Standard Operating Procedure for:

Human-Associated *Bacteroidales* HF183 Marker in Water by Quantitative Polymerase Chain Reaction (qPCR) Assay

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BIOLOGY PROGRAM
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1. SCOPE AND APPLICATION

1.1. This method describes a quantitative polymerase chain reaction (qPCR) procedure that targets a sequence in the 16S rRNA gene found in multiple *Bacteroides* species which are associated with human feces.

1.2. This procedure is used for samples with LIMS test ID of PCR-HF183, PCR-HF183B, PCR-HF183S, and PUR-HF183.

2. SUMMARY OF METHOD

2.1. Bacteria present in a water sample are collected by filtration. The filtered sample is processed by a bead beater in the presence of zirconium silica beads to extract the DNA from the bacterial cells into a buffer solution. The clarified extract may also be purified using a purification kit. The extract is analyzed with qPCR amplification and detection in a thermocycler system using a primer/probe assay specific to human associated *Bacteroidales*. Solid samples are weighed and purified prior to PCR amplification and detection.

2.2. Interferences

2.2.1. Colloidal or suspended particulate materials in water can clog the filter and prevent filtration of enough sample needed to detect the target sequence. Some substances, including humic and tannic acids, can act as inhibitors and reduce efficiency by either binding to the DNA polymerase or interfering with the annealing and extension cycles. Sample purification or dilution can help alleviate the effects of inhibitors.

2.3. Definitions

2.3.1. qPCR: Quantitative Polymerase Chain Reaction is a procedure in which the amount of DNA amplified in each PCR cycle is measured using fluorescent dyes to determine the original amount of DNA in the sample.

2.3.2. Target sequence: The DNA sequence in the Gene of Interest (GOI) that is amplified by the primers/probe in each PCR cycle. The primers/probe set is designed to attach specifically to this sequence.

2.3.3. *Bacteroides*: A genus of anaerobic, Gram-negative rod bacteria that are typically found in the gastrointestinal tract of humans and animals. *Bacteroides dorei* (*B. dorei*) is used as the reference standard for this method.

2.3.4. Standard Curve: A dilution series of standards with known DNA concentrations is analyzed and plotted on a graph with the log of the known concentrations on the x-axis and the quantification cycle (C_q) value of each dilution on the y-axis to create a standard curve. The standard curve is used to determine the

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concentrations of unknown samples and the performance of the reaction using several parameters including the slope and correlation of the graph.

3. EQUIPMENT AND SUPPLIES

3.1. Gloves, latex or nitriles

3.2. Tube racks

3.3. Dispersion beads, 0.1 mm, zirconium silica

3.4. Extraction tubes, conical, 2 mL screw cap with o-ring seal

3.4.1. Weigh out 0.3 ± 0.01 g of dispersion beads into each extraction tube. Seal tube tightly, making sure the O-ring is seated properly. Autoclave @ 121°C for 15 min (15 psi). Record in LIMS Autoclave Log.

3.5. Centrifuge tubes, 0.5, 1.5, and 2.0 mL

3.6. Micropipettes with sterile PCR grade filter tips

3.7. 96-well Reaction Plate, Clear

3.8. 96-well plate holders and stands

3.9. Optical adhesive Seal for reaction plates

3.10. Autoclave (S/N 031761851)

3.11. Mettler Precision Balance Model XA105DU (S/N B314195659)

3.12. CBS Optimizer PCR Workstation #1 (S/N 007631)

3.13. CBS Optimizer PCR Workstation #2 (S/N 007632)

3.14. Fisher Mini Vortexers

3.15. Fisher Minicentrifuges

3.16. Thermo Sorval T3 Centrifuge (S/N 309020018)

3.17. QuantStudio3 Thermocycler (S/N 272311740)

3.18. PCR3 – Step One Plus Thermocycler (S/N 2720010389)

3.19. PCR#1, Thermo Refrigerator (S/N 0121179101121219), maintained at 4ºC ± 2ºC, Room B246A

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3.20. PCR#2, Thermo Freezer (S/N 0121174101121218), maintained at -20ºC ± 10ºC, Room B246A

