ANALYSIS OF SELECTED METALS BY DUAL VIEW INDUCTIVELY COUPLED PLASMA-ATOMIC EMISSION SPECTROSCOPY (ICP-AES)

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1. SCOPE AND APPLICATION

1.1. This standard operating procedure (SOP) details the analysis of metals in waters, soils, wastes, and other environmental samples utilizing an Inductively Coupled Plasma-Atomic Emission Spectrometer (ICP-AES). It covers analyses by EPA Method 200.7 (test ID's W-ICP, W-ICP-F, W-SI-ICP, and W-SI-ICP-F), EPA Method 6010 (test ID's W-ICP-R, W-ICP-R-F, S-ICP, SA-ICP, WASTE-ICP, T-

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ICP, SPLP-ICP, TCLP-ICP, and S-ICP-TO), and Standard Method 2340B (test IDs W-HARD and W-HARD-F, see DEP SOP **MT-058**).

1.2. The Perkin Elmer ICP-AES (models Optima 5300 & 7300 Dual View) is an atomic emission spectrometer possessing the capability of both axially and radially oriented viewing positions. This instrument uses inductively coupled plasma as the excitation source and a **S**egmented array, **C**harge-coupled **D**evice ("SCD") as the detector. It is capable of analyzing up to 76 elements simultaneously, offers wide linear dynamic range, and resolution of approximately 6 pm at 200 nm. The procedure performed in this laboratory analyzes 29 elements simultaneously.

2. SUMMARY OF METHOD

2.1. Digested water, sediment, waste, and tissue samples are analyzed and reported as 'total recoverable metals'. With the exception of silver, where EPA method 200.7 is approved for the determination of metals and metalloids in drinking water, samples may be analyzed directly by pneumatic nebulization without acid digestion if the sample has been properly preserved with acid and has a turbidity of < 1 NTU at the time of analysis. EPA Method 200.7 is approved for drinking water for Ba, Be, Cd, Ca, Cr, Cu, Mg, Ni, SiO_2, Na, Al, Fe, Mn, and Zn. Sediment samples digested using hydrofluoric acid and microwave radiation are reported as 'total metals'. Filtered water samples are analyzed without digestion and reported as 'dissolved metals'. TCLP (Toxicity Characteristic Leaching Procedure) and SPLP (Synthetic Precipitation Leaching Procedure) extracts are also analyzed. Soils and sediments are reported on a dry weight basis; wastes and tissues are reported on a wet weight basis.

2.2. The following is a list of the 29 elements currently analyzed along with the approximate spectral line in nanometers (nm) used:

Element	Spectral Line	Element	Spectral Line	Element	Spectral Line
Al	396.164	Cu	327.391	Na	589.611
Sb	206.834	Fe	273.935	Sr	421.549
As	188.980	Pb	220.350	Tl	190.791
Ba	233.528	Mg	279.080	Sn	189.926
Be	313.099	Mn	257.606	Ti	334.938
B	208.890	Mo	202.028	V	292.399
Cd	228.800	Ni	231.602	Zn	206.195
Ca	317.930	K	766.500	SiO_2	251.609
Cr	267.709	Se	196.023	Li **	670.759
Co	228.613	Ag	328.067	Sc *	361.384
				Sc *	357.251

* Internal standard

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** Lithium analyzed by the Perkin Elmer ICPDV 7300 only.

2.3. The instrument is calibrated with 1 standard (see section 4.10.) and a blank. The calibration equation is linear. Since only two points are used, the calibration curve is forced through the blank. **Data are not reported outside of this established range of linearity unless they are properly diluted to within this established range of linearity.**

The calibration curve linearity is verified on every run by analyzing a second source standard, a continuing calibration verification (CCV) sample, and a limit of quantitation (LOQ) sample. The calibration is verified by analyzing the second source standard purchased from a vendor that is different from the ones used for the calibration standards (ICPMINC, ICPMETC, LIC and AG-SIO2C; sections 4.12.1 - 4.12.4.). The analytes present in these second source standards are at the same concentration as the appropriate calibration standard. If the determined concentration for any analyte in these standards is less than 95 % or greater than 105 % of the stated value then the data for that analyte cannot be used unless it is properly qualified and commented.

Initial calibration verification standards (CCVC i.e. ICPMINCCVC, ICPMETCCVC, LICCVC and AG-SIO2CCVC; sections 4.11.1 - 4.11.4.) are analyzed for each analyte and these standards must be determined to be no less than 95 % or no greater than 105 % of the stated value for each analyte in the standard and the Relative Standard Deviation (RSD) from the four replicate integrations must be less than 3 % for each analyte. If the determined concentration or RSD for any analyte is outside of this range then no data can be reported for that analyte unless properly qualified and commented. In addition, the instrument practical quantitation limit (PQL) solution (see 4.12.7. and 4.12.8.), which corresponds to the LOQ, is run after the instrument calibration on every analytical batch with established control limits of 70 – 130 %. If an analyte is outside of the control limits, samples requiring results for the affected analyte must be either rerun or qualified using the criteria for positive and negative bias in DEP SOP **LB-027.**

3. APPARATUS AND EQUIPMENT

3.1. Perkin Elmer Optima models 5300 or 7300 Dual View Inductively Coupled Plasma Atomic Emission Spectrometers (ICP-AES).

3.2. Perkin Elmer AS-93 Plus or S10 autosampler.

3.3. A three-channel peristaltic pump is used for sample introduction, internal standard introduction, and drain for waste. A second peristaltic pump (located on the auto-sampler) is used to supply the rinse solution.

3.4. A Polyscience Refrigerated Recirculator.

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3.5. 17 x 100 mm polypropylene round bottom culture test tubes - FS - Cat. No. 14-956-7E.

3.6. 0.045 in. ID, PVC manifold (pump) tubing - Red/Red - FS - Cat. No. 14-190-507.

3.7. 0.015 in. ID, PVC manifold (pump) tubing - Orange/Green - FS - Cat. No 14-190-501.

3.8. 0.030 in. ID, PVC manifold (pump) tubing - Black/Black - FS-Cat. No 14-190-504.

3.9. Argon gas supplied from the laboratory's MicroBulk argon delivery system.

3.10 HF Scientific Inc. MicroTPW Portable Turbidimeter, S/N: 201205062.

3.11 Class A glass volumetric pipettes, assorted volumes.

3.12 Eppendorf pipettes, 100 µL – 1000 µL.

3.13 Finnpipette, 2 – 10 mL.

4. CHEMICALS AND STANDARDS

Note: Be careful not to contaminate solutions with trace metals or minerals during preparation and use. Be sure to mix all solutions thoroughly immediately after they are prepared. Standard solutions must be stored in Teflon bottles. Use class A glass volumetric pipettes, Finnpipette 2-10 mL variable pipettes, or Eppendorf type pipettes in the preparation of standards, standard reference materials (SRMs), and spiking solutions. Check the calibration of Fisher and Eppendorf pipettes every month and record the information in the Pipette Calibration Form MT-PIPCAL-1.0. The template can be found in \\FLDEP1\DEAR\Labs\Lab5\Chemistry\InorganicPrep\METALS\pipette calibration.

Note: All calibration standards, calibration verification standards, calibration check solutions, and PQL solutions should contain 2 % HNO_3 and 1 % HCl unless otherwise noted. Silver solutions tend to be unstable at high concentrations and should contain 4 % HNO_3. The expiration date for standards prepared from purchased metal standards will be the same as the expiration date for the purchased standard. All purchased and prepared standards must be logged into the LIMS standard prep tracker. A label with a serial number and expiration date must be placed on the bottle of standard.

