Pour Plate Method for Heterotrophic Plate Count

1. SCOPE AND APPLICATION

1.1. The heterotrophic plate count estimates the number of viable heterotrophic bacteria in water. The pour plate method can be used for 0.1-2.0 mL of sample. This method is used in the laboratory to conduct several quality control tests including, water suitability test, use test, monthly water quality tests and precision and recovery tests for analytical methods.

2. SUMMARY OF METHOD

2.1. Small aliquots of sample are pipetted into duplicate Petri dishes. 10-12 mL of Heterotrophic Plate Count Agar (HPC) is added to each plate. The plates are carefully swirled to mix the sample with the agar. After the media solidifies at room temperature, the plates are inverted and incubated for 48 ± 3 hours. The ideal colony count should be between 30 and 300 colonies for at least one dilution of sample. The reporting limit is <1000 CFU/mL.

2.2. Interferences

2.2.1. HPC is not selective and can support the growth of many types of bacteria. Aseptic techniques must be used to ensure that there is no contamination of the sample via the media, reagents, supplies, or the analyst.

2.2.2. HPC should be tempered to 44-46˚C after melting. If it is too warm when mixed with the sample, bacteria in the sample could undergo heat shock and could potentially die. This would cause the results to be less reliable.

3. EQUIPMENT AND SUPPLIES

3.1. Petri dishes, sterile, 100 mm x 15 mm, 4-8/sample

3.2. Volumetric pipets, 1 mL and 10 mL – each new lot is checked per SOP MB-03.3

3.3. Hot plate

3.4. 1000 mL beaker

3.5. Heratherm IMH 180 Model #51028 incubator (S/N 41784672), maintained at 35 ± 0.5 °C

4. REAGENTS AND STANDARDS

4.1. Heterotrophic Plate Count Agar (HPC), 1-2 bottles per sample (prepared per SOP MB-02.33)

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4.2. Dilu-Lok 99 mL sterile buffered water bottle, 2 per sample

5. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

5.1. Samples must be stored at <4˚C until analyzed. See SOP FS 1000 and Bureau of Labs Quality Manual for general sample collection and preservation.

5.2. Holding time

5.2.1. Potable: within 30 hours of collection

5.2.2. Non-Potable: received and filtered within 8 hours of collection

6. SAMPLE PREPARATION

6.1. See SOP BB-030 for sample custody and label instructions.

6.1.1. Label duplicate Petri dishes (two plates per sample dilution) with bar code label and dilution of sample.

6.1.2. Sample dilutions should be based on expected pollution levels. The ideal range should be 30-300 colonies for at least one dilution. For samples where no growth is expected, analyze as a potable water sample.

6.1.2.1. Potable water sample: use 1 mL and 0.1 mL dilutions

6.1.2.2. Non-Potable water sample: use 0.01 mL and 0.001 mL dilutions

6.1.2.3. Polluted Non-Potable water sample: use 0.0001 mL dilution

6.2. Melt HPC

6.2.1. Place 2 bottles of media in a 1000 mL beaker and fill with water (approximately up to the necks of the bottles). Each sample requires 1-2 bottles of media, so more than one beaker may be needed.

6.2.2. Place beaker(s) on a hot plate(s) and bring water to a boil until agar is completely melted.

6.2.3. Remove bottles from the beakers and allow to cool to 44.0-46.0˚C before pouring. Media should still be liquid, but cool enough to handle. Do not prepare any more samples than can be diluted and plated within 20 minutes.

****Note:** Prepared agar should be melted only once. Any unused melted agar should be discarded.

7. SAMPLE ANALYSIS

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7.1. Vigorously shake the Whirl-Pak or bottle containing the sample approximately 25 times.

7.2. Potable water samples

7.2.1. Use sterile pipettes to transfer 1 mL and 0.1 mL of undiluted (100%) sample into duplicate Petri dishes (1 pipette per plate, 4 plates total).

7.2.2. Add 10-12 mL of the melted HPC to each dish

7.2.3. Mix the inoculated medium carefully to prevent spilling. Swirl the plate counterclockwise 5 times, clockwise 5 times, and back and forth 5 times.

7.2.4. Allow medium to solidify at room temperature (about 10 minutes).

7.2.5. Invert plates and incubate in Heratherm Incubator #51028 at 35.0±0.5˚C for 48±3 hours.

7.2.6. After incubation, select the plates with 30-300 colonies. Use a colony counter to count all the colonies on each of those plates. For each plate, record the count on the lid with a permanent marker.

7.3. Non-Potable water samples

7.3.1. Prepare a 1:99 dilution of the original 100% sample.

7.3.1.1. Use a sterile pipette to transfer 1 mL of the original sample into a Dilu-Lok 99 mL sterile buffered dilution water bottle.

7.3.1.2. Mix thoroughly by shaking vigorously approximately 25 times.

7.3.2. Use sterile pipettes to plate 1 mL and 0.1 mL of the 1:99 sample dilution into duplicate Petri dishes (1 pipette per plate, 4 plates total). The new dilutions will be 0.01 mL and 0.001 mL, respectively.

