Pseudokirchneriella subcapitata[*] Culture for *Ceriodaphnia dubia* Food ([*]Formerly *Selenastrum capricornutum*)

1. SCOPE AND APPLICATION

    1.1. This method is based on EPA 821-R-02-012 Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms, 5th Edition. It can also be used for *Daphnia pulex* and *Daphnia magna* food.

2. SUMMARY OF METHOD

    2.1. Nutrients are added to sterilized-deionized water to prepare media in a flask. Algae is added to the media and allowed to grow for 3-5 days. The flask is moved to walk-in refrigerator for algae to settle. Supernatant is removed. Concentrated cells are counted. Cell concentration is adjusted by adding sterilized deionized water or further settling. Food is stored and used as needed.

    2.2. Interferences

        2.2.1. Unsterile glassware, media and reagents as well as airborne algae from other cultures create the potential for contamination of the *Pseudokirchneriella subcapitata* culture.

3. EQUIPMENT AND SUPPLIES

    3.1. Coulter® Counter Model Multisizer 4e (SN BA23044)

    3.2. Refrigerator, temperature range >0-≤6 ºC (temperature monitored per SOP TA-06.01)

    3.3. Erlenmeyer flask (quantity and volume can be adjusted to meet laboratory needs)

    3.4. 7-10-day old *Pseudokirchneriella subcapitata* stock culture, per SOP TA-04.08

    3.5. 2 mL Volumetric pipet for aeration

    3.6. 2, 5, or 10 mL sterile volumetric pipets for adding solutions

    3.7. Airline tubing, ~1-meter length

    3.8. Parafilm®

4. REAGENTS AND STANDARDS

    4.1. Autoclaved deionized (DI) Water

    4.2. PAAP stock solutions (listed in SOP TA-03.03)

    4.3. Alga-Gro®

    4.4. 2 µg/mL working solution for sodium selenate, prepared per SOP TA-09.08

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5. CULTURE PREPARATION

5.1. Obtain a clean, autoclaved Erlenmeyer flask. label the flask "*Pseudokirchneriella subcapitata* for *C. dubia* food" along with the batch number, expiration date, and the analyst's initials.

5.1.1. The batch number is the date that the ingredients are mixed and poured into the flask. Format is mm/dd/yy.

5.1.2. Expiration date is 3 months after the preparation date.

5.2. Fill the flask halfway with autoclaved DI water. Add 1 mL per L of each of the eight PAAP stock solutions to the flask using sterile pipets.

5.3. Add 1 mL of sodium selenate working solution per liter of solution in the flask.

5.4. Add 5 mL of Alga-Gro® per liter of solution in the flask.

5.5. Inoculate the flask with 100 mL (adjust as needed) of concentrated *P. subcapitata* cells from the algal stock culture.

5.5.1. Refer to SOP TA-04.08. One flask per liter of ~100 mL of 7-10-day old *P. subcapitata* can be substituted for the concentrated *P. subcapitata*.

5.6. Fill the flask to the neck with autoclaved DI water.

5.7. Place the flask on culture tray *A7*. This tray has continuous lighting (cool-white fluorescent bulbs) with an intensity of ~300-400 foot-candles. Any culture tray with continuous lighting of similar intensity (within 10%) can be used.

5.8. Connect the 2 mL pipet to the airline tubing. Place the pipet in the flask. Cover the flask with Parafilm®. Adjust the airflow to provide moderate aeration.

5.9. After 4 (±1) days (does not include set-up day), remove the aeration pipet from the flask. Seal the flask with Parafilm® and place the flask in the walk-in refrigerator. A visual inspection of the flask should show a significant increase in turbidity (should be green) from the initial inoculation.

5.9.1. Flask can be left for up to seven days prior to placing in the refrigerator, if visual observation suggests continued growth (greening) in the flask.

5.10. When the algae have settled, and a distinct clear layer has formed above a distinct green layer (usually within 2-3 days), siphon or pour off the clear supernatant.