Note: See DEP SOP M**T-069** for instruction on reagent preparation documentation and expiration dates. All bottles of reagents in the laboratory are required to have unique identifiers. This is accomplished by logging purchased and prepared reagents into the LIMS standard prep tracker. A label, which includes a serial number, must be printed and placed on the bottle of reagent.

4.1. De-ionized (DI) water - From the laboratory DI water supply system.

4.2. Trace metal grade nitric acid (HNO_3) - Stored in acid cabinet under the hood. Expiration date provided by the manufacturer.

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4.3. Trace metal grade hydrochloric acid (HCl) - Stored in the acid cabinet under the hood. Expiration date provided by the manufacturer.

4.4. Rinse Water - 200 mL HNO_3 and 100 mL HCl in 10 L of DI water. Expires 1 year after the preparation date.

4.5. Acid Rinse Solution - Dilute 50 mL HNO_3 and 10 mL of HCl to 1 L with DI water - this solution contains 5 % HNO_3 and 1 % HCl. Expires 1 year after the preparation date.

4.6. Mn Profile Solutions – Used to automatically align the torch viewing position for the highest signal intensity.

Expires according to the manufacturer's expiration date of the stock manganese, however, this standard's expiration may be extended as long as the manganese concentration is stable and a stable signal is produced when aspirating the solution.

4.6.1 Radial View - 10 mg/L Mn solution: Pipette 1.0 mL of a stock supply of 10,000 mg/L Mn solution into a 1 L volumetric flask containing 900 mL of DI water, 20 mL of HNO_3, and 10 mL of HCl. Dilute to the mark with DI water.

4.6.2 Axial View - 1 mg/L Mn solution: Pipette 0.10 mL of 10,000 mg/L Mn solution into a 1 L volumetric flask containing 900 mL of DI water, 20 mL of HNO_3, and 10 mL of HCl. Dilute to the mark with DI water.

4.7. Internal Standard - 10 mg/L Sc: Pipette 2 mL of scandium standard (10,000 mg/L) into a 2 L flask containing 1 L DI water and 40 mL of HNO_3 and 20 mL of HCl. Dilute to the mark with DI water. Expires according to the manufacturer's expiration date of the stock scandium standard, however, this standard's expiration may be extended as long as the scandium concentration is stable and a stable signal is produced when aspirating the solution.

4.8. Primary Single-Element Standards – 10,000 mg/L or 1,000 mg/L NIST-traceable single-element standards are purchased from vendors such as SPEX, High Purity, Exaxol, etc. Used for calibration standard preparation (see 4.10.). Expiration dates assigned by the manufacturer.

4.9. Secondary Single-Element Standards – 10,000 mg/L or 1,000 mg/L NIST traceable single-element standards are purchased from vendors such as SPEX, High Purity, Exaxol, etc. Used for second source calibration verification standard preparation (see 4.12.). The commercial source of the secondary single-element standards must be different than the commercial source of the single-element standards used to prepare the calibration standards. Expiration dates assigned by the manufacturer.

4.10. Calibration Standards

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The expiration date of the prepared calibration standards are the same as the purchased primary single-element standards from which they were made.

4.10.1. Calibration Blank (INITIAL BLANK, CCB, I-BLK) - In a 1 L Teflon bottle, add 700 mL DI water, then 20 mL HNO_3 and 10 mL HCl and dilute to the mark with DI water. This solution contains 2 % HNO_3 and 1 % HCl.

4.10.2. ICPMIN - 200 mg/L Ca, Na; 100 mg/L Fe, Mg; 50 mg/L Al; 20 mg/L K.

4.10.3. ICPMET - 10 mg/L Cr, Cu, Pb, Mn, Sr, V and Zn; 5 mg/L Ba; 1 mg/L Sb, As, B, Cd, Co, Mo, Ni, Se, Tl, Sn, Ti and 0.5 mg/L Be.

4.10.4. AG-SIO2 - 0.25 mg/L Ag; 50 mg/L SiO_2.

4.10.5. LITHIUM - 1 mg/L Li.

4.11. Continuing Calibration Verification (CCV and CCVC) Standards

The expiration date of the prepared CCV standards are the same as the purchased single-element standards from which they were made.

4.11.1 ICPMINCCV/ICPMINCCVC - 100 mg/L Ca, Na; 50 mg/L Fe, Mg; 25 mg/L Al; 10 mg/L K. ICPMINCCV is the continuing calibration verification solution for these six elements and ICPMINCCVC is the initial calibration verification solution for these six elements.

4.11.2. ICPMETCCV/ICPMETCCVC - 5 mg/L for Cr, Cu, Pb, Mn, Sr, V, Zn; 2.5 mg/L for Ba; 250 µg/L for Be; 500 µg/L for Sb, As, B, Cd, Co, Mo, Ni, Se, Tl, Sn, Ti. ICPMETCCV is the continuing calibration verification solution for these 20 elements and ICPMETCCVC is the initial calibration verification solution for these 20 elements.

4.11.3. AG-SIO2CCV/AG-SIO2CCVC - 0.125 mg/L Ag; 25 mg/L SiO_2. AG-SIO2CCV is the continuing calibration verification solution for silver and silica and AG-SIO2CCVC is the initial calibration verification solution for silver and silica.

4.11.4. LICCV/LICCVC - 0.5mg/L Li. LICCV is the continuing calibration verification solution for lithium and LICCVC is the initial calibration verification solution for lithium.

4.12. Calibration Check Solutions

The expiration date of the prepared calibration check standards are the same as the purchased single-element standards from which they were made.

4.12.1. ICPMINC - Second source check standard 200 mg/L Ca, Na; 100 mg/L Fe, Mg; 50 mg/L Al; 20 mg/L K. This is the minerals calibration verification solution. Must be prepared from the secondary single-element standards (see 4.9).

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4.12.2. ICPMETC - Second source check standard 10 mg/L Cr, Cu, Pb, Mn, Sr, V and Zn; 5 mg/L Ba; 0.5 mg/L Be, 1 mg/L Sb, As, B, Cd, Co, Mo, Ni, Se, Tl, Sn, Ti. This is the trace metal calibration verification solution. Must be prepared from the secondary single-element standards (see 4.9).

4.12.3. AG-SIO2C - Second source check standard 0.25 mg/L Ag; 50 mg/L SiO_2. This is the Ag and SiO_2 calibration verification solution. Must be prepared from the secondary single-element standards (see 4.9).

4.12.4. LIC - Second source check standard 1 mg/L Li. This is the lithium calibration verification solution. Must be prepared from the secondary single-element standards (see 4.9).

4.12.5. ICS2007 - Interference check solution. Pipette 1 mL of Sb Stock Standard (1000 mg/L), 1 mL of ICS # 3 Standard, 1 mL of ICS # 18 Standard and 50 mL of ICS # 5 Standard into a 1 L volumetric flask containing 600 mL of DI water, 20 mL of HNO_3 and 10 mL HCl. Dilute to the mark with DI water. Refer to **APPENDIX A** for concentrations.

4.12.5.1 ICS # 3 – Custom NIST-traceable standard prepared by SPEX, Inorganic Ventures, or another suitable vendor. Contains B at 500 mg/L, Mo at 300 mg/L, SiO_2 at 492 mg/L, and Ti at 1000 mg/L.

4.12.5.2 ICS # 18 – Custom NIST-traceable standard prepared by SPEX, Inorganic Ventures, or another suitable vendor. Contains As, Pb, and Tl at 1000 mg/L; Ba, Cd, Cr, Co, Cu, Ni, Ag, V, and Zn at 300 mg/L; Be at 100 mg/L, Mn at 200 mg/L, K at 20,000 mg/L, and Se at 500 mg/L.

