Mounting of Worms, Midges, and Aquatic Mites

(Based on Epler, 2001, Identification manual for the larval Chironomidae (Diptera) of North and South Carolina. A guide to the taxonomy of the midges of the southeastern United States, including Florida and Pluchino, 1984, Guide to the common water mite genera of Florida)

1. SCOPE AND APPLICATION

1.1 This method details the slide mounting of macroinvertebrate specimens, such as worms, midges and aquatic mites (if necessary), or parts of specimens, such as labrums, mouthparts, parapodia, etc., for taxonomic identification.

1.2 This method is to be used for all samples with Laboratory Information Management System (LIMS) test codes beginning with MI- and not ending in -SORT.

2. SUMMARY OF METHOD

2.1 Specimens are mounted on glass microscope slides under glass coverslips using CMC-10 Mounting Media. Slides are then either air dried or dried in an oven at 85° to 95° C for 1-2 hours. Slides can be dried for 2-4 hours at 50° C if there are problems, such as boiling media or air pockets forming under the coverslips, at the higher temperatures.

3. APPARATUS AND EQUIPMENT

3.1 Dissecting microscope
3.2 Forceps
3.3 Glass microscope slides, 25 x 75 mm
3.4 Cover slips, round, 12 and 18 mm diameter
3.5 Labels for slides

4. REAGENTS AND CHEMICALS

4.1 Mounting medium (CMC-10)

5. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

5.1 Samples are collected, preserved, and delivered to the lab according to SOP FS 7000 – General Biological Community Sampling, SCI 1000 – Stream Condition Index Methods, or BRN 1000 – Biological Reconnaissance Field Methods.

5.2 Samples are checked into the laboratory per Standard Operating Procedure (SOP) IZ-01.

6. SAMPLE PREPARATION PROCEDURE

6.1 Samples are prepared per SOP IZ-02, IZ-03, IZ-04, or IZ-39

7. SAMPLE ANALYSIS PROCEDURE

7.1 Samples are analyzed per SOP IZ-06 and IZ-07.

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7.2 Using the Laboratory/Receiving/Print Preparation Labels utility in LIMS, print out slide labels. All labels are printed from the Sato printer.

7.2.1 To print slide labels, go to the sample Log-in station, open LIMS and select "Laboratory", "Receiving", "Print Preparation Labels." Go to "Invert_Labels". For Stream Condition Index (SCI) samples with replicates, choose either "Invert Slide Labels Rep 1" or "Invert Slide Labels Rep 2" as appropriate. For all other samples, choose "Invert Slide Labels". Scan the sample bar code, select "File", "Print", select the "Generic / Text only" printer, and print the number of labels needed.

7.3 Affix a slide label to each slide by wrapping the label around the right edge of a slide.

7.4 Starting on the far left of the slide, place a drop of CMC on the slide for each specimen being mounted. Use enough CMC to form a seal around the coverslip when the specimen is pressed down.

7.4.1 Mount 4 specimens per slide under 12 mm cover slips. In the case of very large specimens, two are mounted per slide under 18 mm coverslips.

7.5 Using a needle or forceps, place the specimen in the CMC. Midges and mites (if necessary) should be mounted ventral side up and upside down.

7.5.1 Mounting of aquatic mites or other large specimens requires more CMC, otherwise air will accumulate under the coverslip and the necessary structures will not be visible. Mites should be cleared in a 1N KOH solution and then rinsed in DI water before mounting.

7.6 Using forceps, place the coverslip over the specimen.

7.6.1 Place the coverslip over the specimen at an angle in order to reduce air bubbles and roll the specimen into position.

7.7 Using forceps or the eraser of a pencil, press the coverslip onto the specimen. At this point, orient the specimen by pushing or pressing the coverslip in various ways.

