Measuring Total Residual Chlorine using the Hach® Pocket Colorimeter™ and Colorimeter™ II Test Kits

1. SCOPE AND APPLICATION

1.1. This method is adapted from the HACH® Pocket Colorimeter™ and HACH® Colorimeter™ II Test Kits instruction manuals. As adapted, it is equivalent to SM 4500-Cl G. The method is used for determining total residual chlorine (TRC) in aqueous samples.

1.2. Use of this SOP constitutes acknowledgement that the Hach Pocket Colorimeter™ and the Pocket Colorimeter ™ II user's manuals have been read and understood by the operator.

1.3. **WARNING** This laboratory is not NELAC certified for this analysis. See Method **STEP 5.1** on page 2 of this SOP.

2. SUMMARY OF METHOD

2.1. Operation of meter is checked. The sample is placed in a vial. Instrument is "blanked" with the sample. DPD (diethyl-p-phenylenediamine) reagent is added to the vial. The timer is started. TRC is read. A duplicate measurement is made. Results are averaged and recorded in appropriate log books.

2.2. Interferences:

2.2.1. Include: acidity > 150 mg/L $CaCO_3$, alkalinity > 250 mg/L as $CaCO_3$, bromine, hardness > 1,000 mg/l as $CaCO_3$, iodine, oxidized manganese and oxidized chromium, monochloramine and ozone.

2.2.2. Damage to the meter by water/liquid entering the cell holder.

3. EQUIPMENT AND SUPPLIES

3.1. Hach pocket colorimeter™ or colorimeter™ II chlorine test kits (S/N 05110C042029, S/N 05050C028378 or S/N15070E276527) containing:

3.1.1. Pocket colorimeter™ or colorimeter™ II

3.1.1.1. The meters have two modes. The LOW mode resolution is 0.01 mg/L. The HIGH mode resolution is 0.1 mg/L.

3.1.2. Two 2.5 cm sample cells

3.1.3. Two 1.0 cm sample cells

3.1.4. 1 cm sample-cell adapter

3.1.5. DPD total chlorine reagent powder pillows

3.2. Disposable pipet (10 mL) with manual pipetter

3.2.1. One per sample to be tested (volume of each new lot is checked per SOP MB-03.3)

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3.4. Laboratory timer (count-down or count-up with alarm)

3.5 Kimwipe®

4. REAGENTS AND STANDARDS

4.1. Disposable pipet (10 mL) with manual pipette

4.2. Chlorine demand free water

4.2.1. Deionized (DI) water for dilution of higher level samples

4.3. Hach-Spec√™ Secondary Standards and if needed potassium permanganate standards (1.0 mg/L and 0.1 mg/L)

4.3.1. The Hach® secondary standards come four to a box and include a blank, a low-range standard, a mid-range standard, and high-range standard. The potassium permanganate standard (1.0 mg/L) is prepared per Standard Methods 20th Edition: SM 4500-Cl G, procedure 4-2 page 4-64. Dilute the 1.0 mg/L standard 9:1 with deionized water to make the 0.1 mg/L standard. Record the preparation in the *Tox Record of Preparation Logbook* as batch # (batch # = date prepared: ex. 1/05/14), date, expiration (date prepared + 7 days). Make fresh potassium permanganate standards whenever the standard(s) are > ± 10% from nominal even if "unexpired." (This presumes that the secondary standards are within specified ranges and there is not a problem with the meter).

5. METHOD

5.1. **Note: Results** of chlorine measurements performed on samples in the Bioassay laboratory **are not reportable for compliance purposes.** The results are used to determine potential causes of observed toxicity and as a measure of the success of dechlorination efforts.

5.2. The following methods apply to both the Colorimeter™ and the Colorimeter™ II when referring to the instrument.

5.3. Use Spec√™ Secondary Standards and if needed Potassium Permanganate Standards for calibration. Total Residual Chlorine PT samples can be used in lieu of Potassium Permanganate. Ensure that the standards have not expired.