4.12.5.3 ICS # 5 – Custom NIST-traceable standard prepared by SPEX, Inorganic Ventures, or another suitable vendor. Contains Al at 1200 mg/L, Ca at 6000 mg/L, Fe at 5000 mg/L, Mg at 3000 mg/L, and Na at 1000 mg/L.

4.12.6. Instrument PQL Stock Solution - Prepared by a dilution of purchased single-element stock standards. This intermediate solution should contain all analytes listed in Section 2.2, except SiO_2 (refer to 4.12.8), at concentrations of 50X the PQL levels. Make sure the most recent PQL levels are used when making the solution. Excel files containing the current MDLs and PQLs may be found in \\FLDEP1\DEAR\Labs\Lab5\Chemistry\Common\MDLs_Control_Limits\Metals_Mercury.

4.12.7. Instrument PQL Solution - CPQLDV and IPQLDV (a suffix is added to these abbreviations (e.g. 9, 10, etc.) to indicate current PQL solution used) - Add 600 mL of DI water, 20 mL HNO_3, and 10 mL HCl to a 1 L

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volumetric flask. To this flask, pipette 20 mL of the Instrument PQL Stock Solution (see section 4.12.6.). Dilute to mark with DI water.

4.12.8. Instrument SiO_2 PQL Solution - CPQLSILICA and IPQLSILICA (a suffix is added to these abbreviations (e.g. 2, 3, etc.) to indicate current PQL solution used) - Add 600 mL of DI water, 20 mL HNO_3, and 10 mL HCl to a 1 L volumetric flask. Add the appropriate amount of individual stock SiO_2 standard and dilute to mark with DI water.

4.13. Spiking Solutions

The expiration date of the prepared spiking solutions are the same as the purchased single-element standards from which they were made.

4.13.1. Digestion PQL Solution - DPQLDV (a suffix is added to this abbreviation to indicate the current PQL solution used (e.g. 9, 10, etc.)). This solution is prepared from purchased single-element stock standards in 2 % HNO_3 and 1 % HCl at a concentration of 50X the final PQL. Make sure the most recent PQL levels are used when making the solution. Excel files containing the current MDLs and PQLs may be found in \\FLDEP1\DEAR\Labs\Lab5\Chemistry\Common\MDLs_Control_Limits\Metals_Mercury.

4.13.2. Trace Metal Spike Solution - HW6. This spike is prepared in two parts, HW6A and HW6B using the purchased second source single-element standards. HW6A is prepared with 2 % HNO_3 and 1 % HCl while HW6B is prepared with 2 % HNO_3. Refer to **APPENDIX B** for spike concentrations.

4.13.3. Mineral Spike Solution - ICPMINA. For both digested and instrument spikes (digested and filtered samples) use a purchased solution from SPEX. Refer to **APPENDIX B** for the spike concentrations. The expiration date is provided by the manufacturer.

4.13.4. Sediment Spike Solution - SEDSPK4. This spike is prepared in two parts, SED SPK4A and SED SPK4B using the second source single-element standards. SED SPK4A is prepared with 2 % HNO_3 and 1 % HCl while SED SPK4B is prepared with 2 % HNO_3. Refer to **APPENDIX B** for spike concentrations.

4.13.5. SiO_2 Spike Solution - The second source silica stock solution is used as the spiking solution. Add 200 uL of this solution to 10 mL of sample or DI water to produce a spike of 20 mg/L SiO_2.

Note: The current silica stock standard may have been made from sodium metasilicate in which case it cannot be combined with any standards used for sodium analysis.

5. SAMPLE PREPARATION PROCEDURE

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5.1. Filtered Water Samples - For undigested, filtered and acidified samples, which are checked out directly from the custody room, prepare sample matrix spikes as follows. For trace metals, pipette 200 uL of spiking solution HW6 (A and B) (refer to **APPENDIX B** for spike concentrations) into 10.0 mL of sample in a centrifuge tube and mix well. For minerals spikes, pipette 200 uL of solution ICPMINA (refer to **APPENDIX B** for spike concentrations) to 10.0 mL of sample. For samples requiring silica results, pipette 200 uL of the 1000 mg/L second source SiO_2 stock standard into 10.0 mL of sample in a centrifuge tube and mix well. **Each batch must contain at least one set of duplicate matrix spikes and at least one spike for every ten samples (two different samples must be spiked for batches over 10 samples).**

One method blank (see 4.10.1), one IPQL (see 4.12.7 – 4.12.8), and one LFB (see 4.13.2 – 4.13.3) are also required for each batch of filtered water samples.

Note: If a precipitate is formed in a filtered sample during acidification, transport, or storage, the sample must be digested. Notify your supervisor immediately if precipitate or particulates are observed in a filtered sample.

Note: If, after spiking a QC sample, the concentration of an analyte is above the instrument's calibration range, rerun the QC sample replicates and spikes for that analyte diluted appropriately. The QC sample replicates and the spikes should be diluted by the same factor.

5.2. Unfiltered (digested) water samples - See the DEP SOP **MT-024** for details on preparation.

5.3. Solid samples (soils, sediments, and wastes) - See the DEP SOPs **MT-060 and MT-063** for details on sample preparation.

5.4. TCLP and SPLP samples - See the DEP SOP **MT-068** for TCLP and SPLP extractions.

6. INSTRUMENT START-UP/SHUTDOWN/STANDBY MODE PROCEDURE: 5300 or 7300

6.1. START-UP - In the event that the instrument has been previously powered down completely, the following procedure must be used to begin operation.

6.1.1. Turn on the Polyscience chiller and allow it to equilibrate the temperature to ~ 17 °C.

6.1.2. Switch the Optima 5300 or 7300 MAIN POWER toggle switch, located on the right side panel of the instrument, to the ON position.

6.1.3. Open the software as in sections 7.1 and 7.2 below.

Note: If the instrument has been powered down for a minimal amount of time, the spectrometer will require approximately 74 minutes to become operational. However, if the instrument has been powered down for more than a few minutes it should be allowed to stabilize

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overnight while having everything turned on except the plasma and peristaltic pump. This will allow the optics bench and detector to reach a stable temperature. After that time, the plasma can be turned on and allowed to warm up.

6.2. SHUTDOWN - In the event that the instrument must be completely powered down, the following procedure should be followed.

6.2.1. Extinguish the plasma and turn off the pump (see 6.3).

6.2.2. Exit the Winlab32 software.

6.2.3. Flip the MAIN POWER toggle switch located on the right side panel of the instrument to the OFF position

6.2.4. Turn off the Polyscience chiller.

6.3. STANDBY MODE - SHUT DOWN. If desired, at the end of an analysis the instrument can be put in this mode if it is not to be used for several weeks. In Standby Mode, the argon purge gas to the detector and the detector coolant system is shut down. The command to enter this mode can be accessed two ways.

6.3.1. Manual Standby Mode Access: Click on the SYSTEM icon, Auto Shutdown/Start-up, then check "Shutdown" either with "Immediately on OK" (i.e. right now!) or "At end of Automated Analysis," finally "Put Spectrometer into Standby" and click "ok" to apply settings.

6.3.2. Automated Standby Mode Access: Click on the "Auto" icon, which brings up the Automated Analysis Control window. Under the SET UP tab, click on Auto Shutdown, Set... and the same screen as in 6.3.1. will appear. Check "Shutdown" either with "Immediately on OK" (i.e. right now!) or "At end of Automated Analysis", finally "Put Spectrometer into Standby" and click "ok" to apply settings.