3.21. PCR#4, Thermo Freezer (S/N 0121174201121218), maintained at -20ºC ± 10ºC, Room B246B

3.22. PCR#5, Thermo Fridge/Freezer (S/N 0121174701121218), maintained at 4ºC ± 2ºC and -20ºC ± 10ºC, Room B246

3.23. PCR#6, Thermo UXF Freezer (S/N 0132376801130826), maintained at -80ºC ± 10ºC, Room B246

3.24. PCR#7, Thermo Ultra Low Freezer (S/N 128473401090813), maintained at -80ºC ± 10ºC, Room B246

4. REAGENTS AND STANDARDS

4.1. Deionized Water

4.2. Molecular Biology grade water – Corning Molecular Biology Grade Water tested to USP Sterile water (Cat.# MT46000CV)

4.3. Bleach solution, 10% (v/v)

4.4. Bovine Serum Albumin (BSA), 2mg/mL – Prepare per SOP PCR-3.0.

4.5. PerfeCta® qPCR ToughMix™, ROX™ to use with StepOne Plus Thermocycler

4.6. TaqMan™ Environmental Master Mix 2.0 to use with QuantStudio3 Thermocycler

4.7. Prepare the following per SOP PCR-3.1.

4.7.1. Salmon DNA Standards (10^3, 10^2, 10^1)

4.7.2. Salmon DNA Extraction Buffer

4.7.3. Salmon DNA Forward and Reverse Primers, 500 µM Stock

4.7.4. Salmon DNA Probe, 100 µM stock

4.8. Prepare the following per SOP PCR-3.2.

4.8.1. *B. dorei* DNA Standards (10^5, 10^4, 10^3, 10^2, 10^1)

4.8.2. *B. dorei* DNA spikes (10^5, 10^4)

4.8.3. *Bacteroides* primers 500 µM stock

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4.8.3.1. Forward Primer HF183F: 5'ATCATGAGTTCACATGTCCG

4.8.3.2. Reverse Primer BthetR1: 5'CGTAGGAGTTTGGACCGTGT

4.8.4. *Bacteroides* Probe 100 µM stock

4.8.4.1. Probe BthetP1: 5'-/6FAM/CTG AGA GGA /ZEN/ AGG TCC CCC ACA TTG GA/3IABkFQ/-3'

5. CALIBRATION AND STANDARDIZATION

5.1. See DEP Laboratory Quality Manual, Section 5.5.

5.2. Check thermometers annually. See SOP LB-035.

5.3. Check calibration of micropipettes quarterly for accuracy and precision. See SOP PCR-2.4.

5.4. Refer to SOP PCR-2.3 for use and maintenance of thermocycler.

5.5. Refer to SOP BB-036 for calibration and use of the balance.

5.6. Refer to SOP PCR-2.5 for calibration and maintenance of support equipment.

5.7. After each use of the UV workstation, wipe down all surfaces with 10% bleach, then rinse with PCR grade water. Turn on UV lamps for at least 15 minutes. The UV workstations have timers for the use of the UV lamps.

5.8. Clean work area counters after use. Spray surface of work area with 10% bleach solution and wipe down after 15 minutes. Spray and wipe down with sterile DI water.

6. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

6.1. See DEP Field SOP FS 2000 for sample collection and preservation.

6.2. See SOP PCR-5.0 for details on sample custody and printing sample labels.

7. SAMPLE PREPARATION

7.1. Refer to SOP PCR-4.0 for sample filtration.

7.2. Refer to SOP PCR-4.1 for sample extraction.

7.3. Refer to SOP PCR-4.2 for purification of inhibited or solid samples.

8. ASSAY PREPARATION

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8.1. Primer/Probe Mix – In PCR Workstation #1, prepare primer/probe mix for each assay (SKETA and HF183) separately as follows:

8.1.1. HF183 Assay: Add 4 µL of 500 µM forward primer stock, 7 µL of 500 µM reverse primer stock, and 6 µL of 100 µM probe to 583 µL of PCR grade water to make primer/probe mix. Use a blue 1.5 µL centrifuge tube labeled "PP" for the primer/probe mix. Mix by tapping then centrifuge briefly.

8.1.2. SKETA Assay: Add 10 µL of each 500 µM primer (forward and reverse) stock and 9 µL of 100 µM probe to 571 µL of PCR grade water to make primer/probe mix. Use a pink 1.5 µL centrifuge tube labeled "PP" for the primer/probe mix. Mix by tapping then centrifuge briefly.

8.1.3. Label tubes with the date. Store at 4°C and use within one week.

8.1.4. See Table 15.1 for a summary.

8.2. qPCR Master Mix assay – Create a reaction plate worksheet per SOP PCR-5.0. In PCR Workstation #1, prepare Master Mix for each assay (SKETA and HF183) separately as follows:

8.2.1. Count the number of wells needed for each assay to determine the total volume of reagents needed for the assay. Enter the number of wells in the reaction plate worksheet under the Master Mix Preparation section under '#Rxns' cell. The calculator adds an additional 3-4 wells in the calculations to ensure enough Master Mix is available (as some can be lost in mixing and pipetting).

