WEIGHING CHRONIC TESTS

1. SCOPE AND APPLICATION

1.1. This procedure is used to determine the dry weight of organisms used in chronic bioassays described in test methods found in EPA 821-R-02-013 and EPA 821-R-02-014.

2. SUMMARY OF METHOD

2.1. Labeled weigh boats are prepared in advance of test termination. They are dried in a drying oven and stored in a desiccator. Tare weights of the weigh boats are determined and recorded. Organisms to be weighed are placed in labeled weigh boats. Weigh boats containing organisms are dried in a drying oven and then stored in a desiccator. Final weights are measured and recorded in the appropriate logbook.

2.2. Prior to weight measurements, the user must weigh and record on the appropriate bench sheet the weights of two S-weights (or other certified weights) that bracket the expected weight of the object(s) to be weighed. After the final measurement, the user must bracket the measurements by weighing and recording on the appropriate bench sheet the values of two S-weights or other suitable weighs, such as Bio-R-weights.

2.3. Interferences include airflow and vibrations which interfere with the balance, weigh boats contaminated by moisture or body-oils, improperly calibrated balance and contaminated S or R-weights.

3. EQUIPMENT AND SUPPLIES

3.1. Weigh Boats

3.1.1. Aluminum foil

3.1.2. Fine point, black permanent marker

3.1.3. 4 inch Carolina dish

3.1.4. Scissors

3.1.5. Volumetric flask stopper (#13)

3.1.6. Forceps

3.1.7. Ruler, 30 cm

3.1.8. Weigh-boat form block (located in Cabinet UB in Room B246)

3.1.8.1. The weigh-boat form block is a block of wood approximately 1.85 cm × 7 cm × 2 cm with 12 circular holes drilled to an approximate depth of 6 mm.

3.1.9. Desiccator with desiccant

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3.1.9.1. Desiccant must be blue or white in color. A pink color indicates that the desiccant is hydrated. Dry desiccant in oven before use until blue or white color returns.

3.1.9.2. Keep the desiccator depressurized even when not in use.

3.2. Weighing

3.2.1. Weigh boats

3.2.2. Filter forceps

3.2.3. Autoclave gloves

3.2.4. Organisms to be weighed

3.2.5. Mettler XA105 balance (SN B314195659) (located in the micro lab), calibrated and used per SOP PCR-2.2.

3.2.6. Drying oven maintained at either 100 ˚C (dry time >6 but <8 hours during business hours only) or 60 ˚C (dry time ~ 24 hours when drying overnight)

3.2.7. Vacuum source with attached tubing (if using desiccator with vacuum attachment)Certified S-weights (in B246 Cabinet A2) or R-weights (beside XA105 balance)

3.2.8. Appropriate *Chronic Bioassay Bench Sheet* for the test being conducted

4. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

4.1. See SOP FS 1000 for sample collection and preservation.

5. PREPARATION OF WEIGH BOATS

5.1. Wearing gloves, lay a (~30 cm X 30 cm) sheet of aluminum foil on a flat surface. Using a ruler, mark 2 cm intervals along opposite ends of the foil. Use the ruler to connect the marks. Do the same on the other two sides of the sheet. The foil should now be marked off into 2 cm squares.

5.2. Carefully cut along one set of 2 cm lines to form 2 cm × 30 cm long aluminum foil strips, with lines every 2 cm. This should form 16 squares per strip.

5.3. Stack the strips and clip them on one end using a small binder clip. Hang them on the nail between Cabinet UA and Cabinet UB in Room B246 until needed.

5.4. As needed, lay the appropriate number of strips shiny side down on a flat surface. (Number of strips is dependent upon the test being taken down.) Use the permanent marker to clearly number both sides of the strips (same number and same square). The numbers should be written as close to the center of each square as practical. Begin with number 1 and continue to 24 (more or less, depending on the test).

5.5. After numbering, cut the squares from the strips.

5.6. Using forceps, find the first square in series and center over one of the holes in the weigh-boat form block. Carefully form the aluminum foil boat by placing the bottom

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of the #13 flask stopper on top of the foil square and gently pressing into the hole of the form block. Remove the stopper and carefully remove the boat from the hole using the forceps.

5.6.1. After the boat is formed, make sure the number is still legible. If not, discard the boat and make a new one.

5.7. Place formed boats in a small Carolina dish in a logical order, such as a spiral, so they can be found in numeric order when needed.

5.8. Repeat step 5.6 - 5.7 until all weigh boats have been made. Cover the dish with foil to prevent dust and dirt particles from falling into the weigh boats.

5.9. Pressurize the desiccator if necessary and remove the top. Place the Carolina dish in the center of the desiccator, place the top back on, and evacuate the desiccator using a vacuum source. Keep boats in desiccator until needed.

5.9.1. When pressurizing the desiccator, close the desiccator valve 30 seconds after you can no longer hear air entering.

5.9.2. Monitor the desiccant as described in Section 3.1.9.

6. WEIGHING CHRONIC TESTS

6.1. Day 6/144 hours of the 7 day chronic test

6.1.1. Remove the Carolina dish containing the weigh boats from the desiccator and place it in the drying oven. The foil should be folded or removed to allow air exchange. Set the oven temperature to 100 ˚C and allow the weigh boats to dry for at least one hour.

6.1.2. After at least one hour at 100 ˚C, remove the Carolina dish containing the weigh boats from the drying oven and return it to the desiccator.