6.4. STANDBY MODE - START UP. If the instrument is currently in Standby Mode, the detector system will need to cool back down to operating temperature before data can be collected. To exit Standby Mode and prepare for analyses, enter the AUTOMATIC SHUTDOWN/START-UP window either via the "System, Auto Shutdown/Start-Up" path or via the AUTO icon, SET UP tab, "Auto Shutdown, Set..." box. Click on "Start-Up" and choose a time. Once the instrument begins the start-up process, it will take approximately 74 minutes before the spectrometer is ready.

7. SAMPLE ANALYSIS PROCEDURE

Note: Read the Optima 5300 or 7300 Family Hardware and Software guides, and online help in WinLab32 for details.

7.1. Turn on the computer, log on as "metals" (see supervisor for password) and click on the WinLab32 icon to enter the software. If the software is already loaded there is no need to exit and re-enter.

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7.2. Upon entering WinLab32, a "System Status" window will open immediately, pictorially indicating the process of checking the status of the spectrometer, plasma, and autosampler subsystems. Green colored "pipes" indicate successful connections. This window may also be seen during analyses from the main WinLab32 screen under the "System, Diagnostics" path. If the instrument was powered down completely prior to this step, see note in section 6.1.3.

7.3. Pre-Check the following:

7.3.1. Peristaltic pump tubing - Change the sample introduction (Black/Black), internal standard (Orange/Green), and pump drain tubing (Red/Red) daily. Change the rinse station pump tubing if it is flat or has visible damage. Pre-stretch the tubing before placing it around the pump to avoid slack generation and flow rate reduction later on during a run.

7.3.2. Rinse water level - The rinse water should be at least one-fifth full. If it is not, prepare more (see 4.4.).

7.3.3. Ar gas pressure - The Ar pressure should read approximately 100 psi for the Ar flow. **DO NOT adjust the regulator at the wall. The instrument can be damaged if the pressure going to the instrument is too high!!!**

7.3.4. Nebulizer - Check the nebulizer to be sure it is clean and produces a smooth mist. Suitable nebulizers include glass Meinhard, PEEK Mira Mist, or any other nebulizers approved by the group supervisor. Normal operations utilize a PEEK Mira Mist nebulizer. If using a glass Meinhard, be sure to use a "C" type and not a "K" type. The intensities can change and cause instrumental difficulties if the wrong type is used under operating conditions for the other type.

7.3.5. Torch - Normal operation utilizes a modified wet aerosol torch assembly. Check the following: the torch is centered in the RF coil; the slot for radial viewing is centered in the quartz window; the bonnet is clean and the injector tip is clean. If there are deposits on the tip, the torch should be cleaned in the following way: disassemble the torch, soak the torch parts in Aqua Regia (3 parts HCl to one part HNO_3) for 48 hours, rinse well with DI water, and dry thoroughly before reassembling. Refer to the Optima 5300 or 7300 Family Hardware Guide from Perkin Elmer for details on removal and cleaning. The torch itself does not have to be completely clean at the top as the emission lines are viewed through the slot in the torch and/or through the end of the torch. After reassembling the torch, re-align its viewing position (Mn alignment) using the solutions in 4.6 and the spectrometer control option in the WinLab32 software. Document the torch viewing positions and intensities in the instrument maintenance log. The intensity reads should be similar, though slightly lower than previous alignments. The radial and axial positions do not

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have to be exact, but should be around 0.0 nm for radial viewing and 0.0, 15.0 nm for axial. Refer to the WinLab32 Instrument Control Software Guide from Perkin Elmer for details on aligning the torch.

7.3.6. Spray Chamber - For normal operations a Cyclonic Spray Chamber with inner baffle is used. The spray chamber should be visually inspected for particles or abnormalities in the glass. Once the sample/internal standard flow has started the spray chamber should be examined for a smooth mist with no beading on the walls of the chamber.

7.3.7. Chiller - Ensure that the power is on, the temperature setting is 17 °C, the recirculating water pressure is ~ 60 psi and there is enough water for cycling. If the water level is too low, fill the chiller with distilled water and a few drops of algaecide or other acceptable solution.

7.3.8. Drain line - Ensure that the condensate from the spray chamber is being pumped through its drain line in a pulsed stream.

7.3.9. Sample/Internal Standard Solution Flow Rate - When the instrument is turned on, place both the sample and internal standard probes in an acid rinse solution. Check both lines to ensure the flow rates are smooth and continuous by generating several bubbles in each line and following the passage of the bubbles all the way to the spray chamber. Ensure that the bubbles do not get stuck in the introduction lines especially at line connections and the 'y' connector. The sample introduction flow rate should be smooth. The internal standard flow will pulsate somewhat, but this cannot be avoided. Once set, the peristaltic pump tension should not need daily adjustment.

7.3.10. Method Selection in Software - Select the desired method in the WinLab32 software. It contains the associated MSF and IEC files as well as proper plasma and flow rate parameters.

Note: All instrument maintenance should be documented in the instrument maintenance log.

7.4. Torch Lighting and Warm-Up - After all pre-start checks pass inspection, perform the following steps:

7.4.1. Torch Ignition - Open the Plasma icon from the main menu, and click on the "On/Off" toggle button located at upper left. The plasma ignition sequence consists of an initial 45 second gas purge, a brief stabilization period in which the pump is turned off, and then a 15-second "final purge" before ignition. Once the plasma is ignited, the pump will be turned back on automatically after ~ 15 more seconds.

7.4.2. Sippers - Manually place the sample and internal standard sippers (probes) in the rinse solution while the instrument warms up.

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7.4.3. Warm-up - Allow the torch assembly/RF generator system to warm up for at least one hour to ensure that the torch and optical system are at equilibrium before starting a run. The detector system is ready to go unless the instrument was placed in "STANDBY" Mode from a previous analysis (see 6.3).

7.5. Autosampler Table Set-up - While the instrument is warming up, a new "Sample Information File" can be created for a new analytical run. Create the new file as follows:

7.5.1. Click on the "SamInfo" icon at the top of the main menu to bring up the file from the most recent run.

7.5.2. The "Sample Information Editor" window will appear with the name of the file at the top. The lower section of the window displays "Parameters that vary by sample". Modify this section for each new run as follows.

7.5.2.1. The leftmost column labeled "Sample No." cannot be edited. To insert a row in the table, click on a row in this column and select "Ctrl-I" on the keyboard to insert a row before the highlighted row, to delete a row, click on the row in this column and press "Ctrl-D" to delete the highlighted row.

7.5.2.2. In the column labeled "A/S Location" type in the autosampler rack position number in which you want that sample analysis tube. Rack position number 16 is the first available entry, after the initial QC samples.

7.5.2.3. Enter the "Sample ID" in its column and include in the sample ID, any dilution factors, reruns, or other important notes about the sample.

Note: Only use letters and numbers in this column. Do **not** use any non-alphanumeric characters (commas, parentheses, asterisks, etc.) in the Sample ID column; these will cause errors when converting the file into QC manager.

7.5.2.4. In the column labeled "ANALYZE QCs BEFORE" and in the first row, make sure the proper sequence of QC solutions appears. See Appendix C for an example of the autosampler sequence that includes the name, rack position number, and purpose of each beginning QC. Place these QC solutions in the rack according to their position in Appendix C (these positions can also be found in the method file in the WinLab32 software).

7.5.2.5. Following the initial QC set and at the end of the run, insert the QC sequence number 13, 14, 15, 16, 17, 20 (see Appendix C or the method file) in the column. Either QC #13 (silver CCV), QC #14 (minerals CCV), #15 (trace metals CCV), or QC #20 lithium

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CCV) may be left out if none of those analytes are needed in the run.