5.4. *Sample preparation*

5.4.1. Warm sample to between 20-25 ºC.

5.5. Meter Calibration and Verification of Instrument

5.5.1. Press the power button on the Colorimeter™II. The display will light up.

5.5.2. Remove the cap from the instrument. Wipe the Spec√™ blank with a clean Kimwipe® and place into the cell holder. Ensure that the alignment mark (looks like half of a diamond) is facing the keypad. Securely cover the cell with the instrument cover.

5.5.3. Press [**ZERO**]. The display will show "**0.00**".

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5.5.4. Remove the instrument cap. Remove the Spec√™ blank from the cell holder. Place the STD 1 cell into the cell holder after wiping it with a clean Kimwipe®, aligning the alignment mark with the keypad. Securely cover the cell with the instrument cover.

5.5.5. Press [**READ**]. Record the displayed value in the chlorine logbook along with the value of the standard.

5.5.5.1. If the value is not within ±10 % of the standard value, ensure that the standard cells are clean and repeat the procedure. If values are still out of range, refer to the instrument manual for trouble shooting instructions.

5.5.6. Repeat **STEP 5.5.4.** and **STEP 5.5.5**, first with the Spec√™ cell labeled STD 2 and then with the Spec√™ cell labeled STD 3.

5.5.7. Refer to all steps of the *Total Residual Chlorine Measurement Low Range, 0 – 2.00 mg/L* procedure using the potassium permanganate standards as the sample. Record the displayed value in the chlorine logbook.

5.5.7.1. In lieu of potassium permanganate standards that can be measured once annually or whenever there appears to be a problem with the secondary standards; semi-annual Total Residual Chlorine proficiency samples can be run. Potassium permanganate standards, if made and used, should read 1.0 mg/L (± 10%) and 0.1mg/L (± 20%). If the values are not within the specified percentage of the standard value, ensure that the standard cells are clean and repeat the procedure. If values are still out of range, prepare a new standard. Test the new standard, and then if a problem still exists, refer to the instrument manual for trouble shooting instructions.

5.6. Total Residual Chlorine Measurement Low Range, 0 – 2.00 mg/L

5.6.1. Use a 10-mL pipet to place 10 mL of sample into a 10-mL sample cell (blank sample).

5.6.1.1. Use one pipet per sample (do not cross contaminate samples).

5.6.2. Wipe the exterior of the cell with a Kimwipe® to remove any dirt, smudges, fingerprints, etc. which may reduce the amount of light passing through the cell, thereby giving an inaccurate reading.

5.6.3. Place the sample cell containing the blank sample in the cell holder. Place the instrument cap over the sample cell (to occlude light).

5.6.3.1. The diamond mark on the sample cell (vial) should be facing the front of the instrument (toward the operator).

5.6.4. Press [**ZERO**]. After the display “zeros,” remove the instrument cap from the cell holder and remove the sample cell.

5.6.4.1. This step “zeros” the meter while correcting for any color inherent in the sample. The colorimeter will turn on and the display will show "**---**" followed by "**0.00**."

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5.6.5. Open a DPD Total Chlorine powder pillow by tearing the top portion off and add the contents to the sample cell (blank sample). Cap the cell and gently shake for 20 seconds to mix.

5.6.5.1. A pink color will form if chlorine is present.

5.6.6. To allow proper color development, wait at least three minutes but not more than six minutes before performing **STEP 5.6.8**.

5.6.6.1. Set the timer for 3 minutes. (If used, else monitor the wall clock).

5.6.6.2. Any undissolved powder does not affect accuracy.

5.6.7. Check the sample cell for bubbles that sometimes form on the sides of the cell. These bubbles will cause a false high result. If bubbles are seen, gently tap the side of the cell and swirl the contents of the cell to dislodge the bubbles.

5.6.8. After three minutes, place the prepared sample in the cell holder. Securely (to occlude light) place the instrument cap over the sample cell (flat part of cap away from operator).