6.2. Day 7/168 hours of the 7 day chronic test

6.2.1. Remove the Carolina dish containing the weigh boats from the desiccator and place it next to the Mettler XA105 balance in the micro lab. Fold back the foil and allow the Carolina dish to set for 15-20 minutes to acclimate the weigh boats to the room conditions.

6.2.2. Weigh the weigh boats. Use the forceps to avoid touching the weigh boats with bare flesh. Weigh the weigh boats one at a time in ascending alphanumeric order. After the fifth weigh boat and each subsequent 5 weigh boats, reweigh the first weigh boat. If at any time during the weighing process the weight of the first weigh boat varies by more than ± 0.05 mg from the initial weight, return the weigh boats to the desiccator for approximately 30 minutes. Continue until all weigh boats have been weighed without an exceedance of the 0.05 mg limit.

6.2.3. Record the weights on the appropriate *Chronic Bioassay Bench Sheet*.

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6.2.4. Take the weigh boats to the test termination area in B246. Load the boats per the *Test Termination* procedure in the appropriate chronic bioassay SOP (TA-07.09, TA-07.14,TA-07.15).

6.2.5. Place the Carolina dish containing the loaded weigh boats in the desiccator. Leave the weigh boats in the desiccator overnight.

6.3. Day 8 of the 7 day Chronic test

6.3.1. Remove the Carolina dish containing the weigh boats from the desiccator and place it in the drying oven. Weigh boats **with organisms** should be dried at 100 ˚C for a minimum of 6 hours. If dried at 60 ˚C, the drying time should be a minimum of 24 hours.

6.3.1.1. In the event that an oven fails, temperatures between 60 ˚C and 100 ˚C can be used for drying, as long as the laboratory can demonstrate that the organisms are dried to a constant weight.

6.4. Day 9 of the 7 day Chronic test

6.4.1. Remove the Carolina dish containing the weigh boats from the drying oven. Weigh the boats as in Step 6.2.2. and record the weights on the appropriate *Chronic Bioassay Bench Sheet*.

7. DATA ARCHIVAL

7.1. Use a black, waterproof pen for all bench sheets and logbooks.

7.2. Record weights on the appropriate *Chronic Bioassay Bench Sheet*.

7.3. To correct raw data entries, place a single line through the incorrect or illegible entry. Write the corrected entry near the error and initial the correction. Do not overwrite entries.

7.3.1. The corrected entry is written as near its appropriate space as possible or else in the comment box on the appropriate bench sheet.

7.3.2. If the correction requires an explanation, write a comment and reference the error. Initial and date the explanation.

7.3.3. Use sequential alphabetical annotation to identify errors and corresponding comments (use A for first error and use A for corresponding explanatory comment; use B for second error, etc.).

8. QUALITY CONTROL

8.1. The balance is checked before use to ensure it is reading accurately. Two certified S- or R-weights are weighed such that they bracket the expected weights of the objects being weighed.

8.2. The weights are not handled with bare hands to prevent contamination from body oils. They are also kept in a closed box to prevent dirt and dust from contaminating them.

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8.3. Weights are taken using a balance set on a marble slab to minimize vibrations which could interfere with the balance.

8.4. The balance is covered to minimize air flow.

8.5. Weigh boats are desiccated to eliminate moisture as an interference in the weighing process.

8.6. Forceps are used to handle the weigh boats to prevent body oils from contaminating them and affecting the weight.

8.7. If a problem occurs during the sample processing or testing, the following steps are performed:

8.7.1. Lab supervisor is notified.

8.7.2. Client is notified (client makes decision for additional testing).

8.7.3. Section administrator is notified.

8.7.4. A non-conformance report (NCR) is submitted describing the problem and the corrective action.

8.7.4.1. To create an NCR: open LIMS and go to Laboratory/Administrative/Non-Conformance Report

8.8. Refer to http://www.dep.state.fl.us/labs/library/lab_sops.htm for sample collection and preservation, and the Bureau Quality Manual for corrective action and handling out-of-control data.

9. SAFETY/HAZARDOUS WASTE MANAGEMENT

9.1. Proper laboratory protection, including lab hood or ventilation system, lab coat, gloves and safety glasses, is required when working with chemicals and unprocessed samples.

9.2. Refer to Bureau Quality Assurance Manual for sample disposal (http://www.floridadep.org/labs/library/progplan.htm).

9.3. Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management https://www.fldepnet.org/content/dear/bureau-laboratories-safety-documents

10. REFERENCES

10.1. EPA 821-R-02-013 Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, 4th Ed. October 2002.

10.2. EPA 821-R-02-014 Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms, 3rd Ed. October 2002

10.3. DEP SOP PCR-2.2, Calibration, Use and Maintenance of Mettler Toledo XA105DU Balance

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10.4. DEP SOP TA-07.09, Pimephales promelas and Cyprinella leedsi Chronic Toxicity Testing Methods

10.5. DEP SOP TA-07.14, Menidia beryllina Chronic Toxicity Testing Methods

10.6. DEP SOP TA-07.15, Americamysis bahia Chronic Toxicity Testing Methods

10.7. Bureau Quality Assurance Manual

10.8. Bureau Health and Safety Manual

10.9. MSDS Information Sheets

APPENDIX OF CHANGES

08/16/2013	Combined SOPs TA-06.20-1.15 and TA-06.21-2.0. Reformatted the combined SOP to produce TA-06.21-3.0.
02/09/2015	Formatted for 508 compliance.
08/14/2015	Updated links. Weight location. Drying time clarification.

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