Preparation of Samples for qPCR Analysis

1. SCOPE AND APPLICATION

1.1. This method describes the preparation of surface water samples by membrane filtration and weighing of solid (sediment, biological solids) samples for qPCR assays.

1.2. This procedure is used for samples with LIMS test ID of PCR-HF183, PCR-HF183B, PCR-HF183S, PCR-GULL2, PCR-GULL2B, PCR-GULL2S.

2. SUMMARY OF METHOD

2.1. A water sample is filtered through a membrane filter. The filter containing the bacteria is agitated in a centrifuge tube containing zirconium silica beads using a bead beater to extract the DNA from the bacterial cells into a buffer solution. The clarified extract may also be purified using a purification kit. The extract is analyzed with real-time PCR amplification and detection in a thermocycler system using a primer/probe assay marker.

2.2. Solid samples are weighed on a balance and extracted using a purification kit. The extract is analyzed with real-time PCR amplification and detection in a thermocycler system using a primer/probe assay marker.

2.3. Interferences

2.3.1. Colloidal or suspended particulate materials in water can clog the filter and prevent filtration of a sufficient amount of sample. Some substances, including humic and tannic acids, can act as inhibitors and reduce efficiency by either binding to the DNA polymerase or interfering with the annealing and extension cycles. Sample purification or dilution can help alleviate the effects of inhibitors

3. EQUIPMENT AND SUPPLIES

3.1. Gloves, latex or nitriles

3.2. Tube racks

3.3. Forceps

3.4. Dispersion beads, 0.1 mm, zirconium silica

3.5. Extraction tubes, conical, 2 mL screw cap with o-ring seal

3.5.1. Weigh out 0.3 ± 0.01 g of dispersion beads into each extraction tube. Seal tube tightly; making sure the O-ring is seated properly. Autoclave @ 121°C for 15 min (15 psi). Record in LIMS Autoclave Log.

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3.6. Centrifuge tubes, 2.0 mL

3.7. Polycarbonate membrane filter, 47 mm diameter, 0.45 µm pore size

3.8. Glass Fiber Filter, 47 mm diameter, AP20

3.9. Pipette pump with sterile disposable 50 mL tips

3.10. Magnetic Funnels with 47 mm bases, 500 mL

3.11. Vacuum Filtration Manifold

3.12. Sterile, clean spatulas

3.13. MO BIO PowerLyzer® PowerSoil® DNA Isolation Kit

3.13.1. PowerLyzer® Glass Bead Tubes (0.1 mm)

3.14. Steris AMSCO Model #3021 (S/N RG11420101) autoclave

3.14.1. Calibrate and use per SOP TA-6.18.

3.15. Mettler Precision Balance Model XA105DU (S/N B314195659)

3.16. PCR#5, Thermo Fridge/Freezer (S/N 0121174701121218), maintained at 4ºC ± 2ºC and -20ºC ± 10ºC, Room B246

3.17. UV Sterilizer unit

4. REAGENTS AND STANDARDS

4.1. Deionized Water

4.2. PCR grade water –Autoclave ultrapure water for 15 min. Record in LIMS Autoclave Log.

4.3. Bleach solution, 10% (v/v)

4.4. Phosphate Buffered Saline (PBS) – Prepare per SOP PCR-3.0.

4.5. Ethanol 80%

5. CALIBRATION AND STANDARDIZATION

5.1. See FDEP Bureau of Labs Quality Manual, Section 5.5

5.2. Refer to SOP PCR-2.2 for calibration and use of the balance.

5.3. Refer to SOP PCR-2.5 for calibration and maintenance of support equipment.

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5.4. Clean work area counters after use. Spray surface of work area with 10% bleach solution and wipe down after 15 minutes. Spray and wipe down with sterile DI water.

6. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

6.1. See DEP SOP FS 1000 for sample collection and preservation.

6.2. Refer to SOP PCR-5.0 for Sample custody and labels.

7. SAMPLE FILTRATION

7.1. Turn on UV sterilizers and let warm up for at least 10 minutes.

7.2. Unwrap 3-6 funnels and bases. Place bases with funnels in vacuum manifold. Extra funnels may be placed in the UV sterilizer. Use a new set of funnels and bases daily.

7.3. Place forceps in a small beaker of ethanol. Fill a clean, sterile, dry rinse bottle with buffered dilution water.

7.4. Use sterile forceps to place a sterile glass fiber filter and then a polycarbonate filter on the filter base and attach the funnel to the base. The vacuum may be opened briefly to help seat the filters.

Note: Keep forceps in a small beaker of ethanol during filtration process. Dip in the ethanol and shake off excess before grasping each filter. If forceps are accidentally dropped or placed on counter, replace with a new pair of sterile forceps from the UV sterilizer. Make sure no ethanol residue from the forceps touches the filter.

7.5. Shake the sample container vigorously, approximately 25 times to uniformly distribute bacteria. Carefully open container. Avoid touching the mouth of the container to prevent contamination of the sample.

7.6. With a 50 mL pipette tip and pipette pump, pour 50 mL of sample into the funnel. Continue adding 50 mL aliquots of sample (maximum of 200 mL) until filtration of sample slows down. Typical filtration volumes are 100-200 mL. When sample has filtered through, rinse the walls of the funnel with 20-30 mL of PBS (rinse all the way around 3 times). Record the total volume of sample filtered on the extraction tube label and in prep worksheet manager.

