Pimephales promelas and *Cyprinella leedsi* Chronic Toxicity Testing Methods

DEPARTMENT OF ENVIRONMENTAL PROTECTIONLABORATORY
BIOLOGY PROGRAM
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1. SCOPE AND APPLICATION

1.1. This method is based on EPA 821-R-02-013 *Short-Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, 4th Edition, Test Method 1000.0*. It is used to estimate the chronic toxicity of freshwater whole effluent and receiving water on *Pimephales promelas* (fathead minnow) and *Cyprinella leedsi* (bannerfin shiner) using < 24-hour old (or < 48 hours if shipped) organisms in a 7-day static-renewal exposure. Tests conducted by this procedure are BSY-CS-FFS and BSY-CD-FFS.

2. SUMMARY OF METHOD

2.1. Samples are received and prepared in the laboratory. Sample physical-chemical (phys-chem) parameters are measured, and samples are adjusted as needed to meet permit or method test requirements. Sample is filtered through a 60µm screen to remove debris and/or indigenous organisms. Sample dilutions and control waters are placed into test chambers. Test organisms are collected, fed and held for observation prior to loading into test chambers. Tests are incubated in laboratory incubators during the duration of the test. Physl-chem parameters are measured and recorded daily. Survival/mortality is recorded daily. The test endpoints are survival and growth (measured as dry weight) (IC25) of the organisms. Statistical endpoints are obtained from the EPA WET Statistics Excel spreadsheet per SOP TA-07.17. Results are entered in the Laboratory Information Management System (LIMS).

2.2. Test acceptability criteria: 80% or greater survival in the controls and average dry weight per surviving control organism equal to or greater than 0.25 mg.

2.3. Currently, *C. leedsi* has not been approved by EPA for NPDES compliance testing. The certification request (submitted ~1997) was inactivated in 2011 at the request of the Biology Program.

2.4. Interferences: Refer to EPA-821-R-02-013 Section 11. Includes toxic substances introduced by contaminants in dilution water or on hands, glassware, sample hardware, and testing equipment; dissolved oxygen < 4.0 mg/L; pH < 6.0 SU or > 9.0 SU; salinity > 1 part per thousand; ammonia; chlorine; debris; and indigenous organisms that may attack or be confused with the test organisms.

3. EQUIPMENT AND SUPPLIES

3.1. *P. promelas* or *C. leedsi* test organisms (< 48 hours old).

3.1.1. Test organisms are fed live *Artemia* nauplii at least 2 hours prior to test initiation. The FDEP Laboratory currently purchases test organisms. Refer to SOP TA-04.20.

3.2. 1 L Test chambers, autoclaved, 4 per test concentration.

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Note: Test chambers can be dried in a drying oven for eight hours at 90 °C rather than being autoclaved.

3.2.1. A test with 5 test concentrations and a control, using 4 replicates per treatment would require 24 chambers.

3.2.2. A minimum of four replicates per concentration is required.

3.2.3. The minimum test chamber size is 500 mL (recommended minimum) and the minimum volume per replicate is 250 mL (recommended minimum). A loading of 10 organisms per replicate is recommended and a minimum of 40 organisms per concentration is required.

3.2.4. We generally use 1L test chambers and 500 mL of sample. Adjust chamber size and replicate volume as necessary to accommodate available sample.

3.3. 10 - 1000 mL Volumetric flasks (as needed)

3.3.1. One set for toxicant/sample and one for diluent

3.4. 100 mL, 500 mL, 1000 mL and 2000 mL, Graduated cylinders (as needed).

3.4.1. For measuring amount of toxicant/sample or diluent needed for each concentration. One set for toxicant/sample and one for diluent.

3.5. 1 L, 2 L, 4 L and 6 L Erlenmeyer flasks (as needed).

3.5.1. For mixing test dilutions.

3.6. 1mL and 10 mL Serological pipettes, graduated.

3.7. Pipettor for volumetric pipettes.

3.8. Magnetic stir bars and stirrer.

3.8.1. For mixing test dilutions.

3.9. Carolina dish, 8" diameter.

3.9.1. Transfer fish from holding container to Carolina dish for easier loading of bioassay chambers.

3.10. Droppers and glass tubing with fire polished edges, 4 mm ID, for transferring organisms.

3.10.1. Nets or Nytex® screen cannot be used to transfer organisms as the screen can injure them.

3.11. Meters to measure phys-chem parameters:

3.11.1. pH (Calibrate per SOP TA-06.43).