7.5.2.6. The calibration standards will automatically appear when beginning an analysis in the Manual or Automated Analysis Control windows and are not listed in the sequence.

7.5.3. Wash Time - To add additional wash time to the preset wash time in the method (e.g. 60 seconds), type in the number of seconds in this column in the Autosampler file. Use extra wash after high-conductivity (over 20,000 µmhos/cm) samples or samples with a heavy matrix. The wash time that you enter in the Autosampler file will be added to the default time after the sample ID on that same row. The default wash time is preset in the Method Editor, "Sampler" bottom tab, "Autosampler" side tab.

7.6. Save and Print Autosampler Table

7.6.1. From the main menu select *File*, *Save as, Sample Information File* and assign a name in the form of *yyyymmdd-instrument #* to save the file. For example, for a run on the 5300 instrument, on May 1, 2014 the sample information file name would be 20140501-5300 (the extension is automatically set to ".sif").

7.6.2. Select *File*, *Print*, *Active Window* to print the file.

7.7. Loading Samples

Pour the calibration standards, QC standards, and samples into the autosampler tubes and place them in the appropriate positions according to the sample information file. See Appendix C or the method file for positioning of the calibration and QC standards. Make sure there is sufficient QC solution volume in the QC tubes for the number of times they will be analyzed. The instruments usually use between 4 and 8 mL of volume depending on the analyte list and method.

7.8. Running Samples

7.8.1. Starting the analytical run

7.8.1.1. From the main menu select the *Auto* icon to bring up *the Automated Analysis Control* window. In the "Set Up" tab, make sure the proper method is displayed.

7.8.1.2. If the Sample Information File listed is not the desired one, click on OPEN and choose the appropriate file.

7.8.1.3. Choose a results data set name by clicking on OPEN and entering the desired name (the results data set name cannot be manually entered directly into its field). Create the results data set name in the format: "yyyymmdd-instrument" (e.g. 20140501-5300 for May 1, 2014 on the 5300).

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7.8.1.4. Make sure that the "Use Sample Info" and "Save Data" boxes are checked.

7.8.1.5. If the analyst will monitor the analytical run to its completion, skip to 7.8.1.8. and see 7.8.4. for manual shutdown of plasma. Otherwise, to set the instrument for automatic shutdown, select the "SET..." button next to "Auto Shutdown."

7.8.1.6. In the *Automatic Shutdown/Start-Up* window, check "Shutdown," "Begin at end of analysis," "Wash before shutdown" (assign at least 5 minutes time), and "Turn off Plasma and Pump."

7.8.1.7. Do not check the "Put spectrometer into Standby" option unless the instrument will be off for several days (see section 6 above). Also, do not select the "Start-Up" menu in the lower half of the window UNLESS the Standby option has been selected. The start-up option will initiate the 74-minute stabilization period for the SCD system. Under the Start-Up menu do NOT select "Turn on plasma and pump." Click on OK.

7.8.1.8. Select the "Auto Wavelength Realign" and "Set..." buttons. In the *Automatic Wavelength Realign* window, select "At start of analyses" and click OK.

The Hg realignment process closes the shutter to the spectrometer and turns on the Hg lamp. It is used to align the optical signal properly on the SCD. The Hg realignment should be done at the beginning of an analytical run when the instrument has had major maintenance done or where repetitive instrument QC failures occur. When this realignment is performed, document the slit adjustment values and the date completed in the instrument maintenance logbook.

Note: Any adjustments made by the Hg realignment process will affect ALL analyte wavelengths. If an individual peak is found to be shifting during analyses, using the Hg realign again will not correct the problem.

7.8.1.9. Do NOT select the "Print log during analyses" box.

7.8.1.10. Select the "Analyze" tab in the *Automated Analysis Control* window.

7.8.1.11. Select the "Rebuild List" button and scroll through the sample IDs to make sure that the sample information file is correct, including proper placement of QC samples. See Appendix C for an example table.

7.8.1.12. Place the internal standard sipper into a 10 mg/L Sc solution and the sample sipper into the autosampler wash well. Let these

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solutions aspirate for at least 2 minutes to clear bubbles from the lines.

7.8.1.13. Click on the "Analyze All" button and the analysis should begin with the Hg lamp realignment as sequence #1. If the Hg realignment does not need to be completed (see 7.8.1.8), **immediately** click the "Analyze All" button again to stop the run. Click the "Analyze All" button a second time to restart the run at the appropriate sequence number (not the autosampler location number).

7.8.1.14. From the main menu select the *Results* icon to monitor the results. If you wish to monitor the four individual replicate results, select from the main menu the *MethEd* icon, "Options" tab, and click the "Replicate Data" box under "Sample Items."

7.8.2. Calibration Procedure - Monitor the calibration standards as they are analyzed. Check that the internal standard counts in the INITIAL BLANK standard (displayed in the *Results* window) are similar in magnitude to the previous run. Also check that the precisions (RSD) of the four internal standard replicates and the four calibration standard replicates are not greater than 4 %. If the RSDs are greater than 4 %, stop the analysis and re-optimize the sample introduction system. The instrument calibration must be re-started after optimizing the sample introduction system. To monitor the calibration slope for one or more analytes, view the *Results* window immediately after the final standard has been analyzed. The Optima 5300 and 7300 use a "Linear, calculated intercept" fit whereby the calibration blank intensities (relative to the internal standard) are set at zero concentration.

7.8.3. Check the analytical run - Ensure that recoveries of the initial QC samples are within the control limits (see DEP SOP MT-058).

Note: Any QC sample that was entered into the Sample Information File under the "Analyze QCs Before" column will yield a "Passed/Failed" flag displayed in the *Automated Analysis Control* window during the analytical run. More detailed information on which specific analytes passed/failed and their percent recoveries can be found in the results display window.

7.8.4. Manually extinguishing the PLASMA - If the operator is still present when the analytical run is complete, shut down the plasma by pushing the F9 key on the keyboard. If the software is hanging for some reason or communication between software and instrument is not working properly, push the red EMERGENCY PLASMA OFF button on the front of the instrument to bypass the software. This option will not harm the instrument. However, to be able to re-ignite the plasma in this case, you

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must reset the system by clicking on the *System* icon, and "Reset Emergency Plasma Off."

8. GENERATION OF REPORTS AND DATA ANALYSIS FILES

8.1. Printing Raw Data. The raw data, consisting of the individual replicate data for each sample analyzed, should NOT be printed out unless special circumstances warrant a hard copy.

8.2. Data Report Generator. At the end of an analytical run, the raw data is to be saved both in a "Report" format and in a .csv file format using the "Export" feature. The report file format is saved directly to the directory on \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV.

8.2.1. From the main menu in WinLab32, select *File, Utilities, Data Manager* (this application can also be accessed from the windows start menu).

8.2.2. Highlight the data set desired and click on the *Report* icon to bring up the "Data Reporting Wizard" window.

8.2.3. Select "Use existing design" and the Browse button to bring up the current list of report templates.

8.2.4. Open the template "vertical_detailed 2.rep" and click "Preview" from the Data Reporting Wizard window. Do not click on "Finish."

8.2.5. When the actual report is displayed in the Preview window, click on the icon that looks like an envelope. This will bring up the "Export" window.

Note: This "Export" window does not mean the same thing as the *EXPORT* function in Data Manager.