7.7. Turn off vacuum and aseptically fold polycarbonate filter into a cylinder (fold in half, then half again, then roll) with sample side facing inward using sterile forceps and being careful to handle only the edges.

7.8. Place the rolled polycarbonate filter into an extraction tube containing dispersion beads and cap.

7.9. Place a new polycarbonate filter and filter a second aliquot of the sample in the same manner as the first aliquot. Place the second rolled filter in a 2.0 mL centrifuge

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tube. Record the final sample volume on the tube labels. Discard the glass fiber filter.

7.10. Repeat Steps 7.4.-7.9 until all samples are filtered. Make sure funnels have been UV sterilized for at least 2 minutes before starting each new sample.

7.11. Store sample filters at -20°C until analysis. For long term storage (>2 weeks) store at -80°C.

8. SOLID SAMPLE WEIGHING

8.1. Before turning on the balance, snap the weighing basket onto the grid weighing pan.

8.2. Spin down all MOBIO PowerLyzer ® Power Soil® DNA isolation kit bead tubes, to ensure beads are at the bottom of the tube and do not interfere with sample weighing.

8.3. Place an open MOBIO PowerLyzer ® Power Soil® DNA isolation kit bead tube into the basket. Ensure that the doors of the draft shield are all closed and press the [→T←] key to tare the balance. "0.00000 g" should be displayed.

8.4. Add a small amount of the solid sample to be weighed into the bead tube using a sterile spatula. Close the door of the draft shield to prevent drafts from skewing the weight.

8.5. Wait for the weight to stabilize. Add more sample in small amounts at a time, to reach 0.25 ± 0.01 grams of solid.

8.6. Record the weight associated with the sample in grams (g) on the sample label.

8.7. Carefully remove the MOBIO PowerLyzer® Power Soil® DNA isolation kit bead tube containing the sample and cap. Place the sample label on the tube.

8.8. Repeat Steps 8.2-8.7 until all samples are weighed. Make sure spatulas have been washed with 10% bleach and triple rinsed with DI water before starting each new sample.

8.9. Turn the balance off and clean any spilled substances from the balance or surrounding area.

8.10. Store samples at -80°C until analysis.

9. SAMPLE ANALYSIS

9.1. Refer to SOP PCR-1.0 for analysis of samples with LIMS ID: PCR-HF183, PCR-HF183S, PCR-HF183B.

9.2. Refer to SOP PCR-1.2 for analysis of samples with LIMS ID: PCR-GULL2, PCR-GULL2B, PCR-GULL2S.

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10. DATA ARCHIVAL

10.1. Refer to SOP PCR-5.0 for worksheet instructions.

11. QUALITY CONTROL

11.1. Method Blank (MB) – labeled MBLK_#

11.1.1. Analyze a method blank for each batch of samples filtered (maximum of 13 per batch). Filter 50 mL of sterile PBS at the beginning of each sample batch. Extract and analyze with the same batch of samples.

11.2. Replicate Sample

11.2.1. Analyze at least one replicate per filtration batch (maximum of 13 per batch). Filter and analyze replicate in the same manner as original sample is analyzed and within the same batch.

11.2.2. Analyze at least one replicate per weighing batch (maximum of 13 per batch). Purify and analyze replicate in the same manner as original sample is analyzed and within the same batch.

12. SAFETY/HAZARDOUS WASTE MANAGEMENT

12.1. For sample filtration/ weighing, wear safety glasses, gloves, and a clean lab coat.

12.2. When sample filtration is complete, discard remaining sample in the sink, and place sample containers in a trash bag. When trash bag is full, label with the date range of the samples and request a pickup from the Receiving section. After report certification, sample containers are discarded by the Receiving section.

12.3. Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management.

12.4. Read MSDS information for reagent handling and disposal.

13. REFERENCES

13.1. FDEP Bureau of Labs Quality Manual
http://www.dep.state.fl.us/labs/library/progplan.htm

13.2. Health and Safety Plan https://www.fldepnet.org/system/files/Division-of-Environmental-Assessment-and-Restoration/governing-documents/Health%20%20Safety%20Plan_0.docx

13.3. EPA Method B (June 2010)

13.4. 2009 TNI Standard

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13.5. NELAC Quality Systems Manual

13.6. SOP PCR-1.0, *Human-Associated Bacteroidales HF183 Marker in Water by Quantitative Polymerase Chain Reaction (qPCR) Assay*

13.7. SOP PCR-1.2, *Gull-Associated GULL2 Marker by Quantitative Polymerase Chain Reaction (qPCR) Assay*

13.8. SOP PCR-2.2, *Calibration, Use and Maintenance of Mettler Toledo XA105DU Balance*

13.9. DEP SOP PCR-2.5, *Calibration, Use, and Maintenance of Support Equipment*

13.10. SOP PCR-3.0, *Preparation of Reagents for quantitative Polymerase Chain Reaction (qPCR)*

13.11. SOP PCR-5.0, *Sample Custody, Preparation Labels and Worksheet Instructions for Molecular Biology*

13.12. DEP SOP TA-6.18 *AMSCO Renaissance Series 3021 Autoclave Calibration and Use*

13.13. DEP SOP FS 1000, *General Sampling*

Appendix of Changes

06/08/2016 Added LIMS Analysis codes for GULL2 marker and method SOP reference.

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