3.11.2. Dissolved Oxygen (DO) (Calibrate per SOP TA-06.39).

3.11.3. Temperature (Digital thermometer; Calibrate per SOP TA-06.13).

3.11.4. Conductivity/salinity (Calibrate per SOP TA-06.36).

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3.12. Hach® kits to measure:

3.12.1. Total Residual Chlorine (TRC) (Measure per SOP TA-06.22).

3.12.2. Alkalinity (Measure per SOP TA-06.08).

3.12.3. Hardness (Measure per SOP TA-06.08).

3.12.4. Ammonia (Measure per SOP TA-06.35 or SOP TA-06.33).

3.13. Incubator should maintain selected test temperature (25 ± 1°C recommended) and a photoperiod of 16-hours light to 8-hours dark. Light intensity should be 50-100 ft candles. Test temperature must not deviate (maximum minus minimum) more than 3°C in the test chambers for the duration of the test.

3.14. Other incubators or the water bath can be used to regulate test temperature.

3.15. 30 mL disposable plastic cups, with parameter tray.

3.15.1. For aliquots of concentrations for parameters.

3.16. Fine mesh fish net.

3.16.1. To aid in euthanizing fish at test termination.

3.17. Ice bath.

3.17.1. Use 8'' Carolina dish with ice and cold water. For euthanizing fish during test termination.

3.18. Waste bucket.

3.19. Live *Artemia* nauplii (prepare per SOP TA-01.03).

3.20. Plastic transfer pipette.

3.20.1. Used for feeding.

3.21. Drying oven.

3.21.1. Capable of sustaining 100° C for 8 hours or 60° C for 24 hours. Refer to SOP TA-06.21.

3.22. Computer with access to LIMS and the EPA WET Excel spreadsheet for statistical analysis.

3.23. *Chronic Bioassay Bench Sheet* logbook.

4. REAGENTS AND STANDARDS

4.1. Dilution water - MHW (prepared per SOP TA-03.15).

4.2. Sample to be tested.

5. SAMPLE COLLECTION, PRESERVATION, AND HANDLING

5.1. See SOP FS 1000 for sample collection and preservation.

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5.1.1. For a valid test: a) the sample should be held at 0-6°C prior to test initiation, and b) sample holding time should not exceed 36 hours from time of collection (time of last sample collect for grab samples or the end of the 24-hour period for composite samples) to test initiation.

5.1.2. A series of three samples (designated as samples 1, 2, and 3) are collected and used upon receipt. Sample 1 is used for 2 days, sample 2 is used for 2 days, and sample 3 is used for 3 days. These samples typically arrive on Tuesday, Thursday and Saturday in 4L amber jugs or other Receiving approved container.

5.1.3. The remainder of each sample is stored in the Biology Walk-in cooler for solution renewal the following day(s).

5.1.4. See SOP TA-06.14 for bioassay sample log-in procedure.

6. TEST INITIATION

6.1. Place clean test chambers onto laboratory cart. Label each chamber to identify the replicate (A–D), test concentration, and facility sampled.

6.1.1. Place tape across front of chamber, beneath jar ID, and use waterproof marker to identify. Four chambers for each set of control replicates needed for each sample tested, plus four chambers per concentration for each sample tested.

6.2. Calibrate meters and record calibration readings.

6.2.1. Calibrate meters per respective SOPs. Record calibration readings in LIMS (report results in accordance with SOP TA-06.42).

6.3. Fill a 30 mL disposable plastic cup with sample and take initial pH measurement per SOP TA-06.43. Record value in the *Bioassay Sample log-in* logbook and LIMS TOX_pH logbook.

6.3.1. If the pH is outside the acceptable range of 6.0–9.0 SU, then an additional test with a pH-adjusted aliquot will be run. Adjust pH of sample per SOP TA-08.06. Record pH adjustment and/or measurement in the appropriate bench sheet. The test with the non-adjusted sample is then analyzed as a screening test, regardless of whether the pH adjusted aliquot is used for a screening or definitive test.

6.4. In the same plastic cup containing the sample, take initial DO measurement per SOP TA-06.39. Record value in *Bioassay Sample Log-in* logbook and LIMS TOX_DO logbook.

6.4.1. If DO is < 4.0 mg/L or $\geq$ 9.0 mg/L, aerate the entire sample to near saturation prior to test initiation. Record aeration (initial and final DO as well as aeration rate and duration) in the appropriate bench sheet. Samples with a potential DO problem generally show a downward trend in DO during the

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first few hours of the test. Monitor the DO closely; if the DO begins to fall, set up gentle aeration (< 100 bubbles/min) to ALL test replicates for duration of test using micropipettes and the aeration manifolds. No action is required if DO increases after test initiation.