5.6.9. Press [**READ**]. The display will show "**---**" followed by the total chlorine measurement in mg/L.

5.6.9.1. If the sample temporarily turned yellow at **STEP 5.6.5** or the display is flashing "**2.20**", (indicating an over-range concentration), the chlorine concentration is too high to measure without dilution. The measurement must be repeated with a diluted sample.

5.6.10. Record the displayed reading in the Chlorine logbook and the appropriate bench sheets. If an out-of-range indication is displayed, dilute the sample with DI water. Perform **STEPS 5.6.1-5.6.10** to measure TRC in both the dilution water (DI water) and diluted sample. Multiply the result by the dilution factor.

5.6.10.1. Readings that are ≤0.03 ppm are recorded as <0.03. Readings >0.03 are recorded as displayed on the meter.

5.6.10.2. Dilution water should be chlorine demand free water. If TRC is detected in the dilution water, contact the lab supervisor immediately.

5.6.11. Repeat **STEPS 5.6.1-5.6.10** for the replicate sample. If sample readings are ≤0.03 ppm, two readings are sufficient. If any sample readings are ≥0.03 and vary by more than 0.02 ppm, or whenever one reading is ≤0.03 and the other is > 0.03, a third reading must be taken.

5.6.12. When finished with the instrument, replace the cap, turn off the Colorimeter™ II by pressing the power button, and return the instrument to the carrying case. Rinse all sample cells (excluding Spec√™ Standards) with DI water. Allow the cells to dry before returning them to the carrying case. Clean up the work area.

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6. DATA ARCHIVAL

6.1. Use a black, waterproof pen for all bench sheets and logbooks.

6.2. Record information in *Chlorine Bench Sheets* and *Bioassay Bench Sheets* logbooks.

6.3. To correct raw data entries, place a single line through the incorrect entry, write the corrected entry near the error and initial and date the correction. Do not overwrite entries.

6.3.1. The corrected entry is written as near its appropriate space as possible or else in the comment box on the appropriate bench sheet.

6.3.2. If the correction requires an explanation, write a comment and reference the error. Initial and date the explanation.

6.3.3. Use sequential alphabetical annotation to identify errors and corresponding comments (use A for first error and use A for corresponding explanatory comment; use B for second error, etc.).

7. QUALITY CONTROL

7.1. Refer to QA manual for corrective action and handling out-of-control data.

8. SAFETY/HAZARDOUS WASTE MANAGEMENT

8.1. Proper laboratory protection, including lab hood or ventilation system, lab coat, gloves and safety glasses, is required when working with chemicals and unprocessed samples.

8.2. Refer to the Laboratory Quality Manual for sample disposal

8.3 Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management

9. REFERENCES

9.1. HACH® Pocket Colorimeter™ Instruction Manual.

9.2. Standards Methods for the Examination of Water and Wastewater, 20th Edition: SM 4500-Cl G.

9.3. Laboratory Quality Manual, http://www.floridadep.org/labs/library/progplan.htm

9.4. Laboratory Health and Safety Manual and Hazardous Waste Management, https://www.fldepnet.org/content/dear/bureau-laboratories-safety-documents

9.5. DEP SOP MB-03.03, *Calibration and Use of Mechanical and Disposable Pipettes*.

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APPENDIX OF CHANGES

1/04/2008	Moved *Scope and Application* and *Summary of Method* from comments to inserted rows. Moved subtitle to *Scope and Application*. Added *Appendix of Changes*.
12/30/2008	Added Interferences and References Sections
1/05/2011	Updated reference to Biology Quality Manual. Defined batch # and changed expiration information in comment for Materials Step 3. Changed comment for Step 6 *Repeatability check* to read quarterly rather than annually.
11/07/2013	Reformatted SOP.
02/09/2015	Formatted for 508 compliance.
09/24/2015	Updated links and references.
10/07/2016	Updated links and references.
09/28/2017	Updated links and references

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