Measuring Ammonia using HACH® Ammonia Nitrogen Test Kits for Freshwater and Saltwater Samples

1. SCOPE AND APPLICATION

1.1. These methods are adapted from the HACH® Ammonia Nitrogen Test Kit instruction manuals. These methods are used to determine ammonia in both freshwater and saltwater aqueous samples.

1.2. Use of this SOP constitutes acknowledgement that the HACH® Ammonia Nitrogen Test Kit NI-8 (0-3mg/L) and HACH® Ammonia Nitrogen Test Kit NI-SA (0-2.5mg/L) user's manuals have been read and understood by the operator.

2. SUMMARY OF METHOD

2.1. A color disc is placed in the color comparator box.

2.2. Sample is placed in two plastic viewing tubes.

2.3. For freshwater samples: Test kit NI-8 (0-3mg/L)

2.3.1. One tube is placed in the left opening of the color comparator box; Nessler reagent is added to the second tube and swirled to mix. After 1 to 5 minutes, the second tube is placed in the right opening of the color comparator box. The comparator box is held in front of a light source and the color disc is turned until there is a color match. Results are read and recorded.

2.3.2. Interferences include:

2.3.2.1. Contamination when tubes are not thoroughly rinsed after each use

2.3.2.2. Wet disc

2.3.2.2.1. If disc is wet, disassemble and use a soft cloth to dry all parts. Reassemble when fully dry.

2.3.2.3. Temperatures deviating from 20ºC

2.3.2.3.1. Warmer temperatures cause high results while lower temperatures cause low results.

2.3.2.4. More than 6 gpg (~102 mg $CaCO_3$/L) hardness causes a white precipitate to develop

2.3.2.4.1. To remove the interference, add 1 drop Rochelle salt reagent to the tube.

2.4. For saltwater samples: Test Kit NI-SA (0-2.5mg/L)

2.4.1. One tube is placed in the left opening of the color comparator box. One ammonia salicylate powder pillow is added to the other tube and shaken. After 3 minutes, one ammonia cyanurate powder pillow is added to the same tube and shaken. After 15 minutes, tube is placed in the right opening of the color comparator box. The comparator box is held in front of a light source

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and the color disc is turned until there is a color match. Results are read and recorded.

2.4.2. Interferences include:

2.4.2.1. Contamination when tubes are not thoroughly rinsed with DI water after each use, resulting in a second reading lower than the first

2.4.2.2. Wet color disc

2.4.2.2.1. If disc is wet, disassemble and use a soft cloth to dry all parts. Reassemble when fully dry.

2.4.2.3. Measuring brackish water, resulting in readings higher than actual value. With this kit, error should be <10%.

2.4.2.4. Measuring low salinity or freshwater, resulting in readings higher than actual value. With this kit, error can be as high as 16%.

2.5. If results are unusually high or inconsistent, a replicate is necessary.

3. EQUIPMENT AND SUPPLIES

3.1. Disposable pipette (10mL) with manual pipettor

3.2. Laboratory timer (count-down or count-up), with alarm

3.3. For freshwater samples: HACH® Ammonia Nitrogen Test Kit NI-8 (0-3mg/L) containing:

3.3.1. Two 18mm plastic viewing tubes

3.3.2. One color disc

3.3.3. One color comparator box

3.3.4. One bottle of Nessler reagent

3.4. For saltwater samples: HACH® Ammonia Nitrogen Test Kit NI-SA (0-2.5mg/L) containing:

3.4.1. Two 18mm plastic viewing tubes

3.4.2. One color disc

3.4.3. One color comparator box

3.4.4. Ammonia Salicylate Reagent Powder pillows

3.4.5. Ammonia Cyanurate Reagent Powder pillows

3.4.6. Two stopper caps for tubes

4. REAGENTS AND STANDARDS

4.1. Deionized (DI) water, for dilution of high level samples and for thoroughly rinsing viewing tubes

4.2. Nessler reagent, for freshwater samples.

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4.3. Ammonia Salicylate Reagent, for saltwater samples

4.4. Ammonia Cyanurate Reagent Powder pillows, for saltwater samples.

5. METHOD

5.1. Keep samples at room temperature

5.1.1. Warmer temperatures cause high results while colder temperatures cause low results. (See table in NI-SA kit instruction manual).