8.2.2. For each assay add 2 µL X total wells of BSA, 10 µL X total wells of PerfeCta® qPCR ToughMix™ Rox™ for the StepOne Plus or TaqMan™ Environmental Master Mix 2.0 for the QuantStudio3, and 3 µL X total wells of primer/probe mix (PP). Use a pink 1.5 µL centrifuge tube labeled "MM" for the SKETA Master Mix. Use a blue 1.5 µL centrifuge tube labeled "MM" for the HF183 Master Mix.

8.2.3. See Table 15.2 for a summary.

9. REACTION PLATE PREPARATION

9.1. Collect a 96 well plate and holder. In PCR workstation #1, add 15 µL of appropriate qPCR Master Mix assay to each well of a reaction plate using the electronic Xplorer pipette with a 100 µL tip. Use Figure 15.1 as a guide. Add the HF183 Master Mix to the blue wells shown in Figure 15.1. Add the SKETA Master Mix to all the pink wells in Figure 15.1. Place the Master Mix, BSA, and any labeled extra PP mix back in Thermo Refrigerator B246A. Carefully move the plate to PCR workstation #2.

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9.1.1. Make sure to re-mix the Master Mix occasionally by tapping the tube, followed by a pulse spin in the minicentrifuge. This ensures that the Master Mix is evenly distributed into all the wells.

9.1.2. Master Mix is more viscous than water and should be pipetted slowly to ensure accuracy. Bubbles can easily form in the pipette tip if pipetted too quickly.

9.2. In PCR workstation #2, add 5 µL of DNA sample extract or control to each well using a 10 µL pipette. Use Figure 15.1 as a guide. To ensure proper mixing, tap mix then centrifuge briefly. Switch tips for each sample or control. If any liquid falls outside of the well or sprays, the plate should be discarded and hood thoroughly cleaned. Use the following order for addition of the template to wells:

9.2.1. Add the HF183 Standard Curve controls (see Section 12.1) to wells A1-E3.

9.2.2. Add the SKETA Standard Curve controls (see Section 12.2) to wells F1-H3

9.2.3. Add PCR grade water for the No Template Controls (NTC, SURNTC) and 3rd well spike control (LFB_BDSPK) to wells A6, B6, and C4-D6. See Section 12.3.1 and 12.4.3).

9.2.4. Add 1 µL of the *B.dorei* spike standard (LFB_BDSPK, and _SPK_BDSPK) to the spike control and 3rd well samples in wells A6-B6, C7-C12, and G5-G12. Use Figure 15.1 as a guide. See Section 12.4.3.

9.2.5. Add the method blank (MBLK) control (see Section 12.3.2) to wells A4-B4.

9.2.6. Add the extraction blank (EXBLK) control (see Section 12.3.3) to wells A5-B5.

9.2.7. Add the extraction control (LFB_BDLFB, SUR_BDLFB) to wells E4-H4. See Section 12.4.1.

9.2.8. Add the sample extracts to appropriate wells (4 wells per sample) using Figure 15.1 as a guide. Add the sample to the three HF183 wells first, change tip, and then add the sample to the final SKETA well.

9.3. Seal plate with optical adhesive. Vortex slowly, on Speed 1, for 30 seconds to mix.

9.4. Centrifuge plate at 2500 rpm for 2 minutes 30 seconds (Program 4 on Thermo Sorval T3 centrifuge). Check to make sure the reaction mix is at the bottom of each well. If not, centrifuge for another minute.

10. SAMPLE ANALYSIS

10.1. Place plate in thermocycler and analyze. Refer to SOP PCR-2.3 for setup and use of the thermocycler. Use the default thermocycler cycling conditions for a standard ramp speed (See Table 15.3).

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11. DATA ARCHIVAL

11.1. Refer to SOP PCR-2.3 for instrument file setup.

11.2. Refer to SOP PCR-5.0 for worksheet instructions.

11.3. Refer to SOP PCR-5.1 for data upload instructions.

12. QUALITY CONTROL

12.1. HF183 Assay Standard Curves

12.1.1. Analyze an HF183 Assay standard curve per plate. Use 5 serially diluted standards at 10^5, 10^4, 10^3, 10^2, and 10^1 concentrations in triplicate. These are added to A1-E3 wells on the plate. Use Figure 15.1 as a guide.