6.5. In the same plastic cup containing the sample, measure the conductivity and temperature of the sample. Check the conversion table to determine salinity or read salinity directly from the Fisherbrand™ Traceable™ Bench Conductivity meter.

6.5.1. If conductivity is above 2,000 µmhos/cm or the salinity is more than 1 ppt, then freshwater organisms are probably not the appropriate test organisms for this effluent sample. Check with FDEP District personnel and also check the Facility's NPDES permit for bioassay requirements.

6.6. Measure total residual chlorine (TRC) in the sample per SOP TA-06.22. Record the result in the LIMS TOX_CHLORINE logbook and on the *Chronic Bioassay Bench Sheet*. Check the field data sheet supplied by sampler for field TRC and atypical field measurements.

6.6.1. If TRC > 0.1 mg/L and non-chlorine toxicity is to be measured, dechlorinate a subsample (volume needed to initiate the test) of the sample using sodium thiosulfate per SOP TA-09.18. Setup a concurrent screening test using the 100% sample that has not been dechlorinated and a control. Record chlorine result, amount of sodium thiosulfate used and final chlorine result after dechlorination in the logbook and in the designated section in LIMS.

6.7. Measure the desired volumes of diluent and sample into a 6 L Erlenmeyer flask using two graduated cylinders. Add the final amount of each solution with a 10 mL pipette when nearing the desired volume, to prevent over pouring. Place flask on stir table and stir until homogenized. Measure 2000 mL of the mixture back into a 2000 mL cylinder. Transfer 500 mL from the 2000 mL cylinder to a 500 mL cylinder. Empty the 500 mL cylinder into test chambers A through D for that concentration. Repeat for each test concentration.

6.7.1. For each sample tested, use four chambers per concentration in a dilution series chosen based on previous knowledge of toxicity level. There are typically 5 concentrations assigned by the permit and a control water.

6.7.2. Example of sample volumes for four 500 mL replicates of each concentration (0.5 dilution) of a chronic toxicity test:

Concentration	Diluent	Sample	Total Volume
Control	2000 mL	0 mL	2000 mL
6.25%	2000 mL	2000 mL of 12.5%	4000 mL
12.50%	2000 mL	2000 mL of 25%	4000 mL
25%	2000 mL	2000 mL of 50%	4000 mL
50%	2000 mL	2000 mL of 100%	4000 mL
l00%	0 mL	2000 mL of 100%	2000 mL

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6.8. Perform QA standard reference toxicant (SRT) tests monthly (or concurrent with bioassay if bioassays performed less frequently; a concurrent SRT with one bioassay can be used as the SRT for a series of tests as long as the time interval does not exceed 30 days prior to the start of a bioassay test), using potassium chloride toxicant (other toxicants can be used) for the SRT.

Example of sample volumes for a chronic SRT test with potassium chloride as the toxicant:

Concentration	Diluent	Sample	Total Volume
Control	2000 mL	0 mL	2000 mL
0.0625 g/L	2000 mL	2000 mL of 0.125 g/L	4000 mL
0.125 g/L	2000 mL	2000 mL of 0.25 g/L	4000 mL
0.25 g/L	2000 mL	2000 mL of 0.5 g/L	4000 mL
0.5 g/L	2000 mL	2000 mL of 1 g/L	4000 mL
1 g/L	0 mL	2000 mL of 1 g/L	2000 mL

6.9. Pour a subsample from all A replicates into the disposable plastic cups in the parameter tray. Measure pH, DO, temperature (can be measured from test chamber), and conductivity of the solutions from the subsamples. Record all readings. Record any comments pertaining to the condition or appearance of the sample. On the following seven days, rotate through all of the replicates and take parameters before and after renewing the sample.

6.9.1. Conductivity readings can be rounded to the nearest 5 µmhos or 0.1 mmhos. Readings for pH, DO, and temperature are read to one decimal place. Take measurements from lowest to highest toxicant concentration.

6.9.2. Record all readings in the appropriate columns on the *Chronic Bioassay Bench Sheet*.

6.10. Using a wide bore pipette, load **two** (20% of test organisms per replicate) fish into each test chamber. Repeat the loading process until **ten** fish have been loaded into each replicate. Verify count over a light table. Record counts in appropriate columns on the *Chronic Bioassay Bench Sheet*.