8.2.6. Choose "PDF" under Format, and Disk file (default). Click Okay. This will open the "Choose Export File" window.

8.2.7. Change the destination path (Save in) to \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV. Change the file name to that of the results data set name and click 'Save'. Close the Preview window and return to the main Data Manager window by choosing "Cancel" in the "Data Reporting Wizard" window.

8.3. Export File (.csv format) Generation. The Export file (.csv) is written automatically to the directory C:\pe\Metals\Reports (but note - this is not the "report" version of the data). The Export file is then copied in 'Windows Explorer' to the directory \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV.

Note: Sections 8.3.1-8.3.6 only need to be completed if the results are not auto exported or if the data was reprocessed.

8.3.1. From the main menu in WinLab32, select *File, Utilities, Data Manager*.

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8.3.2. Highlight the data set desired and click on the *Export* icon to bring up the "Data Reporting Wizard" window.

8.3.3. In the "Data Exporting Wizard" window, select "use existing design" and the browse button to bring up the current list of export templates.

8.3.4. Open the "exportICP.xpt" design.

8.3.5. Click "Finish" to get to window # 8, "Export Data Set." and click "Export data".

8.3.6. Click "Finish" and exit the Data Manager program.

8.3.7. Go to Windows start menu and copy the Exported file (.csv format) from the directory C:\pe\Metals\Reports directory to the directory \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV.

8.4. Exporting the Method Parameters. The method parameters for each run should be exported using the following steps:

8.4.1. Click on the MethEd tab to open the method editor.

8.4.2. Click File, Print, Active Window.

8.4.3. Change the printer name to the XPS document writer.

8.4.4. Save method parameters to \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV folder named as yyyymmdd-instrument Method Parameters.

8.5. Exporting the MSF Model. The MSF model for each run should be exported using the following steps:

8.5.1. Click on the Examine tab to open the examine spectra/MSF window in MSF mode.

8.5.2. Load current MSF file and open model from the FILE tab.

8.5.3. Right click on the blue background, and chose MSF File Viewer/ Editor from the dropdown menu.

8.5.4. Click expand all, Print.

8.5.5. Change the printer to the XPS document writer.

8.5.6. Save MSF model to \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV folder named as yyyymmdd-MSF File.

8.6. Exporting the Sample Information Report. The sample information report for each run should be exported using the following steps:

8.6.1. Click on the Sample Information Editor window.

8.6.2. Click File, Print, Active Window.

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8.6.3. Change the printer name to the XPS document writer.

8.6.4. Save sample information report to \\FLDEP1\DEAR\Labs\Lab3\Chem_Data\Inorganic\Metals\ICPAESDV folder named as yyyymmdd-Sample Info File.

8.7. Converting Data File. The exported (.csv) file on the network is converted and loaded in the QC Manager.

8.7.1. In LIMS, follow the path "Laboratory," "QC," "QC Manager" and open the program.

8.7.2. Make sure the Instrument window at upper left reads "Metals: ICP-AES-Perkin Elmer 5300 or 7300." If not, select this option from the drop-down window and ensure you have the appropriate instrument selected.

8.7.3. Select the convert instrument file option, F9 function key, to convert files. This will bring up the "Select Instrument File" window. Choose the proper data set from the list and click Open.

8.7.4. You will immediately be taken to the "Save To LIMS" window and the .csv file name will be listed under 'Save To LIMS as…'. Choose OK to save the data.

8.7.5. You will then be asked to either "Convert" or to "Close" the instrument file. Choose "Convert" and the RAW data will appear in QC Manager, ready for review.

8.8. Archiving and Deleting Data. On a bi-monthly basis archive the WinLab32 Results database to a zip directory, and then delete all old data sets located in the directory

8.8.1. Open Data Manager from outside WinLab32 by selecting *Start, Programs, PerkinElmerWinLab32, Data Manager*. Close the WinLab32 software.

8.8.2. Highlight all files needed to be archived, then press the icon *Archive* and save it as *monthyear.zip* file in the directory E:\\Archived data.

8.8.3. Again, highlight the same files and press the icons *Delete*, then *Pack*, then *Check* (if error appears, press *Check* again).

9. DATA PROCESSING, REVIEW, AND CALCULATIONS

The procedure for data review, cross-checking and any calculations is summarized in DEP SOP **MT-058**, which also lists the method performance acceptance criteria for standard reference materials, matrix spikes, internal standards, and precision data. All aspects relating to tolerance limits, corrective actions for out of control data, and contingencies for handling unacceptable data are contained in this SOP as well as DEP SOP **LB-027**.

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10. SAFETY/HAZARDOUS WASTE MANAGEMENT

10.1. Review the Health and Safety Plan found on the DEAR website.

10.2. Use **CAUTION** with strong irritants such as acids and bases. Avoid breathing the fumes of these irritants by using them in a hood when possible and keeping the face away from open containers of these chemicals. Avoid contact of these irritants with skin and clothing by appropriate use of gloves, apron, face-mask, hood shield, etc. Safety glasses and lab coats must be worn all the time in the laboratory.

10.3. Store any wastes that cannot be disposed of down the sink in a hazardous waste container (properly labeled with **Hazardous Wastes** labels) in the hood. Make sure the bottle stays capped after use and place the bottle in the cabinet below the hood in the metals lab when it is 90 % full. When all containers are nearly full dispose of the waste into the appropriate acidified metals hazardous waste drum located in Bldg C.

10.4. All the non-hazardous wastewater can be disposed of down the sink if the acid or base concentration is diluted to 2 % or diluted by 50 or greater. If it is not, the wastewater has to be neutralized before disposing.

11. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

All samples are stored in the DEP Central Laboratory's Receiving Room. The sample pH for water matrices is checked upon arrival at the laboratory by receiving room staff to ensure it is less than 2. Samples consisting of water matrices are stored at room temperature and samples consisting of solid matrices are stored in one of the receiving room refrigerators.

12. INTERFERENCES

See Appendix E for a list of possible interferences that can be encountered during the analysis of environmental samples.

13. REFERENCES

13.1. EPA Method 200.7, Revision 4.4, 1994

13.2. EPA Method 6010C

13.3. SM 2540 B

13.4. Optima 5000 and 7000 Family Hardware Guide and ICP WinLab Software Guide by Perkin Elmer.

13.5. DEP SOP MT-024, Preparation of Water Samples for Metals Analysis by ICP-AES and ICP-MS

13.6. DEP SOP MT-055, Determination of Percent Dry Solids

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13.7. DEP SOP MT-058, Procedure for Metals Data Review and Authorization

13.8. DEP SOP MT-060, Hot Block Digestion of Soil, Sediment, Waste and Tissue Samples for Total Recoverable Metals

13.9. DEP SOP MT-063, Microwave Assisted Hydrofluoric Acid Digestion of Solid Matrices for Total Metals Determinations

13.10. DEP SOP MT-068, The Toxicity Characteristic Leaching Procedure (EPA Method 1311) and the Synthetic Precipitation Leaching Procedure (EPA Method 1312)

13.11. DEP SOP MT-069, Metals Standard and Reagent Preparation Procedure

13.12. DEP SOP LB-027, Standard Operating Procedure for Reporting Qualified Data

13.13. Quality Manual, Bureau of Laboratories, DEP

13.14. Laboratory Health and Safety Plan

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Appendix A

ICP-AES Interference Check Standard Concentrations (ICS2007 see 4.12.5.)

