cup 1. If calibration is not necessary, the run starts with a CCV and CCB.

6.4. Analysis Setup/Sequence Setup:

6.4.1. Turn on the AQ2/AQ400 Analyzer (switching the reagent cooling switch (small switch) first, and then the main power switch (large switch)) and the instrument computer.

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6.4.2. Double click on the AQ software icon. Select the appropriate instrument as the login name from the drop-down list and enter the password to display the Main Operation Screen. The password is "password".

6.4.3. Perform the daily maintenance as listed in the instrument logbook if it is the first run of the day. If another chemistry was run on the instrument prior to running W-Color, perform extra cleaning. Otherwise, continue to 6.4.4.

Note: Daily maintenance for Color may be performed using either the SEAL Cuvette Cleaning Solution (4.1.4.) or the Alkaline EDTA Rinse (4.4.). If TP is to be run on the instrument, it may be better to use the EDTA rinse, as it is safe for phosphate analysis.

Note: If the instrument was off, allow the instrument to warm-up for at least 15 minutes before performing the Daily Start-up procedure.

Note: A bleach cleaning solution (4.3.) can substitute for DI water in the cleaning procedure in the instrument logbook. Cleaning with DI water must follow a bleach cleaning.

6.4.4. Check that the DI reservoir is full and empty the waste container (if needed). Check that the waste pH is within the range of 2-12.5 before emptying in the sink with running water.

Note: pH that is below 2 or above 12.5 is considered hazardous waste and must be disposed of properly in hazardous waste containers.

6.4.5. Analytical Setup - Select *Scheduling* from the Main Operations Screen. Select a *Tray Number* (first FREE #) and the color reagent set. Enter the tray name which will be the run file name as follows: CTYYMMDDX# . Where, "YY" is the year, "MM" is the month", "DD" is the day and "X" indicates the instrument being used. The "#" represents a number (1, 2, etc.) to indicate the 1st, 2nd, etc. run of the day. Press *OK*. The sequence screen should appear. Starting at line 1, type the sample sequence in the 'Sample ID' column.

An example run sequence includes:

- **Calibrant(s)** (6.2.2.) – if calibration is needed
- **CCV** (60 PCU, 4.2.2.)
- **CCB** (4.1.3.)
- **ALTQC** (4.1.2.)
- **PQL** (6.0 PCU, 4.2.2.)
- **MBLK** (filtered, 4.1.3.)
- **LFB** (filtered, 45 PCU, 4.2.2.)
- Batched samples, including one sample duplicate per batch of up to ten samples
- **CCV**
- **CCB**

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Next, highlight the typed lines and select the **"TRUE_CL"** test. Set the "Type" column to "U#" where U means "unknown" and # indicates the cup number.

Finally, click on the *Save* icon to save the schedule and the *Print* icon to print the tray report/bench sheet.

6.4.6. Pour filtered samples and batch QC into the 1.2 mL instrument sample cups according to the schedule. All samples and QC that appear on the run schedule are poured in individual sample cups.

Note: The MBLK and LFB should be prepared as described in 5.2. and kept with their associated batch samples.

6.4.7. Start the run by double-clicking on the *Run* icon. A dialog box should appear. Select the tray (run file) to be analyzed and click *OK*. On the *Options* dialog box, check the auto-standardize box **only** if you intend to re-calibrate the instrument using the auto-calibration function (6.2.3.). Click *OK*.

6.4.8 Samples with concentrations above 150 PCU are determined by quantitative dilution. All samples should be analyzed undiluted. Alternatively, two different independently prepared dilutions are analyzed. Dilutions should be made with a mechanical pipette. Fill out the dilution recipes used on the pipette form, located at \\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RunData\MetaData\Color\Forms\.

6.4.9. After the analysis is complete, determine the pH of the undiluted aliquot of the sample and any diluted sample aliquots and record on the NU-COLORpH sheet located at \\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RunData\MetaData\Color\Forms\.

6.4.9.1. Check the pH of the filtered samples using pH strips with range 5.0 – 9.0 (3.3.). If the pH of a sample is observed as 5 on this strip range, verify the sample pH using an indicator strip with range 0.0 – 6.0 (3.3.).

6.4.9.2. If the pH of a sample aliquot is < 4 or > 9 it must be verified with a pH meter (See DEP SOP NU-026). If the pH is verified outside the 4-10 pH range add a test comment in the QC Manager indicating the pH at which the sample result was obtained.

6.4.9.3. If the pH of a sample is < 2, perform a $BaCl_2$ test to check for improper preservation. To perform the Barium Chloride test, place an aliquot of sample into a beaker and drop a small amount of $BaCl_2$ into the sample. If the sample turns cloudy then the sample was acidified with sulfuric acid (see your supervisor if the sample has been preserved with acid).

6.5. Quantitation/Calculations:

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6.5.1. The instrument will give the analysis results for Color in PCU.

