NUTRIENTS: QUALITY ASSURANCE AND QUALITY CONTROL

1. SCOPE AND APPLICATION

1.1. The purpose of this Standard Operating Procedure (SOP) is to establish, maintain and document general quality control (QC) for the Nutrients workgroup. The controls used are bias, precision, and detection limit statistics.

1.2. These procedures and calculations are used for all Nutrients analyses: sediments, surface, ground and wastewaters. The procedures described in this SOP are general; more specific or stringent requirements are documented in the method and SOP for the relevant test.

2. QUALITY CONTROL

2.1. In general, for every batch of samples (max. 20 samples) digested and/or analyzed, the following quality control samples are analyzed:

A Laboratory Fortified Blank (LFB)

Digestion or Method Blank (MBLK)

One Matrix Spike

Matrix Spike Duplicate or Sample Duplicate

2.2. Accuracy.

2.2.1. Standardization:

2.2.1.1. Standards are prepared and documented in the Standard Preparation Tracker module of the Laboratory Information Management System (LIMS) program. Guidelines for the preparation of standards are documented in DEP SOP NU-042. Calibration curves are generated at the beginning of each analytical run or at the frequency specified in the method SOP. The correlation coefficient for a standard curve should be 0.995 or greater and the recovery for each calibrant above the practical quantitation limit (PQL) should be 90 - 110 %. These recoveries verify the working calibration range with every calibration. The curves are also checked against a certified or other independently prepared (from a stock of a different lot or manufacturer) standard during each analytical run. The continuing validity of the calibration is verified throughout the run by analyzing check standards (may also be called “continuous calibration verification” standards) at least every 20 samples. Along with check standards, blanks are analyzed to monitor for contamination, carry over, or baseline problems.

2.2.1.2 All analytical equipment must be calibrated on a regular basis (e.g. calibrate pipettes on a monthly basis). Guidelines for the

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calibration of auto-pipettes and dispensers are given in DEP SOP CM-032. The calibrations must be documented. All appliances used for storage or analysis must be monitored regularly and documentation must be maintained. See the supervisor for instructions on equipment calibration and monitoring. All glassware used to measure volumes for analytical purposes should be class A.

2.2.2. Spike Recoveries:

2.2.2.1. A representative sample is spiked with known quantities (preferably approximately 2 to 10 times the PQL) of the analyte before processing. Percent recoveries are calculated for the added analyte. Matrix spike recoveries are indicators of sample matrix interference and contamination. The confidence range is set at 85 - 115 % for water matrices and 80 - 120 % recovery for soil matrices.

2.2.2.2. A blank is spiked with known quantities of the analyte (usually at the same concentration as a matrix spike) before processing. Percent recoveries are calculated for the added analyte. LFB recoveries are indicators of method failure and contamination. The confidence range is set at 85 - 115 % for water matrices and 80 - 120 % recovery for soil matrices.

2.2.3. Blanks:

2.2.3.1. Blank results are used to control contamination and to set minimum detection limits (MDL). In general, there are two types of blanks associated with an analytical run: method blanks (MBLK) and check blanks (CCB).

2.2.3.2. Method blanks are a part of the batch QC and processed together with the samples through all preparation steps. They are used to monitor the preparation process for possible contamination. One MBLK is prepared for each batch. MBLKs are also used for ongoing method detection limit (MDL) verification. MDLs are set such that the risk of reporting a false positive is less than 1% (Please see ref. 6.6. and 6.7 for MDL verification procedures).

2.2.3.3. A check blank is analyzed at the beginning, at least every 20 samples, and at the end of each run. Use deionized water for blanks and adjust the water to match the sample matrix (i.e. acidify, basify or add salt). Check blanks are used to monitor contamination and carry over that may occur on the instrument during the analytical run.

2.2.4. Practical Quantitation Limit (PQL):

2.2.4.1. The PQL is the concentration above which the analyte can be quantified with the method-required accuracy and precision. Above PQL results are reported unqualified (if all QC measures

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meet acceptance criteria). Method control limits are not applicable to results below PQL; these results are flagged with an "I" qualifier. The PQL is determined during the method validation and verified yearly according to the procedures delineated in Ref. 6.6 and 6.7.

2.2.4.2. PQL level spikes are used for two purposes: a) for ongoing MDL and PQL verification and b) for evaluating instrument performance at the low concentration range. Digested PQL standards are required with each tray of digested samples. For un-digested chemistries, at least one PQL level standard is analyzed with every analytical run. The acceptance range for a PQL is 70 - 130 % recovery. For analytical runs that continue for 24 hours or more, at least one PQL standard should be analyzed every 24 hours.

2.2.4.3. Ongoing MDL verification requires analysis of a spike at the same concentration level as was used during the initial method validation study. Typically, this concentration is the same as PQL. In these cases, the PQL standard is utilized for ongoing MDL verification. There may be cases where PQL and MDL verification spike are at different concentration levels. In such cases the MDL verification spike may be used instead of a PQL level spike with the same acceptance limits.