12.1.2. Standard curve efficiency should be between 90-115% and R^2 should be ≥ 0.99. If efficiency is < 70% or R^2 is < 0.97, samples should be re-analyzed. Notify supervisor.

12.2. SKETA Assay Standard Curves

12.2.1. Analyze a SKETA Assay standard curve per plate. Use 3 serially diluted standards at 10^3, 10^2, and 10^1 concentrations in triplicate. These are added to F1-H3 wells on the plate. Use Figure 15.1 as a guide

12.2.2. Standard curve efficiency should be between 80-120% and R^2 should be ≥ 0.99. Samples are not affected if the efficiency or R^2 are outside limits.

12.3. Negative Controls

12.3.1. No Template Control (NTC)

12.3.1.1. Include in triplicate on each plate for each target analyzed as a negative control. For each Master Mix used on a reaction plate, add 15 µL of Master Mix and 5 µL of sterile water to three wells.

12.3.1.2. The HF183 NTCs are labeled NTC_1, NTC_2, and NTC_3. The SKETA NTCs are labeled SURNTC_1, SURNTC_2, and SURNTC_3.

12.3.1.3. If any of the NTCs have a result of less than 35 C_q, the reaction plate may have contamination and should be re-run if samples are affected. Notify supervisor.

12.3.2. Method Blank (MB) – labeled MBLK_#

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12.3.2.1. Analyze a method blank for each batch of samples filtered (maximum of 13 samples per batch). Filter 50 mL of sterile PBS at the beginning of each sample batch. Extract and analyze with the same batch of samples. Add 5 µL of the blank extract to duplicate wells containing 15 µL HF183 Master Mix.

12.3.2.2. If the MB has a result above the MDL, there may be possible sample contamination. All sample results in the batch that are less than ten times the amount of contamination should be "V" qualified and the following comment should be added: "Result is V-qualified due to a control blank failure." For samples that are greater than 10X the blank, add the following comment: "Control blank failure. Result is >=10 times blank result."

12.3.3. Extraction Blank (EB) – labeled EXBLK_#

12.3.3.1. Analyze an extraction blank for each batch of samples extracted (maximum of 13 per batch). Place a sterile filter in an extraction tube labeled EB. Add 600 µL of salmon buffer and extract along with the samples. Add 5 µL of the blank extract to duplicate wells containing 15 µL HF183 master mix.

12.3.3.2. If the EB has a result less than 35 C_q, there may be possible contamination at the extraction step. Samples may be affected and should be re-analyzed. Notify supervisor.

12.4. Positive Controls

12.4.1. Extraction Control (EC)

12.4.1.1. Analyze an extraction control for each batch of samples extracted (maximum of 13 per batch). Place a sterile filter in an extraction tube labeled EC. Pipette 5 µL of 10^5 *B.dorei* standard into the filter, and add 595 µL of salmon buffer.

12.4.1.2. On the plate, the HF183 assay EC is labeled, LFB_BDLFB#. The SKETA assay EC is labeled, SUR_BDLFB#.

12.4.1.3. The extraction control helps determine extraction recovery. Percent recovery of extraction control for both the HF183 and SKETA assay should be 75-150%. If recovery is low, extraction efficiency may be low.

12.4.2. Sample Processing Control (SPC) (SKETA Assay)

12.4.2.1. The SKETA assay serves as a sample processing and inhibition control. Salmon DNA is added to the extraction buffer and each sample is

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analyzed for both the HF183 and SKETA assays. The SKETA assay measures the amount of recovery of Salmon DNA and therefore the efficiency of the extraction process. Low recovery indicates an inefficient extraction process or the presence of inhibition. This control is reported as a surrogate.

12.4.2.2. Analyze each sample, replicate, and extraction control with the SKETA assay alongside the HF183 Assay. See Figure 15.1 for plate layout details.

12.4.2.3. Compare extraction control Salmon DNA recovery to the Salmon DNA recovery in each of the samples. If sample recovery is < 50% of the extraction control recovery, then the sample may be inhibited.

12.4.3. Inhibition control (3rd well spike)

12.4.3.1. A known amount of *B.dorei* DNA is added to each sample as a matrix spike to check for inhibition. For each sample and replicate, spike the 3rd well of the HF183 assay with 1 µL of E4 B.*dorei* spike standard. Also add 1 µL of the standard to the spike control wells.