Purchased Solution	Analyte	Concentration
ICS #5	Al	60 mg/L
ICS #18	As	1 mg/L
ICS #18	Ag	300 µg/L
ICS #3	B	500 µg/L
ICS #18	Ba	300 µg/L
ICS #18	Be	100 µg/L
ICS #5	Ca	300 mg/L
ICS #18	Cd	300 µg/L
ICS #18	Co	300 µg/L
ICS #18	Cr	300 µg/L
ICS #18	Cu	300 µg/L
ICS #5	Fe	250 mg/L
ICS #18	K	20 mg/L
ICS #5	Mg	150 mg/L
ICS #18	Mn	200 µg/L
ICS #3	Mo	300 µg/L
ICS #5	Na	50 mg/L
ICS #18	Ni	300 µg/L
ICS #18	Pb	1 mg/L
Stock Standard	Sb	1 mg/L
ICS #18	Se	500 µg/L
ICS #3	SiO_2	492 µg/L
ICS #3	Ti	1 mg/L
ICS #18	Tl	1 mg/L
ICS #18	V	300 µg/L
ICS #18	Zn	300 µg/L

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Appendix B

Spike Solutions

Metal	Water Spike HW6 (1/50 dilution) µg/L	Sediment Spike SEDSPK4 (1/50 dilution) µg/L	Instrument/Digested Mineral Spike ICPMINA (1/50 dilution) mg/L	SILICA Spike
Al	4000	8000	20	
As	200	400		
Sb	200*	400**		
B	500	400		
Ba	400	800		
Be	2	4		
Cd	10	20		
Ca	4000	8000	80	
Cr	100	400		
Co	100	200		
Cu	100	100		
Fe	4000	8000	20	
Pb	100	400		
Li	100			
Mg	4000	8000	40	
Mn	200	400		
Mo	50*	200**		
Ni	100	200		
K	4000	8000	10	
Se	200	400		
Ag	20*	40**		
Na	4000	8000	80	
Sr	2000	4000		
Tl	50	100		
Sn	100	400		
Ti	40*	200**		
V	100	200		
Zn	500	400		
U	50	50		
SiO_2				10

* Part of HW6B spike
** Part of SED SPK4B

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Appendix C
Autosampler Table Format

Automated Analysis Control Seq. #	Autosampler Rack Position #	QC[a] or Sample	Purpose	QC # (from method)[b]	"ANALYZE QCs BEFORE" sequence in Sample Information File
1	1	INITIAL BLANK	Calibration Blank		
2	2	ICPMIN	Minerals calibration standard		
3	3	ICPMET	Trace metals calibration standard		
4	13	AG-SIO2	Silver/Silica calibration standard		
5	14	AG-SIO2C	second source silver/silica calibration check	1	
6	10	ICPMETC	second source trace metal calibration check	2	
7	9	ICPMINC	second source mineral calibration check	3	
8	5	ACID-RINSE	eliminate carryover	4	
9	11	CPQLDV11	check instrument PQL levels	6	
10	12	ICS2007	interelement interference check	7	
11	6	ICPMINCCVC	initial calibration verification for minerals	8	
12	7	ICPMETCCVC	initial calibration verification for trace metals	9	
13	4	AG-SIO2CCVC	initial calibration verification for silver + silica	10	
14	5	ACID-RINSE	eliminate carryover	11	
15	1	CCB	verify no carryover exists into first sample	12	
16	15	1st Sample			1-4,6-12
17-25	16-24	2nd – 10th Samples			
26	4	AG-SIO2CCV	CCV	13	
27	6	ICPMINCCV	CCV	14	
28	7	ICPMETCCV	CCV	15	
29	5	ACID-RINSE	eliminate carryover	16	
30	1	CCB	see above	17	
31	25	11th Sample			13-17 [c]
32-40	26-34	12th - 20th samples			
41	4	AG-SIO2CCV	CCV	13	
42	6	ICPMINCCV	CCV	14	
43	7	ICPMETCCV	CCV	15	
44	5	ACID-RINSE	eliminate carryover	16	
45	1	CCB	see above	17	
46	35	21st sample			13-17 [c]
47-56	36-44	22nd - 30th samples, etc.	Run must end with CCV's and a CCB		

a For analyte concentrations refer to section 4.

b QC sequence numbers assigned in the Method Editor, "QC" bottom tab. These numbers are used in the Sample Information File.

c The analyst can invert the order of CCVs, skip the acid rinse (but then add extra rinse time after last CCV), or, either the ICPMETCCV, ICPMINCCV or AG-SIO2CCV can be skipped if none of the analytes in those solutions are requested in the 20 samples surrounding this set of QC.

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Appendix D

MSF Model Preparation Notes

Models should be recreated anytime the detector is adjusted or replaced.

1) Read about the MSF model in the WinLab32 Instrument Control Software manual.
2) Open the MSF method from the methods option list. Click on the MethEd icon to make sure all the peak algorithms in the Peak Processing tab of the Process page are set to Peak Area.
3) Make sure there is no MSF model chosen in the Spectral Corrections tab on the Process page.
4) The peak wavelengths for all analytes need to be set to the peak maximum. Do this by pulling up the most recent full analyte list run in the examine window and verifying that each wavelength is in the center of the peak. See the WinLab32 Instrument Control Software manual for details.
5) Make sure that Spectral Profiling is not selected on the Settings tab on the Spectrometer page. The methods currently used do not use Spectral Profiling.
6) The scandium internal standard counts in the initial blank and all calibration standard analytes need to have RSDs less than 3 % to get the best MSF model possible.
7) Run the MSF run using single element standards for all analytes and possible interferences. Use multiple sources and concentrations when possible. Because these are approximate concentration and MSF is concentration independent, expired stock standards can be used to prepare these standards.

Note: All samples must have a different name in the Sample info file.

8) Use the examine window to create an MSF model by selecting “MSF” in the dropdown window at the upper left corner of the examine window (background should turn blue). If working on an old model click “File” “Open” “MSF Model”.
9) Right click on blank and click “Assign global blank”. This designates the initial blank on all analytes. Display the internal standard (radial and axial) in the examine window (you can only look at one at a time), highlight the initial blank in the sample list and type “C” to clear the blank.
10) Assign individual analytes by opening a particular analyte in the examine window, highlighting the sample that corresponds with that analyte (only one concentration) and typing “A” for analyte.
11) Use the examine window to determine interfering peaks and assigning them by highlighting the sample and typing “I” for interferent. Old models can also be used as a basis for what the interferences for a particular analyte may be. It can be helpful to zoom in to be able to see interfering peaks.
12) Save changes to the MSF model by clicking “File” “Save” “MSF Model”.
13) Apply the new MSF model to the current method by clicking on the MethEd icon. In the Spectral Corrections tab on the Process page select the new MSF model (this needs to be reselected even if the model has the same name). Save the method.
14) Reprocess an initial calibration to determine if model is working, make changes as necessary.

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15) After the MSF model is completed, a MSF check needs to be run. This consists of 10 of each: CCB, D-BLK, DPQL, CPQL, and LFBHW. Certified standard reference materials should also be analyzed to verify proper interference correction.

IEC Model Preparation Notes

Models should be recreated anytime the detector is adjusted or replaced.