6.10.1. (10 fish per replicate) x (4 replicates per test concentration) = 40 fish per concentration tested

6.10.2. A person other than the loader should verify the number of fish in each replicate.

6.10.3. Record counts and phys-chem readings in appropriate columns in the *Chronic Bioassay Bench Sheet.*

6.11. Use a plastic transfer pipette to feed fish 0.2 mL *Artemia* nauplii concentrate (800-1100 *Artemia*) from concentrate once in the morning 2 hours prior to renewals, and again after renewals.

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6.11.1. Feed once on Day 1 and then twice on succeeding days. *Artemia* nauplii concentrate is prepared based on SOP TA-01.03.

6.12. Place chambers (use randomization template determined by a dice roll and maintain initial pattern for test duration) in incubator and cover with clear Plexiglass. Covers prevent accidental contamination and minimize evaporation.

6.13. Measure alkalinity, hardness, and total ammonia of sample and control water.

6.13.1. Measure per SOP TA-06.08, TA-06.35, or TA-06.33, respectively.

7. INTERIM READINGS

7.1. Every 24 hours pour subsamples of the appropriate replicates into disposable plastic cups. Measure pH, temperature (can be taken from test chamber), DO, and conductivity in the subsamples before renewing the test solutions and record readings. Count and record the number of fish in each replicate. Starting with the controls and then going in order of increasing toxicant concentration, clean out all dead *Artemia* nauplii at the bottom of each jar. Carefully siphon most, but not all, of the solution in each chamber through a fine mesh net attached to a plastic hose into the waste bucket. Try to keep the fish within the chamber. Replace the contents of each chamber with respective fresh concentrations of the sample.

7.1.1. All counting is done over a light table. Fish are recorded as live if they move when lightly prodded. Record counts and phys-chem readings in appropriate columns in the *Chronic Bioassay Bench Sheet.*

7.1.2. The remaining water after siphoning should measure a half-inch to an inch from the bottom of the chamber. Check the excess mesh net below the end of the hose for trapped fish before siphoning the next jar. If a fish is stuck, gently move the mesh around the water to release the fish.

7.2. Pour subsamples from the appropriate replicate after renewal. Measure pH, temperature (can be measure in the test chamber), DO, and conductivity after renewing the test solutions and record readings.

7.2.1. Subsamples are poured to minimize disturbance to the fish.

7.3. Feed fish 0.2 mL *Artemia* nauplii concentrate once in the morning two hours prior to renewals, and again after the renewals. Cover chambers when done and return to the incubator.

8. TEST TERMINATION

8.1. On day 7 (t = 168 hours ± 6 hours), count the number of surviving fish. Remove and discard any dead fish. Pour subsamples and measure parameters of the solutions. Record all counts and readings. Record any comments pertaining to the appearance or condition of the fish. Record test termination date and time.

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8.1.1. Counting is done over a light table. Phys-chem readings are taken from the subsample to minimize disturbance to the fish.

8.1.2. Record counts, readings, date, and time in appropriate columns in *Chronic Bioassay Bench Sheet.*

8.2. Make weigh boats two days before test termination per SOP TA-06.21.

8.3. Pour the contents of Control replicate A onto a fine mesh net held over the waste bucket. Immediately euthanize the fish by immersing the fish in ice water in the Carolina dish. Keep the fish immersed for two minutes after the last movement is noted in the fish.

8.4. When the fish are dead, take a pair of forceps and place each fish into the respective weigh boat.

8.5. Repeat steps 7.3-7.4 for all test concentrations and replicates.

8.6. The day after test termination, dry the weigh boats containing the fish for eight hours in the sediment drying oven. Store the Carolina dish in the desiccator in a vacuum before and after they are dried.

8.6.1. Refer to SOP TA-06.21.

8.6.2. Dry for eight hours (minimum of six hours) at 100°C.

8.6.3. Fish can also be dried for 24 hours at 60°C.

8.7. Remove the dish after cooling and weigh the weigh-boats. Record the weights in the *Chronic Bioassay Bench Sheet*.

8.7.1. Refer to SOP TA-06.21.

9. STATISTICAL ANALYSIS

9.1. Complete the weight page in the *Chronic Bioassay Bench Sheet* logbook.

9.2. Enter test results into statistical program to obtain NOEC, LOEC and IC25.

9.2.1. Refer to SOP TA-07.17.

9.3. Enter test results into LIMS.

9.3.1. Data is verified by a second person prior to being uploaded into LIMS.

10. DATA ARCHIVAL

10.1. Use a black, waterproof pen for all bench sheets and logbooks.

10.2. *Bioassay Sample Log-In* logbook. Record per SOP TA-06.14.