12.4.3.2. Compare spike control recovery to sample matrix recovery. If sample amplification is <50% of the control recovery, the sample matrix may have inhibitory effects.

12.4.4. If both the SPC and 3rd well spike show inhibition, re-run the sample from a diluted or purified extract. The purification kit may remove inhibitors.

12.4.4.1. If only one of the two controls show possible inhibition, the sample may need to be purified or diluted and re-analyzed. Notify supervisor.

12.5. Replicate Sample

12.5.1. Analyze at least one replicate per filtration batch (maximum of 13 per batch). Filter and analyze replicate in the same manner as original sample and within the same batch. Replicate samples are numbered SAMPLE ID_DUP_A and SAMPLE ID_DUP_B and 100 mL is filtered for each.

12.5.2. The target replicate relative percent different (RPD) is ≤ 25%. If RPD > 25%, the duplicate should be “J” qualified. A comment regarding precision is automatically added to the report.

12.5.3. If a replicate sample is not run, all samples in the batch should have the comment “Precision data unavailable for Analysis Batch#_____” added to the results.

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12.6. Refer to SOP PCR-5.1 for further details on uploading quality control data into LIMS, including calculations and control limits.

13. SAFETY/HAZARDOUS WASTE MANAGEMENT

13.1. All plates must be autoclaved for 45 minutes before disposal.

13.2. Refer to the Health and Safety Plan. Read MSDS information for reagent handling and disposal.

14. REFERENCES

14.1. EPA 815-B-04-001, Quality Assurance/Quality Control Guidance for Laboratories Performing PCR Analyses on Environmental Samples (October 2004)

14.2. PerfeCta® qPCR ToughMix™, Rox™ Protocol

14.3. DEP Laboratory Quality Manual DEP Laboratory Quality Assurance Manual

14.4. Health and Safety Plan Health and Safety Plan

14.5. DEP SOP FS 2000, *General Water Sampling*

14.6. DEP SOP LB-035, *Calibration of Lab Thermometers using a Constant Temperature Calibration Bath*

14.7. DEP SOP BB-36, *Calibration, Use and Maintenance of Mettler Toledo XA105DU Balance*

14.8. DEP SOP PCR-2.3, *Calibration, Use, and Maintenance of StepOne Plus Thermocycler*

14.9. DEP SOP PCR-2.4, *Calibration and Use of Mechanical Pipettes using PCS® Pipette Calibration System and Artel® Pipette Tracker™*

14.10. DEP SOP PCR-2.5, *Calibration, Use, and Maintenance of Support Equipment*

14.11. DEP SOP PCR-3.0, *Preparation of Reagents for quantitative Polymerase Chain Reaction (qPCR)*

14.12. DEP SOP PCR-3.1, *Preparation of SKETA assay standards and reagents for qPCR analysis*

14.13. DEP SOP PCR-3.2, *Preparation of Bacteroides dorei (B. dorei) Assay Reagents and Standards for qPCR analysis*

14.14. DEP SOP PCR-4.0, *Preparation of samples for qPCR*

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14.15. DEP SOP PCR-4.1, *Crude Lysate Extraction Method for qPCR samples*

14.16. DEP SOP PCR-4.2, *Purification of Filtered Samples using the Qiagen DNeasy PowerLyzer PowerSoil DNA Isolation Pro Kit*

14.17. DEP SOP PCR-5.0, *Sample Custody, Preparation Labels, and Worksheet Instructions for Molecular Biology*

14.18. DEP SOP PCR-5.1, *Data upload, Verification, and Sample Authorization for Molecular Biology*

15. TABLES AND DIAGRAMS

Table 15.1 - Primer/Probe Mix Summary

	HF183 Assay	SKETA Assay
Forward Primer (500µM stock)	4 µL	10 µL
Reverse Primer (500µM stock)	7 µL	10 µL
Probe (100µM stock)	6 µL	9 µL
Sterile H_2O	583 µL	571 µL

Table 15.2 – Master Mix Summary

	HF183 Assay		SKETA Assay	
Reagent	Volume per reaction (µL)	Volume (µL) per full plate (68 + 4 extra wells)	Volume per reaction (µL)	Volume (µL) per full plate (28 + 3 extra wells)
BSA 2 mg/mL	2	144	2	62
ToughMix MM	10	720	10	310
Primer/Probe Mix	3	216	3	93
Total of 15 µL of Master Master Mix + 5 µL of DNA template = total volume of 20 µL per reaction				