6.5.2. The QC Manager calculates relative percent differences (RPDs) for duplicate samples as well as recoveries for QC standards. For general calculations including RPDs, see Section 5.9 of the FDEP Quality Manual.

6.5.3. The high calibration standard is prepared according to Section 4.2.1. The other standards used in the linear calibration are prepared from the high standard. The correlation coefficient for the calibration is calculated by the instrument software.

6.6. Matrix Considerations:

6.6.1. Note any equipment/field/trip blanks or field duplicates on the backlog. The backlog fields Matrix, Field ID, and Sampling Location are useful for identifying these sample types.

6.6.1.1. All equipment/field/trip blanks should have a color reading of < 2.0 and > - 2.0 PCU. If the measured color is not in this range, rerun the blank for confirmation on the same run (if possible) and add a comment for the sample in QC Manager.

6.6.2. Saltwater or other matrix interferences may cause results <-2.0 PCU. Dilute sample until the result is within the -2.0> and <2.0 PCU range.

6.6.3. The RPD for the **field** duplicates should be less than 20 % if the color values are above the practical quantitation limit (PQL). If the RPD is 20 % or greater, both field duplicates should be rerun for confirmation on the same run, if possible, and commented in QC Manager. Also note any visible differences between the field duplicates on the tray report and in the comments section of QC Manager.

6.7. Post-Run Procedure:

6.7.1. Click on the *Data Review* menu located on the left side of the screen.

6.7.2. Make any necessary changes to the sample list (dilutions, additional samples, etc.). Highlight and *cancel* any extra cup spaces that were not used for the run, then click on the *Accept All* icon. This adds 'ACC' to the results. A pop-up window appears where the file name needs to be entered. Follow the naming convention described in 6.4.6. Accepting the run in the software creates the excel transfer file used for uploading the data to the Laboratory Information Management System (LIMS). This file should be saved here:

\\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RawData\DiscreteAnalyzers\

Under the appropriate instruments "COLOR" folder and in the current year's subfolder.

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6.7.3. Under the *Maintenance and Set Up* tab at the top of the screen, select the *test* icon , then highlight the color test and print the Color Test Parameters Report.

6.7.4. Click on the *Reporting* icon on the left side of the screen. Check the reporting format 'Color Report' and print the run report. Select the chronological order report function. A browser window will open containing the report with the calibration and all the data.

6.7.5. Convert the excel file (6.7.2.) into QC Manager raw data.

7. DATA ARCHIVAL

7.1. The raw data is archived in paper format. A report folder is created for each analytical run. Occasionally more than one run may be in one folder. The folder is labeled with the test (w-color), filename/s (CTYYMMDDX#) and initials of the analyst.

7.2. Each report folder should contain the following items –

- AQ2/AQ400 run report
- AQ2/AQ400 test parameters
- AQ2/AQ400 tray report/bench sheet
- NU-RGTSTD_W-Color sheet
- NU-COLORpH sheet
- Backlog

and a copy of the data and QC assessments uploaded from QC Manager to LIMS at the time of test authorization. If dilutions were performed, a copy of NU-PIPETTE should be included in the folder.

7.3. The report folders are on file with the nutrients and general chemistry supervisory personnel. Periodically, the files are moved to the State Records Center for long time storage according to DEP SOP LB-006.

7.4. Color instrument data excel transfer files for upload to QC manager are stored in electronic format at:

\\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RawData\DiscreteAnalyzers\

Under the appropriate instruments "COLOR" folder and in the current year's subfolder.

7.5. Current worksheet templates are located in subfolder: \\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RunData\MetaData\Color\Forms\.

8. QUALITY CONTROL

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8.1. For method performance and acceptance criteria for QC measures and corrective action for out-of-control data and contingencies for handling out-of-control or unacceptable data, refer to the following DEP SOPs, NU-038 and NU-043.

8.2. A CCV and a CCB are run as check standards at the beginning of the run, every 10 samples, and at the end of each run. After the initial CCV and CCB a PQL level standard and an ALTQC (4.1.2.) standard are analyzed. Acceptance limits for recoveries of the standards are 70 - 130 % for the PQL and 90 - 110 % for the CCV. The ALTQC acceptance limits are 90 – 110 %. Acceptance limits for the CCB are < 2.0 PCU and > -2.0 PCU.

8.3. Analyze an MBLK (filtered), an LFB (filtered, spike code C1), and a duplicate sample for every batch of ten (or less) samples. Acceptance limits for the MBLK (filtered) are < 2.0 PCU and > -2.0 PCU. Acceptance limits for recoveries of the LFB are 90 - 110 %. The RPD for the batch duplicates should be less than 20 % provided the duplicate results are above the PQL (6.0 PCU).

8.4. Calibration Range - See QA Manual Appendix B Section 7.1.1, 'Protocol for Determining the Test Method Range of Applicability'. Acceptance limits for the % error of the calibration standards above the PQL are ±10 % and ±30 % at the PQL level. The correlation coefficient of the calibration must be equal to or greater than 0.995 for the calibration to be acceptable.