7.3.3. Continue with steps 7.2.2.-7.2.6.

7.4. Polluted Non-Potable water samples

7.4.1. Further dilute the original sample by using a sterile pipette to transfer 1 mL of the 1:99 sample dilution into another Dilu-Lok 99 mL sterile buffered dilution water bottle and mix thoroughly by vigorous shaking approximately 25 times.

7.4.2. Use sterile pipettes to plate 1 mL of the second 1:99 sample dilution into duplicate Petri dishes (1 pipette per plate, 2 plates total). The new dilution will be 0.0001 mL.

7.4.3. Continue with steps 7.2.2.-7.2.6.

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8. DATA ARCHIVAL

8.1. Record counts and sample data in appropriate columns in the **Micro_Data_Log**, found in **LIMS/Laboratory/Technical/Laboratory Logbook**.

8.2. Print a Micro Data Log Report. Go to **LIMS/Laboratory/Technical/Laboratory Logbook**. Click on **Reports** in the top left corner of the window. Choose the appropriate report and print.

8.3. Verify the colony counts and dilutions. Refer to SOP BG-01.

8.3.1. After verification, place dishes in the hazardous waste bin.

8.4. Upload according to SOP BB-31.

9. QUALITY CONTROL

9.1. For each set of samples, into sterile Petri dishes (1 per bottle of media used), pour enough HPC to cover the bottoms of the dishes—only 1 bottle of media per dish. Incubate with the sample dishes.

9.1.1. Growth indicates contamination.

9.1.2. No growth indicates no contamination.

9.2. Refer to QA manual for corrective action and handling out-of-control data.

10. SAFETY/HAZARDOUS WASTE MANAGEMENT

10.1. Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management.

10.2. Read MSDS information for reagent handling and disposal.

11. REFERENCES

11.1. Standard Methods Online Edition, Section 9251A-00 and 9215B-00

11.2. FDEP Laboratory Quality Manual http://www.dep.state.fl.us/labs/library/progplan.htm

11.3. Laboratory Health and Safety Manual https://www.fldepnet.org/content/dear/bureau-laboratories-safety-documents

11.4. DEP SOP MB-03.3 *Calibration and Use of Mechanical and Disposable Pipettes*

11.5. DEP SOP FS-1000 *General Sampling Procedures*

11.6. DEP SOP MB-02.33 *Preparation of Dehydrated Culture Media*

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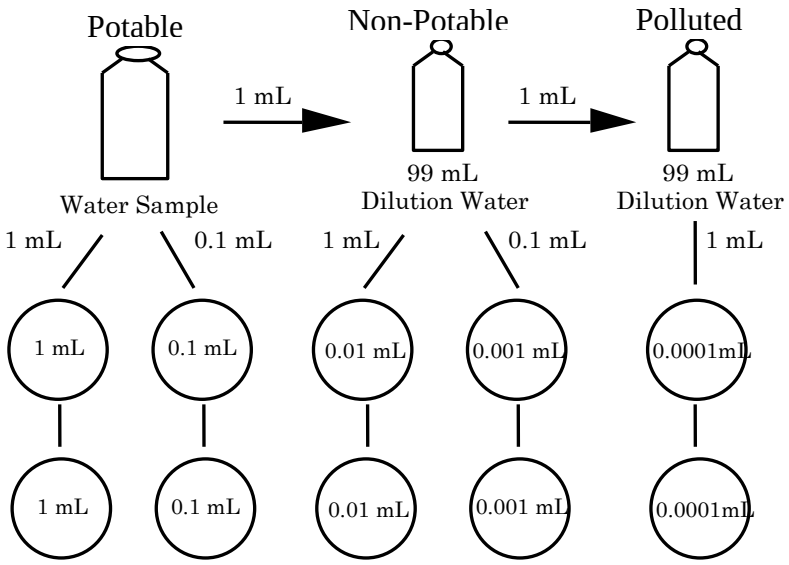
11.6. DEP SOP BB-030 *Sample Custody, Preparation Labels and Worksheet Instructions for Bench Biology Samples*

11.7. DEP SOP BG-01 *Data Entry Verification*

11.8. DEP SOP BB-031 *LIMS QC Manager Data Uploading Instructions for Bench Biology and Microbiology Samples*

12. TABLES, DIAGRAMS

Figure 12.1 Dilution Preparations



Appendix of Changes

08/11/2008	Updated to SM 9215B Online Edition and added a references section.
08/12/2010	Added Introduction and updated reference to new version of safety plan.
08/08/2011	Corrected reference to quality manual, added specific incubator used in materials section.
07/20/2012	Updated format; added DEP SOP FS-1000, DEP SOP BB-030, and DEP SOP BB-032 to References section.
07/16/2013	Changed references to PCA to HPC.
11/16/2015	Remove reference to the old water bath and updated reference to Health and Safety Manual.
09.15.2017	Updated incubator information.

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