5.11. Count culture density using the Coulter® Counter. Refer to SOP TA-06.41 for instructions on use of the counter. A density of 3.9 - 4.6 x 10^7 cells/mL is required to provide a sufficient concentration of cells in the *C. dubia* vials (containing 20 mL of media).

5.11.1. If the cell density is less than required, concentrate by allowing additional settling.

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5.11.2. If the cell density is greater than required, dilute as needed with autoclaved DI water.

5.12. Store between 0º and 6 ºC in the walk-in refrigerator. The batch can be stored for up to six months in the refrigerator; however, as a food source, it is preferable to use a batch within 90 days. Batches of algae less than 90 days old can be combined, especially if there is only a little of the "old" and a lot of the "new."

5.13. Record batch preparation and count information in the *Pseudokirchneriella subcapitata Food for* C. dubia *Log.*

6. DATA ARCHIVAL

6.1. Use a black, waterproof pen for all bench sheets and logbooks.

6.2. Record preparation in the *Pseudokirchneriella subcapitata Food for* C. dubia *Log*.

6.3. To correct raw data entries, place a single line through the incorrect or illegible entry. Write the corrected entry near the error, date and initial the correction. Do not overwrite entries.

6.3.1. The corrected entry is written as near as possible to the error or in the comment box on the appropriate bench sheet.

6.3.2. If the correction requires an explanation, write a comment and reference the error with a letter. Initial and date the explanation. Do not attempt to put all information (correction and comments) into a small space

6.4. Use sequential alphabetical annotation to identify errors and corresponding comments (use A for first error and use A for corresponding explanatory comment; use B for second error, etc.).

7. QUALITY CONTROL

7.1. Sterile, autoclaved glassware and sterile media and reagents are used to prevent contamination of the culture.

7.2. The culture is grown under ~400 foot-candles ±10% of continuous, cool-white lighting.

7.3. Culture can but should not be used for food beyond 90 days from the date of preparation.

8. SAFETY/HAZARDOUS WASTE MANAGEMENT

8.1. Proper laboratory protection, including lab hood or ventilation system, lab coat, gloves and safety glasses, is required when working with chemicals and unprocessed samples.

8.2. Refer to the Laboratory Quality Manual for sample disposal.

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8.3. Refer to the Laboratory Health and Safety Manual and Hazardous Waste Management.

8.4. Read MSDS information for reagent handling and disposal.

9. REFERENCES

9.1. EPA-821-R-02-012 *Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms, 5th Edition*, Test Method 2002.0.

9.2. DEP SOP TA-03.03, Preparation of Preliminary Algal Assay Procedure (PAPP) Medium for Freshwater Algae Cultures.

9.3. DEP SOP TA-04.08, Culture of Algal-Assay Inoculation Stocks.

9.4. DEP SOP TA-06.41, Calibration and Use of the Coulter® Counter Model Multisizer 4e.

9.5. DEP SOP TA-06.19, Temperature Monitoring of Walk-in Refrigerator, Freezer, and Toxicity Laboratory Incubators.

9.6. DEP SOP TA-09.08, Preparation of Selenium Solution for use in 20% DMW.

9.7. Laboratory Quality Manual https://floridadep.gov/dear/florida-dep-laboratory/content/dep-laboratory-quality-assurance-manual-and-sops

9.8. Laboratory Health and Safety Manual Health and Safety Manual

9.9. MSDS Information Sheets.

APPENDIX OF SIGNIFICANT CHANGES

12/12/2007 Added Scope an Application; added Summary of the Method.

06/24/2013 Reformatted SOP.

02/09/2015 Formatted for 508 compliance.

07/11/2016 Updated links.

09/13/2017 Added acceptable algae age range; corrected *Tox Record of Preparation* to Pseudokirchneriella subcapitata *Food for* C. dubia *Log*

08/22/2018 Updated links and changed font to Calibri.

07/02/2019 Updated references.

08/26/2020 Updated order and added sterile pipets to equipment list.

07/02/2021 Edited for clarity and universal language in data archival section. Updated Coulter Counter SOP.

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