5.2. Use the disposable pipette with the manual pipettor to add 5mL of sample water into each plastic viewing tube.

5.3. Place one plastic viewing tube into the left opening of the color comparator box, farthest from the color disc.

5.4. For freshwater samples (refer to Step 5.5 for saltwater samples):

5.4.1. Add three drops of Nessler reagent to the tube that is not in the comparator box.

5.4.1.1. Do not shake the Nessler reagent bottle.

5.4.2. Swirl, do not shake, the tube containing Nessler reagent until a yellow color develops.

5.4.3. Wait at least one minute, and no more than five minutes, to read the results

5.4.3.1. Refer to **STEP 5.6** on how to read results.

5.5. For saltwater samples:

5.5.1. Add one ammonia salicylate powder pillow to the tube that is not in the comparator box.

5.5.2. Cap tube and shake until all powder is dissolved.

5.5.3. Wait three minutes.

5.5.4. To the same tube, add one ammonia cyanurate powder pillow.

5.5.5. Cap tube and shake until all powder is dissolved.

5.5.6. Wait 15 minutes before reading results.

5.5.6.1. Refer to **STEP 5.6** on how to read results.

5.5.6.2. If used for brackish or freshwater, the reading will be slightly higher than the actual value.

5.6. Reading results for freshwater and saltwater samples:

5.6.1. Put the second tube (with Nessler reagent or powder pillows) into the open slot of the color comparator box, closest to the color disc.

5.6.2. Hold color comparator box, with both tubes inside, in front of a light source. Compare the front two color viewing windows.

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5.6.2.1. These windows are in-line vertically with the viewing tubes and next to each other. They are located on the left side midway from the top and bottom of the color comparator box.

5.6.3. While looking at the color viewing windows, turn the color disc until the colors in the front windows match. Record the result at the point in which the colors match.

5.6.3.1. If the color match is between two segments, use the value that is in the middle of the two segments.

5.6.3.2. The result can be found in a single window just below and slightly to the right of the color viewing windows mentioned in **STEPS 5.6.2.** and **5.6.2.1**.

5.7. Record the results in the Ammonia logbook and the appropriate bench sheets.

5.7.1. If an out-of-range result is found (>3 mg/L freshwater kit; >2.5 mg/L saltwater kit), dilute a freshwater sample with DI water and a marine sample with ASW of similar salinity. Perform steps for either freshwater (**STEP 5.4**) or saltwater (**STEP 5.5**) as appropriate on diluted sample. Multiply the result by the dilution factor.

5.7.2. Readings less than 0.2 mg/L are recorded as <0.2 for both kits. Readings >0.2 mg/L are recorded as displayed on the results window.

5.7.3. If saltwater results are unusually high or inconsistent, a replicate is necessary. For the saltwater HACH® Ammonia Nitrogen Test Kit NI-SA, a lower result on the second test indicates contamination of the first test. Troubleshoot and re-run the sample.

5.8. Rinse viewing tubes thoroughly with DI water then repeat **STEPS 5.4/5.5** and **STEP 5.6** for the replicate sample, if needed.

5.9. When finished with the kits, rinse the viewing tubes thoroughly with DI water and allow to dry before placing in kit. If Nessler reagent was used, ensure the lid is secure. Return dry viewing tubes, color comparator boxes, Nessler reagent and any unused pillow packets to appropriate HACH® ammonia kit. Clean up the work area.

6. DATA ARCHIVAL

6.1. Use a black, waterproof pen for all bench sheets and logbooks.

6.2. Record information in *Ammonia* logbook, *Dilution Water Parameter and Instrument Maintenance* logbook and associated *Bioassay Bench Sheets*.

6.3. To correct raw data entries, place a single line through the incorrect entry, write the corrected entry near the error and initial and date the correction. Do not overwrite entries.

6.3.1. The corrected entry is written as near its appropriate space as possible or else in the comment box on the appropriate bench sheet.

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6.3.2. If the correction requires an explanation, or there is very little space for writing, write a comment and reference the error. Initial and date the explanation.

6.3.3. Use sequential alphabetical annotation to identify errors and corresponding comments (use A for first error and use A for corresponding explanatory comment; use B for second error, etc.).

7. QUALITY CONTROL

7.1. Refer to QA manual for corrective action and handling out-of-control data.

8. SAFETY/HAZARDOUS WASTE MANAGEMENT

8.1. Proper laboratory protection, including lab hood or ventilation system, lab coat, gloves and safety glasses, is required when working with chemicals and unprocessed samples.

8.2. Refer to the Laboratory Quality Manual for sample disposal.

8.3. Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management.

9. REFERENCES

9.1. HACH® Ammonia Nitrogen Test Kit NI-8 (0-3mg/L mid-range) Instruction Manual.

9.2. HACH® Ammonia Nitrogen Test Kit NI-SA (0-2.5mg/L) Instruction Manual

9.3. Laboratory Quality Manual, http://www.floridadep.org/labs/librabry/progplan.htm

9.4. Laboratory Health and Safety Manual and Hazardous Waste Management, https://www.fldepnet.org/content/dear/bureau-laboratories-safety-documents

9.5. DEP SOP MB-03.03, *Calibration and Use of Mechanical and Disposable Pippettes.*

APPENDIX OF CHANGES:

12/05/2016 SOP written using TA-6.22-2.17 *Measuring Total Residual Chlorine using the Hach® Pocket Colorimeter™ and Colorimeter™ II Test Kits* as a template. Also, formatted for 508 compliance.

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