7.7.1 Usually, midges are "C" shaped when preserved, and can be rolled into position when the coverslip is placed over the specimen. Sufficient pressure must be applied to spread the mandibles and labrum. Be careful not to break the coverslip, as the pressure required to flatten the specimen may be great.

7.8 Place the slides on a slide tray and set it aside to dry. It may take 1 to 2 days for the slides to dry completely at room temperature. Alternatively, the slide may be placed in a drying oven at 85° to 95° C for. 1-2 hours or at 50° C for 2-4 hours if there are problems, such as boiling media or air pockets forming under the coverslips, at higher temperatures.

7.8.1 Other station identifiers along with the total number of slides, the initials of the mounter, and the date mounted can be written on a small piece of paper and attached to the tray on which the slides are placed to dry. This is often helpful when identifying the organisms.

8. QUALITY CONTROL AND DOCUMENTATION

8.1 Florida Department of Environmental Protection (FDEP) Laboratory Freshwater, Marine Invertebrate, or Macroinvertebrate benchsheet. Use a pen with black ink for all logbook and bench sheet entries.

8.2 To correct raw data entries, place a single line through the incorrect entry, write the corrected entry near the error, and initial and date the correction. If the correction requires an explanation, write a comment and reference the error.

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8.3 If a problem occurs during sample processing, submit a nonconformance report (NCR) describing the problem and the corrective action. To create a NCR, open LIMS and go to Laboratory/Administrative/Non-Conformance Report. For a decision tree regarding issuance of NCR's, refer to SOP LB-002.

8.4 Quality Control (QC) of Sorting is performed per SOP IZ-40.

8.5 QC of Taxonomic Identification is performed per SOP IZ-13.

8.6 Refer to the Laboratory Quality Manual.

9. SAFETY/HAZARDOUS WASTE MANAGEMENT

9.1 CMC-10 Mounting Media can cause irritation to eyes and skin. Avoid contact with eyes and skin and avoid inhaling or ingesting. For skin, wash affected areas with soap and water (MSDS says for at least 15 minutes).

9.2 Triple rinse empty containers with water before disposing.

9.3 Refer to the Material Safety Data Sheet (MSDS) for information about reagents and chemicals in this SOP.

9.4 Refer to the Laboratory Health and Safety Plan.

10. REFERENCES

10.1 FDEP Laboratory Quality Manual, http://www.dep.state.fl.us/labs/library/progplan.htm

10.2 FDEP Laboratory Health and Safety Plan, https://www.fldepnet.org/content/dear/bureau-laboratories-safety-documents

10.3 SOP FS-7000 General Biological Community Sampling

10.4 SOP SCI-1000 Stream Condition Index Methods

10.5 SOP BRN-1000 Biological Reconnaissance Field Methods

10.6 SOP IZ-01 Sample Arrival/Log-in Procedures

10.7 SOP IZ-02 Stream Condition Index (SCI) Sample Preparation

10.8 SOP IZ-03 Invertebrate Hester-Dendy Sample Preparation

10.9 SOP IZ-04 Invertebrate Core/Grab/Dredge Sample Preparation

10.10 SOP IZ-06 Invertebrate Enumeration and Taxonomic Analysis

10.11 SOP IZ-07 Subsampling of Worms and Midges

10.12 SOP IZ-13 Invertebrate Taxonomic Identification Quality Control

10.13 SOP IZ-39 Macroinvertebrate Subsampling Sample Preparation

10.14 SOP IZ-12 Invertebrate Sorting Quality Control (QC)

10.15 SOP LB-002 Non-conformance Reporting System

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Appendix of Changes

12/21/2009 Reformatted the layout following SOP LB-001.

11/10/2011 Updated date for Epler's taxonomic key. Clarified slide drying process. Updated name for the Laboratory Safety Manual.

11/04/2013 Revised scope to apply to any specimen or part of specimen from any type of macroinvertebrate sample.

10/27/2014 Formatted for 508 compliance.

11/05/2015 Updated SOP references.

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