8.5. The pH of each aliquot analyzed for color is recorded on the NU-COLORpH sheet located at \\floridadep\data\labs\Lab3\Chem_Data\Inorganic\Nutrients\RunData\MetaData\Color\Forms\. If the pH of a sample is verified to be below 4 or above 10 units a test comment is added in QC Manager. Refer to 6.4.8.

9. SAFETY/HAZARDOUS WASTE MANAGEMENT

9.1. Review the Laboratory Health and Safety Plan (accessible at https://floridadep.sharepoint.com/dear/SitePages/Lab%20Health%20and%20Safety.aspx).

9.2. Safety glasses, gloves, and lab coats must always be worn handling samples, chemicals and standards.

9.3. Before performing the color analysis for the first time, read the MSDS sheet on the platinum-cobalt standard used in this analysis. Excess standards during preparation and analysis are collected in a beaker and disposed in the designated hazardous waste container.

9.4. All purchased color standards and wastes thereof may be corrosive. Wastes exhibit the characteristic of corrosivity if they are aqueous and have a pH ≤ 2 or ≥ 12.5. Test the pH of any instrument waste before discarding it down the sink. If it is determined to be corrosive it must be discarded as hazardous waste (acidic waste drum in the waste room if the pH is below 2). The standard described in section 4.1.1. must be discarded as hazardous waste (pH < 2). Any waste suitable for sink disposal should be flushed with excess tap water.

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10. REFERENCES/DEFINITIONS/VALIDATION DATA

For definitions see section 3 of the FDEP Quality Manual. All abbreviations used throughout the document are defined at least once in the document usually at the first time of use. Method validation data is on file with the supervisor and on the data drive at \\floridadep\data\labs\Lab5\Chemistry\Inorganic\Nutrient\Validations\Color\.

10.1. Standard Methods Online 2120 C-2011.

10.2. DEP SOP NU-026, Manual Determination of pH.

10.3. DEP SOP NU-033, Summary of Sample Analysis Procedures for the General Chemistry and Nutrients Workgroups.

10.4. DEP SOP NU-038, Nutrients and General Chemistry: Quality Assurance and Quality Control.

10.5. DEP SOP NU-042, Standard Preparation Procedure

10.6. DEP SOP NU-043, Nutrients and General Chemistry Quality Assurance and Data Authorization Guidelines.

10.7. Quality Manual, Bureau of Laboratories, FDEP.

10.8. Laboratory Health and Safety Plan.

10.9. DEP SOP LB-006, Standard operating procedure for records maintenance and storage.

Appendix of Significant Changes

11/17/2011	Section 6.4.5: Added Bleach Cleaning Procedure. Section 6.4.10. and 8.4.: Procedure for recording and adjusting the pH.
01/13/2012	MDL was adjusted to 2.5 PCU. Clarified waste disposal procedure. A filtered MBLK is run at the beginning of every batch (6.4.7.).
02/01/2012	Standard preparation was corrected in Section 4. Naming convention corrected in Section 6.4.6.
05/24/2012	Calibration preparation procedure changed, see Section 4. Calibration and analysis procedure edited in Section 6.
01/08/2015	File naming convention was modified for consistency and to specify the instrumentation used (see 6.4.6. and 7.1.). Data archival information was clarified in section 7.
01/07/2015	Filtering procedure in Section 6.4.7. and pH determination procedure in Section 6.4.10. modified. Appendix C example NU-COLORpH sheet updated.
04/21/2017	Section 6.4 was revised for clarity and passwords were added. Appendix B and C were removed.

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06/25/2018 A prepared standard is used as LFB. One LFB is filtered and analyzed for every batch of samples. Updated the reference method to the 2011 version.

01/19/2022 Added tables for instrumentation, supplies, chemicals, and standards. Updated sample filtering procedure. Removed mention of centrifugation as an option for removing particulate matter. Updated QC requirements. Removed SM 2120 B-2011 as reference method.

02/24/2022 Removed pH adjustment procedure for samples outside the pH 4-10 range. Clarified auto calibration procedure and acceptance criteria for the standards in the calibration.

08/23/2022 Added AQ400 information. Added EDTA solution to Section 4. Clarified maintenance procedure and added reference to instrument logbooks. Added example run sequence. Renamed Appendix A to Appendix of Significant Changes.

09/22/2023 Modified ALTQC acceptance limits. Data drive file path links updated. Removed AQ2-2 instrument.

10/03/2024 Removed procedure of filtering high turbidity samples using 0.20 µm filters. Added instructions for reporting color samples with high turbidity. Updated data drive file path links.

10/01/2025 MDL updated from 2.5 PCU to 2.0 PCU. Removed the need to test for high turbidity if it is not requested by the client. In section 4, the expiration date related procedures. Added clarification in section 6 that two different dilutions of a sample can be analyzed instead of the undiluted and diluted aliquot.

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