1) Run an MSF run sequence using the single element stock standards following the same procedure as dictated in the MSF model preparation notes through #7.
2) View results in examine window and apply background correction points (depicted as green cross hairs along the baseline). One point if the baseline is noisy but flat across, two points if the baseline is at an angle and none if the baseline is clean. The crosshairs should be placed in a location where there are no interfering peaks or raised baselines, it is important to zoom in on the baseline when selecting base points in order to prevent false positives or false negatives due to under or over correction. To increase the number of base points type "M" to pull up the method option window and change the number of background points (none, one or two).
3) Change the integration window to include the whole peak by typing "M" to pull up the update method parameters box. Here you can change the number of points/peak, generally the number of integration points should be between 1 and 4 depending on the peak shape and width but is ultimately peak dependent. The actual pixels integrated are displayed in the examine window as small green squares along the peak lines while the large blacked out rectangle indicates the integration area.
4) Reprocess and save the MSF run as a new file with the new base points and integration windows set.
5) Select "Tools" "IEC Model Builder" and open the MSF results data set.
6) Set limits generically to 20 µg/L. Adjust accordingly based on analyte and MDL.
7) Click on the Calculate Factors tab. Set the interfering analyte to the analyte that corresponds with the sample ID (i.e. for sample ID Al 500 mg/L the interfering analyte would be Aluminum_396). Select Scandium (both axial and radial) from the analyte selection option at the top of the window and select "set to zero" for all analytes.
8) The summarize factors tab gives a table with the interference and the value calculated based on the model. Click "only non-zero factors" to view only the analytes with IEC factors.
9) Click "File" "Save" "IEC Model" and in the IEC model builder click update method. Then save the method.
10) Reprocess the initial calibration data in WinLab32 off-line to check that all QC is passing. Be sure to open the appropriate method.
11) Some samples will have unnecessary IEC factors that can be eliminated without changing the results; this should be done on an analyte by analyte basis.
12) Changes can also be made to the number of integration points and the location of integration window. The yellow line indicates the peaks center and can be moved by clicking in the examine window and using the left and right arrow keys. The peak can be set by typing "P" this will shift the black

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integration window and indicate the current pixels being integrated based on the peak placement. The method parameters can be saved from this window by typing "M" and "S".

13) IEC factors can manually be calculated if the current IEC factor is not working by using the results from the ICS2007 standard. The IEC factor is calculated as the actual concentration of the affected analyte in the ICS2007 standard minus the expected concentration of the analyte in the ICS2007 standard in µg/L divided by the actual concentration of the known or suspected interferent in mg/L. The resulting number is added to the current IEC factor. If these changes cause drastic failures the IEC factor is probably not the problem, try changing the peak location and the number of integration points. If only the PQL is failing try to adjust the location of the background points.

Note: A positive factor will decrease recoveries while a negative factor will increase recoveries.

14) After the IEC model is completed, an IEC check needs to be run. This consists of 10 of each: CCB, D-BLK, DPQL, CPQL, and LFBHW. Certified standard reference materials should also be analyzed to verify proper interference correction.

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Appendix E

Method Parameters

Method Editor: Spectrometer Page: Settings

Spectral		
	Purge Gas Flow	High
	Spectral Profiling	No
	Resolution	Fixed Normal

Read Parameters			**Min**	**Max**
	Time (sec)	Auto	2.0	20
	Delay time (sec)	25		
	Replicates	4		

Method Editor: Sampler Page: Plasma

Plasma (L/min)	15
Aux (L/min)	0.2
Neb (L/min)	0.6
Power (watts)	1500
View Distance	15

Source Equilibrium Delay (sec)	10
Plasma Conditions	Vary by Element

Method Editor: Sampler Page: Peristaltic Pump

Pump Parameters	
Sample Flow Rate (mL/min)	1.5
Flush Time (sec)	30

Method Editor: Sampler Page: Autosampler

Wash		
Location: 0	**Frequency**	After every sample + extra time
	Rate (mL/min)	1.5
	Normal Time (sec)	60
	Extra Time (sec)	120

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Analyte	Approximate Wavelength in nm	Plasma View	Possible Interferences
Al	396	Radial	
As	189	Axial	Cr, Fe
Sb	207	Axial	Cr, Fe
B	209	Axial	
Ba	234	Axial	
Be	313	Axial	
Cd	229	Axial	As, Ca, Fe
Ca	318	Radial	
Cr	268	Axial	
Co	229	Axial	
Cu	327	Axial	
Fe	274	Radial	
Pb	220	Axial	Ca, Fe
Mg	279	Radial	
Mn	258	Axial	Fe
Mo	202	Axial	Fe
Ni	232	Axial	Fe
K	766	Radial	
Se	196	Axial	Ca, Fe
Ag	328	Axial	Fe, Ti
Na	590	Radial	
Sr	422	Radial	
Tl	191	Axial	Fe, Ti, V
Sn	190	Axial	Ca, Ti
Ti	335	Axial	Ca
V	292	Axial	Fe, Mo
Zn	206	Axial	Cr
SiO_2	252	Radial	Mo
Sc	357	Axial	Internal Standard
Sc	361	Radial	Internal Standard
Li*	671	Axial	

*Lithium analysis performed on the Perkin Elmer ICPDV 7300 only.

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Appendix F

If results from TCLP samples are within 20 % of the regulatory limits and the spike recoveries are less than 50 %, the method of standard additions (MSA) must be used to quantify the samples. MSA is used to account for matrix affects in a sample. In this method the calibration standards are made in the sample matrix rather than in blank solution.

MSA requires that the calibration curve be linear up to the most concentrated addition and that all volumes be kept equal for all additions. Four identical aliquots of the sample are taken and prepared according to the chart below. All four aliquots are analyzed using CCB solution (2 % HNO_3 and 1 % HCl) as the blank.

			Unspiked Aliquot				1st Aliquot (50 % RCRA violation levels)		
	Spiking Solution		Sample	Spike Soln	CCB		Sample	Spike Soln	CCB
	mg/L		mL	mL	mL		mL	mL	mL
As	10		4	0	4		4	0.1	3.9
Ba	20		4	0	4		4	1	3
Cd	2		4	0	4		4	0.1	3.9
Cr	10		4	0	4		4	0.1	3.9
Pb	10		4	0	4		4	0.1	3.9
Se	2		4	0	4		4	0.1	3.9
Ag	10		4	0	4		4	0.1	3.9
			2nd Aliquot (100 % RCRA violation levels)				3rd Aliquot (150 % RCRA violation levels)		
			Sample	Spike Soln	CCB		Sample	Spike Soln	CCB
			mL	mL	mL		mL	mL	mL
As			4	0.2	3.8		4	0.3	3.7
Ba			4	2	2		4	3	1
Cd			4	0.2	3.8		4	0.3	3.7
Cr			4	0.2	3.8		4	0.3	3.7
Pb			4	0.2	3.8		4	0.3	3.7
Se			4	0.2	3.8		4	0.3	3.7
Ag			4	0.2	3.8		4	0.3	3.7

Complete the MSAWorksheet template (the template can be found at \\FLDEP1\DEAR\Labs\Lab5\Chemistry\Metals\TCLP analysis worksheets\MSA) by entering the spike values and sample raw results in µg/L. The worksheet will calculate the equation of the line. Fill in the X-intercept and slope boxes based on this equation. The resulting independent X axis value must be multiplied by 2 (to account for MSA dilution) and then by 10 (to account for TCLP preparation dilution). Divide by 1000 to convert this result to mg/L.

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To report the calculated result in the QCM file click on the LIMS result and change the sample result. Comment that the MSA method was used and comment if the sample results exceed the TCLP limit. Attach the MSA worksheet to the run cover page.

Appendix G

Appendix of Significant Changes

1. The Limit of Quantitation requirement, control limits, and corrective actions were added to Section 2, Summary of Method.
2. The matrix spike frequency was added to Section 5, Sample Preparation Procedure.
3. 10/4/2010 Instrument names updated to include 7300
4. 10/4/2010 References updated to include new SOP numbers
5. 10/4/2010 Information regarding interference models was added to Appendix D.
6. 10/4/2010 Appendix F: Method of Standard Additions added to SOP
7. 10/21/2015 Lithium added as an analyte.

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