10.3. Record information in appropriate logbooks (*Chronic Bioassay Bench Sheet*).

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10.4. To correct raw data entries, place a single line through the incorrect or illegible entry, write the corrected entry near the error, date and initial the correction. Do not overwrite entries.

10.4.1. The corrected entry is written as near as possible to the error or in the comment box on the appropriate bench sheet.

10.4.2. If the correction requires an explanation, write a comment and reference the error with a letter. Initial and date the explanation. Do not attempt to put all information (correction and comments) into a small space.

10.4.3. Use sequential alphabetical annotation to identify errors and corresponding comments (use A for first error and use A for corresponding explanatory comment; use B for second error, etc.).

10.5. DEP Laboratory Information Management System (LIMS)

10.6. Log sample receipt, temperature, and condition into *Chronic Bioassay Bench Sheet* logbook. Fill in information requested at top of bench sheets as completely as possible prior to test initiation. Include full name of facility, address, contact person, date and time collected, field TRC, LIMS sample and job numbers, test chamber ID, test chamber size, test solution volume, food type, and food batch numbers. Access scanned Field Sheets via \\floridadep\data\dear_scans\Labs\BizhubScans\Receiving to obtain sampling information.

10.7. Enter the NOEC, LOEC, and IC25 results from the EPA WET Excel spreadsheet into the appropriate logbook, appropriate computer file and LIMS results editor. A second person must verify data prior to uploading into LIMS.

11. QUALITY CONTROL

11.1. Refer to DEP Laboratory Quality Manual for corrective action and handling out-of-control data. Refer to EPA 821-R-02-013 Test Method 1002.0 for test conditions and acceptability criteria.

Perform QA standard reference toxicant (SRT) tests monthly or concurrently.

11.2. If a problem occurs during sample processing or testing, the following steps are performed: 1) lab supervisor is notified, 2) client is notified (client makes decision for additional testing), 3) section administrator is notified, and 4) a nonconformance report (NCR) is submitted describing the problem and the corrective action.

11.3. To create NCR: open LIMS and go to Laboratory/Administrative/Non-Conformance Report. Refer to https://floridadep.gov/dear/quality-assurance/content/dep-sops for sample collection and preservation, and the Laboratory Quality Manual for corrective action and handling out-of-control data.

11.4. Samples should be held at 0-6 degrees C prior to test initiation and between renewals (exception—hand delivered sample day of collection should be cooling

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relative to collection temperature). Sample holding time should not exceed 36 hours from time of collection until initial use of sample.

11.5. Incubator light intensity maintained at ~50-100 ft-candles for a photoperiod of 16 hours light to 8 hours dark.

11.6. Incubator is maintained at the selected test temperature of 25 ± 1°C.

11.7. Test temperature must not deviate more than 3°C in the test chambers for the duration of the test.

11.7.1. If the test vessel temperatures for a facility test exceeds ±3˚C, but results indicate no toxicity, add the comment, "Temperature of test solutions deviated from test specifications." to the reported test result.

11.7.2. If the incubator checkpoint temperatures exceed ±3˚C, but results indicate no toxicity, add the comment, "Continuous temperature monitoring in surrogate test vessels deviated from test specifications." to the reported test result.

11.7.3. If room temp checkpoints are out, but there is no toxicity and the incubator and test vessel temps were fine, do not qualify or add a comment.

11.7.4. If results do indicate toxicity in a sample, the acceptability of the test will depend on the experience and professional judgment of the laboratory analysts and the reviewing staff of the regulatory authority. If the temperature deviation from test specifications is suspected of contributing to the toxicity of a test, J-qualify the result and add the comment "Estimated value. Deviation of test solution temperatures from test specifications suspected to contribute to toxicity."

12. SAFETY/HAZARDOUS WASTE MANAGEMENT

12.1. Proper laboratory protection, including lab hood or ventilation system, lab coat, gloves, and safety glasses, is required when working with chemicals and unprocessed samples.