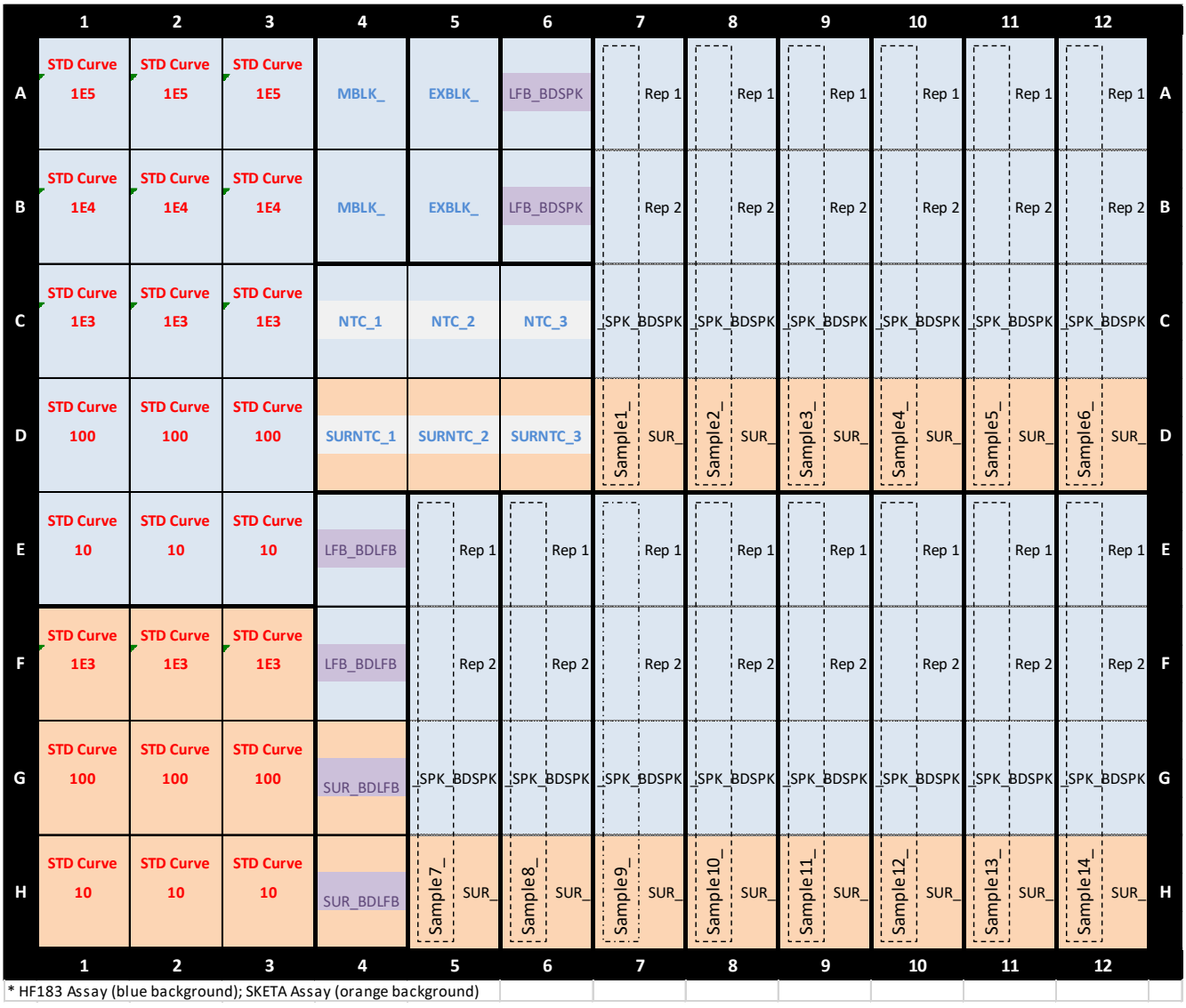
Table 15.3 – Cycling Conditions

		PCR
Step	Enzyme Activation	Cycle (40 cycles)

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	Hold	Denature	Anneal/Extend
Temperature (°C)	95	95	60
Time	10 min.	15 sec.	1 min

Figure 15.1 - Plate Layout Map



* HF183 Assay (blue background); SKETA Assay (orange background)

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Appendix of Changes

09/29/2016 Updated SOP references and master mix reagent.

08/16/2019 Updated new QuantStudio Thermocycler and DEP Laboratory Quality Manual

07/31/2020 Updated Autoclave serial number and DEP Laboratory Quality Manual

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