2.2.5. Other bias assessments:

2.2.5.1. To evaluate any bias possibly introduced during sample preparation, batch QC samples should undergo the same processing steps as samples.

2.2.5.2. If some of the samples are filtered prior to analysis, positive and negative bias checks are required. This can be achieved by filtering the MBLK and an LFB or a check standard. In such cases, samples and QC samples filtered are identified by adding a suffix "_F" to their name.

2.3. Precision.

2.3.1. Any time an instrument takes a replicate reading on a sample or if the sample is analyzed more than one time, the precision needs to be checked.

2.3.2. The relative percent difference (RPD) for two or more data points is calculated either by the instrument or the analyst. When the average value of the concentration is above the PQL then the RPD must be less than 20% in order to be acceptable.

3. QUALITY ASSURANCE

3.1. All data and quality control data are reviewed by a supervisor before being accepted as final. See DEP SOP NU-043.

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3.2. All maintenance or analytical problems are reported to a supervisor. Document all instrument maintenance in the logbook assigned to the instrument. Also, document equipment calibrations (pipettes, balances etc.) in the assigned notebooks or computer files. See the supervisor for instructions on how to document the instrument maintenance and equipment calibration.

3.3. Field sampling problems such as contamination, labeling, and holding times are reported to a supervisor and noted with the results using the NCR module of the LIMS program.

3.4. Results that are suspect for any reason are qualified and/or noted in the 'Ref. Method and Comment' line of the final report.

3.5. For methods that do not have electronic documentation of the time of analysis the start and end time of the analytical run should be documented by the analyst on the raw data. The end time of the run should be used as the time of analysis in the QC Manager. If samples are close to expiration the individual time of analysis should be noted and used for those samples.

3.6. Quality control results are documented electronically through the QC Manager module of the LIMS program.

4. CORRECTIVE ACTIONS

Contact a supervisor and report any non-compliant QC results. There are many different reasons for QC failures. In general, for any failure associated with the initial calibration (failure of independent standard, correlation coefficient out-of-compliance etc.) the run should be aborted, the cause of the failure evaluated and resolved, and the instrument recalibrated. For further information see DEP SOP NU-043.

5. REMARK CODES (SEE ALSO DEP SOP LB-027)

5.1. Sample results that are below the PQLs and/or MDLs have to be reported with a remark code (qualifier). Samples that are prepared and analyzed in duplicate are reported as an average and flagged with a qualifier. Samples that are analyzed after expiration must be reported with a qualifier. Samples associated with quality control failures may be qualified. Incorrectly preserved samples must be qualified. The QC Manager module of the LIMS program will assign most of these codes automatically (i.e. I, U, A). The Q qualifier and J qualifier for failed spike recoveries, RPDs, LFBs or CCVs are assigned automatically in the Data Review/Authorize module of the LIMS program. Appropriate comments for Q qualified data and non-automatically assigned J qualified data should be added in the QC Manager. There are default comments for the most common reasons for qualifying data.

5.1.1. Sample results that are below the MDL are reported as the MDL followed by the U qualifier.

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5.1.2. Sample results that are below the PQL but at or above the MDL are reported with an I qualifier.

5.1.3. For samples that are poured out and analyzed in duplicate, QC manager will calculate the average and append the A qualifier.

5.1.4. Samples that are analyzed after expiration are reported with the Q qualifier.

5.1.5. For the QC sample in batches associated with failing QC, the result is reported with a J qualifier. All sample results in a batch with associated LFB failure are reported with a J qualifier. All sample results where the bracketing continuing calibration verification standard fails are reported with a J qualifier.

5.1.6. For samples that are incorrectly preserved, the result is reported with a Y qualifier and an appropriate comment added in the QC Manager. Make sure the preservation flag in LIMS is adjusted and an appropriate NCR record created if the preservation issue was not detected at login but during analysis. Check with your supervisor before authorizing associated results.

5.2. Some conditions may require two or more qualifiers, use the list below to assign the sequence of qualifiers (a is the first). Multiple codes and comments may be assigned.

a U or I

b J

c Y

d A

6. REFERENCES

6.1. Quality Manual, Bureau of Laboratories, FDEP.

6.2. DEP SOP NU-043, Nutrients and General Chemistry Quality Assurance and Data Authorization Guidelines.

6.3. DEP SOP NU-042, Standard Preparation Procedure.

6.4. DEP SOP LB-027, Standard Operating Procedure for Reporting Qualified Data.

6.5. DEP SOP CM-032, Calibration of Auto-Pipettes and Dispensers using the Check & Track software.

6.6. 2016 TNI Standard, EL-V1-2016-Rev.2.1.

6.7. Definition and Procedure for the Determination of the Method Detection Limit, Revision 2.0, 40 CFR Part 136, Appendix B.

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Appendix of Significant Changes

12/10/2018 Section 2 was modified for clarity and to include new MDL and PQL verification requirements. References 6.6 and 6.7 were added.

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