12.2. Refer to DEP Laboratory Quality Manual for sample disposal.

12.3. Refer to the DEP Laboratory Health and Safety Plan.

12.4. Read MSDS information for reagent handling and disposal.

13. REFERENCES

13.1. DEP Laboratory Quality Manual

13.2. DEP Laboratory Health and Safety Plan

13.3. EPA 821-R-02-013 *Short-Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, 4th Edition, Test Method 1000.0*

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13.4. MSDS

13.5. DEP SOP FS 1000, *General Sampling*

13.6. DEP SOP BG-01, *Data-Entry Verification*

13.7. DEP SOP TA-01.03, *Artemia Nauplii Culture*

13.8. DEP SOP TA-02.00, *Feeding Test-Organism Cultures*

13.9. DEP SOP TA-03.15, *Preparation of Synthetic Moderately Hard Freshwater*

13.10. DEP SOP TA-04.20, *Receiving and Maintaining Purchased Test Organisms*

13.11. DEP SOP TA-06.08, *Measuring Total Alkalinity and Hardness*

13.12. DEP SOP TA-06.13, *Thermometer Accuracy Check and Use*

13.13. DEP SOP TA-06.14, *Bioassay Sample Log-In Procedure*

13.14. DEP SOP TA-06.21, *Weighing Chronic Tests*

13.15. DEP SOP TA-06.22, *Measuring Total Residual Chlorine Using the Hach® Pocket Colorimeter™ II Test Kit*

13.16. DEP SOP TA-06.33, *Measuring Ammonia using HACH® Ammonia Nitrogen Test Kits for Freshwater and Saltwater Samples*

13.17. DEP SOP TA-06.35, *Measuring Total Ammonia Using the Hach® Ammonia Nitrogen Pocket Colorimeter™ II Test Kit*

13.18. DEP SOP TA-06.36, *Calibration and Use of the Fisherbrand™ Traceable™ Bench Conductivity Meter for Conductivity and Salinity*

13.19. DEP SOP TA-06.39, *Thermo Scientific Orion Star A213 Benchtop RDO/DO Meter Calibration and DO Measurement*

13.20. DEP SOP TA-06.42, *LIMS Entry for Laboratory Parameters*

13.21. DEP SOP TA-06.43, *Calibration and Use of Fisherbrand™ Accumet™ AB315 pH/mV Meter*

13.22. DEP SOP TA-07.17, *Using EPA Whole Effluent Toxicity (WET) Statistics for Acute and Chronic Bioassay Tests*

13.23. DEP SOP TA-08.06, *pH adjustment of Samples: Algal Assay and Toxicity Bioassay*

13.24. DEP SOP TA-09.01, *Potassium Chloride Reference Toxicant Stock Solutions*

13.25. DEP SOP TA-09.18, *Dechlorination Procedure using Sodium Thiosulfate*

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APPENDIX OF CHANGES

07/25/2008 Moved *Scope and Application*, *Summary of the Method* and *Interferences* from comments to body of SOP. Added *References* and *Appendix of Changes*. Removed SOP TA-06.06 from Materials # 11 comment. Deleted *Obtain young per SOP #TA05_03* from Materials # 1 comment.

07/13/2009 Updated Quality Manual reference.

08/14/2009 Replaced reference to SOP TA-07.08 with reference to SOP TA-07.16

07/21/2010 Updated reference to Biology Quality Manual. Removed reference to vinyl gloves in the Safety section comment and added a Caution statement.

03/17/2011 Updated references to Biology Quality Manual, now Bureau Quality Manual. Changed 105ºC to 100ºC throughout the SOP.

02/27/2012 Updated reference to Bureau Quality Manual. Added correction of illegible entries to the Documentation Section.

12/20/2013 Reformatted. Deleted step 3 “Count the fish in each replicate. Remove and discard any dead fish. Record number in the *Chronic Bioassay Bench Sheets*” under TEST TERMINATION and added “Repeat steps 6.3-6.4 for all test concentrations and replicates” after step 4.

02/12/2015 Formatted for 508 compliances.

01/13/2016 Updated links.

03/09/2016 Updated SOPs and References.

03/10/2017 Updated SOPs and References.

02/26/2018 Updated links, SOPs and References.

02/22/2019 Updated link and references to meter SOPs

01/06/2020 Updated links, SOPs and added a computer to Equipment and Supplies section.

02/01/2021 Updated SOP for universal language and grammar accuracy.

01/31/2022 Updated SOP for grammar accuracy and added a table of contents.

02/02/2023 Updated SOPs for the pH meter and added an SOP for LIMS entries.

01/18/2024 Added missing References; changed SOP TA-06.37 to new SOP TA-06.43; updated links; edited for grammar, clarity, and continuity.

03/12/2024 Added information about what to do if temperatures exceed the ±3˚C limit.

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